pequeños ANIMALES

evolutior

LU

 Immunotherapeutic Strategies for Canine Lymphoma: Changing the Odds Against Non-Hodgkin Lymphoma Estrategias inmunoterapéuticas para el linfoma canino: cambiar las probabilidades contra el linfoma no Hodgkin

N° 4

- Animal models of cancer metastasis to the bone Modelos animales de metástasis del cáncer al hueso
- Animal models in osteosarcoma Modelos animales en osteosarcoma



N° 4

- Animal models in osteosarcoma......50
 Modelos animales en osteosarcoma





Patrocinado por:





EDITA: IMPULSO VET impulsovet@impulsovet.es Immunotherapeutic Strategies for Canine Lymphoma: Changing the Odds Against Non-Hodgkin Lymphoma

Estrategias inmunoterapéuticas para el linfoma canino: cambiar las probabilidades contra el linfoma no Hodgkin

https://www.frontiersin.org/articles/10.3389/fvets.2021.621758/full

Joana N. R. Dias, Ana S. André, Sandra I. Aguiar, Solange Gil, Luís Tavares and Frederico Aires-da-Silva*

1. Centro de Investigação Interdisciplinar em Sanidade Animal, Faculdade de Medicina Veterinária, Universidade de Lisboa, Avenida da Universidade Técnica, Lisbon, Portugal

Palabras clave:

cáncer, oncología comparativa, linfoma no Hodgkin, linfoma canino, inmunoterapia contra el cáncer

Keywords:

cancer, comparative oncology, non-Hodgkin lymphoma, canine lymphoma, cancer immunotherapy



Tabla de contenido:

- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current
- Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



3 de 72

a nueva era de la inmuno-oncología ha traído complejidades y desafíos que enfatizan la necesidad de identificar nuevas estrategias y modelos para de-

sarrollar terapias exitosas y rentables. La inclusión de un modelo canino en el desarrollo farmacológico de inmunoterapias contra el cáncer está siendo ampliamente reconocida como una solución válida para superar varios obstáculos asociados con los modelos preclínicos convencionales. Impulsado por el éxito de las inmunoterapias en el tratamiento del linfoma no Hodgkin humano (LNH) y por las notables similitudes del LNH canino con su contraparte humana, el LNH canino ha sido uno de los principales focos de la investigación comparativa. En la presente revisión, resumimos una visión general de los desafíos y perspectivas de las inmunoterapias actuales contra el cáncer y el papel que la medicina comparativa podría desempeñar en la solución de las limitaciones que plantea este campo en rápida expansión.

he new era of immune-oncology has brought complexities and challenges that emphasize the need to identify new strategies and models to develop successful and cost-effective therapies. The inclusion of a canine model in the drug development of cancer immunotherapies is being widely recognized as a valid solution to overcome several hurdles associated with conventional preclinical models. Driven by the success of immunotherapies in the treatment of human non-Hodgkin lymphoma (NHL) and by the remarkable similarities of canine NHL to its human counterpart, canine NHL has been one of the main focus of comparative research. Under the present review, we summarize a general overview of the challenges and prospects of today's cancer immunotherapies and the role that comparative medicine might play in solving the limitations brought by this rapidly expanding field. The state of art of both human and canine NHL and the

rationale behind the use of the canine model to bridge the translational gap between murine preclinical studies and human clinical trials are addressed. Finally, a review of currently available immunotherapies for canine NHL is described, highlighting the potential of these therapeutic options.

Introduction

In 2018 alone, cancer was responsible for an estimated 9.6 million deaths worldwide in countries of all income levels, ranking second place in the leading causes of death, behind cardiovascular diseases (1). Owing to population growth, aging, and adoption of lifestyle behaviors associated with cancer risk, this number is expected to rise by about 70% over the next 20 years (2, 3). Still, even though these impressive numbers demonstrate that cancer burden remains a major challenge worldwide, recent developments in personalized medicine and novel treatment

Abbreviations: ADCC, antibody-dependent cellular cytotoxicity; CAR, chimeric antigen receptor; CAR-T, chimeric antigen receptor T-lymphocytes; CDC, complement dependent cytotoxicity; CDV, canine distemper virus; CHOP, cyclophosphamide, doxorubicin, vincristine and prednisolone; cNHL, canine lymphoma; CTLA-4, cytotoxic T-lymphocyte associated protein 4; DLBCL, diffuse large B-cell lymphoma; EGFR, epidermal growth factor receptor; FDA, US Food and Drug Administration; GM-CSF, Granulocyte-macrophage colony- stimulating factor; HER2, human epidermal growth factor receptor 2; hNHL, human non-Hodgkin lymphoma; HSP, heat shock proteins; HSPPC, immunogenic tumor specific peptides; LMI, large multivalent immunogen; mAbs, monoclonal antibodies; NHL, non-Hodgkin lymphoma; PBMC, peripheral blood mononuclear cells; PD-1, programmed-death 1; PD-L1, PD ligand 1; scFv, single chain variable fragment; TERT, telomerase reverse transcriptase; USDA, United States Department of Agriculture.



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current
- Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



approaches, such as immunotherapy, have raised hope of significantly improving cancer survival (2).

The concept of harnessing the host's immune system to treat cancer can be traced back decades, however only in recent years immunotherapies have emerged as a clinically validated and effective treatment strategy (4). Nowadays, cancer immunotherapy has emerged as a fast-growing field and rapidly became the fourth pillar of cancer care, along with surgery, cytotoxic therapy and radiotherapy (5). More recently the successes of clinical breakthroughs, such as checkpoint inhibitors and engineered T cells, revitalized the field and highlighted the opportunities that immunotherapeutic approaches can offer, which culminated in the nomination of "cancer immunotherapy" as 2013's Breakthrough of the Year by Science (6, 7). In 2018, the Nobel Prize in Physiology or Medicine was jointly awarded to James Allison (University of Texas MD Anderson Cancer Center) and Tasuku Honjo (Kyoto University School of Medicine) for their discoveries leading to new approaches in harnessing the immune system to fight cancer (8-12).

However, by transforming the cancer therapeutic landscape, this complex modality brought unique challenges to the drug discovery community. In fact, as more cancer patients have received immunotherapies, some of the major drawbacks of these treatments have become clear. One of the major issues is to determine the sub-populations of patients who will respond and who will experience significant toxicities (13). In fact, the challenge now is to extend the range of patients that benefit from immunotherapy while minimizing treatment-related adverse events. To address this, it is crucial to identify factors predictive of response that may help to properly select patients for treatment, identify rational combination therapies. and define progression and resistance (14). This is particularly critical when developing cancer immunotherapies, considering that the patient's immune system is expected to be as significant as tumor-related aspects when determining response and toxicity (15).

Clinical translation of cancer immunotherapy depends on preclinical investigation and rodent models have been the foundation of preliminary basic investigation and safety assays (16). However, these models underrepresent the heterogeneity and complex interaction between the human immune cells and cancers. Indeed, laboratory mice rarely develop spontaneous tumors, are housed under specific-pathogen free conditions that greatly impact immune development, and incompletely model main characteristics of the tumor/immune microenvironment, cre-

ating challenges for clinical translation. As a result, these murine models have failed to correlate with clinical success rates, demonstrating an urgent need for innovative pre-clinical models (17–19). Thereby, the use of alternative animal models is pivotal to bridge the translational gap between murine models and human clinical studies. In particular, preclinical models displaying intact immune systems that closely resemble the human immune response, present comparable, spontaneous oncogenesis and immune interactions similar to humans, and can model key clinical outcomes such as efficacy, dose response, and toxicity, will be critical for translational cancer immunotherapy research (15).

Thus, comparative medicine offers an important platform with innovative complex cross-species models that allow the research of novel therapeutic strategies and agents for diseases that are common to animals and humans (20, 21). Notably, the canine model represents a powerful resource of models for cancer immunotherapy research. Dogs are an appealing outbred combination of companion animals that experience spontaneous cancer development in the setting of an intact immune system (15). Besides, naturally occurring tumors in dogs present many clinical, pathological, immunologic, molecular, diagnostic and therapeutic sim-



- Tabla de contenido:
- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current
- Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



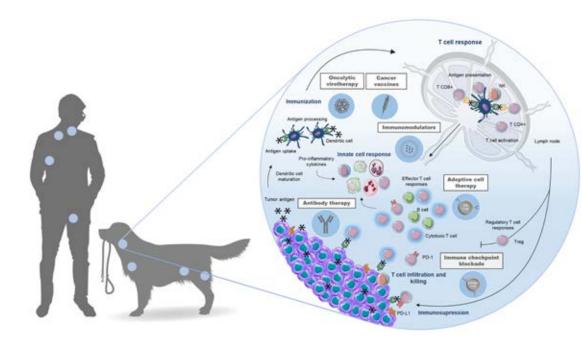
ilarities to those observed in humans, that are difficult to reproduce in other models (22–25). This allows studying the complex immune interactions during the course of treatment while also addressing long-term efficacy and toxicity of cancer immunotherapies (15).

Nevertheless, the integration of the canine model in immunotherapeutic approaches research requires diagnosis, staging and treatment response assessment, optimization and standardization, to perform large and organized clinical trials and to achieve conformity when analyzing data (26).

Driven by the great success accomplished with the application of immunotherapies in the treatment of human non-Hodgkin (hNHL) and by the remarkable similarities of canine non-Hodgkin lymphoma (cNHL) to its human counterpart, cNHL has been one of the main focus of comparative research regarding the development of immunotherapeutic approaches for dogs (Graphical abstract).

Rationale for a Canine Model of Lymphoma

For a long time, research in lymphoma has benefited from traditional mouse models, however the paucity of truly representative models has hindered



GRAPHICAL ABSTRACT. Graphical Abstract. The application of canine lymphoma as an animal model for immunotherapeutic approaches in comparative medicine provides an integrated drug discovery platform that maximize interdisciplinary cooperation and leverage commonalities across humans and dogs for the development of novel immunotherapies against non-Hodgkin lymphoma, benefiting both species.

complete understanding of disease biology and drug development. With the introduction of genomics technology, non-traditional animal models have been more accessible and the leverage of these opportunities may represent a novel strategy to accelerate disease research and new drug discovery (27). Furthermore, there is an increasing number of studies demonstrating that spontaneously arising lymphoma in dogs could be an invaluable resource to study the biology and treatment of this disease (28). As such, the cNHL model may help to bypass many of the limitations associated with the use of murine models while presenting other additional advantages (29, 30).

The cNHL shares many remarkable similarities with its human counterpart (29, 31–34). The incidence of cNHL of 15–30/100 000 is similar to human incidence (35, 36), though additional studies indicate that the incidence of cNHL may be higher (37). Classification and grading schemes of cNHL were designed to reflect the equivalent in peo-



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current
- Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



ple and facilitate comparison. In fact, the 2008 revised World Health Organization classification based on the Revised European American Lymphoma classification system, which attempts to group lymphomas by cell type, phenotypic, genetic and molecular aspects, is the current standard for the diagnosis and classification of human lymphoma, also serves as the basis for the current canine recommendations (38, 39). The use of these current World Health Organization guidelines as a template, allowed describing 20 cNHL entities, among nearly 50 discrete subtypes of hNHL. Moreover, B-cell lymphoma is more prevalent than T-cell lymphoma in both species and diffuse large B-cell lymphoma is the most common type of non-Hodgkin lymphoma (NHL) in both humans and dogs (38). Finally, treatment modalities for cNHL are similar to those used for human lymphoma (radiation, corticosteroids, chemotherapy) and CHOP (cyclophosphamide, doxorubicin, vincristine, and prednisolone)-based chemotherapy agents are typically used to treat it. Response to treatment and resistance also present clinical patterns similar to hNHL (27).

From a drug development perspective, the canine model represents a large and long-lived animal model, evolutionarily more closely related to humans than rodents, that provides a more accurate assessment of the pharmacokinetic/ pharmacodynamic parameters, while determining safety and efficacy of new therapeutic agents and approaches (27, 40). Moreover, the relatively fast disease progression rate allows obtaining early conclusions from clinical trials. In fact, a randomized clinical trial in pet dogs requires ~1–3 years, whereas a human clinical trial takes about 15 years to be completed. This short timeline allows to integrate the findings of pet trials on human trials, including toxicity, response, pharmacodynamics, dose, regimen, schedule, biomarkers and responding histology assessment (28).

Another main advantage of the canine model is that cNHL is a spontaneously occurring tumor in an immune-competent host, in contrast to murine xenograft or genetically engineered mouse models. This natural occurring cancer setting offers genetic diversity similar to human lymphoma and allows studying biological mechanisms, such as tumor initiation and promotion. Moreover, the pet dog model harnessed by the evolutionary conservation allows to identify similarities between canine and human lymphomagenesis, for example in identifying key "driver" gene mutations common to both species (27).

The benefits of the cNHL model extend beyond the biological advantages of a spontaneously occurring tumor in a large animal. Pet dogs share the same living environment as their caregivers,

allowing to study environmental risk factors of developing lymphoma (27, 28). For example, an epidemiological study in France demonstrated a correlation between the incidence of cNHL and hNHL and reported a strong association between cNHL and the distribution of waste incinerators, radioactive waste or other polluted sites (41). Moreover, there is an increased prevalence of lymphoma within specific dog breeds (42) and a breed-specific distribution of B-cell and T-cell lymphomas (43). This in association with the well-organized multi-generational pedigrees kept by many breeders, represents a unique genetic advantage that allows mapping of lymphoma predisposition genes with strategies that are not possible in humans (28).

The final rationale for using dogs with lymphoma as an animal model relies on the dual benefit concept of this comparative research approach. Improved current health care have promoted the increase of dogs lifespan, allowing the diagnosis of late-in-life diseases such as cancer (44). Lymphoma particularly is one of the most common malignancies in dogs (28). In addition, the social status of dogs as companion animals allows them to benefit from high quality health care and the ethical exploration of translational approaches. Moreover, these initiatives are also motivated by the increasing healthcare standards



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current
- Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



demanded by pet owners, creating a need for novel cancer therapies in veterinary settings (20, 21, 45). Altogether, the use of the cNHL model represents a unique opportunity to strengthen the collaboration between human and veterinary medicine in lymphoma research, that ultimately will lead to advances in the care of people and dogs affected by NHL, a critical medical unmet need of today's society (22, 27).

A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology

NHL, an heterogeneous group of cancers characterized by a diverse class of lymphocyte proliferations, represents one of the most common neoplasias in both humans and pet dogs (38, 46). hNHL constitutes the most commonly reported hematological malignancy worldwide, comprising nearly 3% of all cancer diagnoses. The highest incidence rates are found in Australia/New Zealand, Northern America, and Europe. In the United States, NHL is the seventh most common and sixth most common cause of cancer-related death, in Europe is the eleventh most common and the fourteenth most deadly malignancy and its incidence has nearly doubled since the early 70s (47, 48). NHL represents 90% of all lymphomas and encompasses an heterogeneous group of cancers characterized by the proliferation of malignant lymphocytes, 85–90% of which arise from B lymphocytes, whereas the remaining derive from T cells or natural killer cells. This diverse group of malignancies usually develops in the lymph nodes, but can occur in almost any tissue, ranging from the more indolent follicular lymphoma to the more aggressive diffuse large B-cell (DLBCL) and Burkitt's lymphoma (49). NHL patients typically present with persistent painless lymphadenopathy, but some patients may present with constitutional symptoms or with involvement of organs other than those from the lymphoid and hematopoietic system (50).

The basis of treatment selection requires an accurate diagnosis, a careful staging of the disease, and the identification of adverse prognostic factors. Regardless, NHL patients most commonly receive chemoimmunotherapy as initial treatment. Radiation therapy may be performed if patients have early-stage disease (50). Response rates to conventional chemotherapy are generally >50%; however, most patients eventually relapse. Moreover, the toxicity of conventional chemotherapy often limits its efficacy (47).

In the last decades, the scientific community has been reporting cases of

therapeutic success using monoclonal antibodies (mAbs) in the treatment of NHL in humans. One of the most successful examples has been the application of mAbs targeting the surface antigen of CD20 (Rituximab[®]) in combination with chemotherapy regimen CHOP, which has revolutionized the treatment of B-cell lymphoma by significantly improving disease-free interval and overall survival, with minimal toxicity (51, 52). Even though current therapy strategies have significantly improved prognosis of patients diagnosed with NHL. a substantial fraction of patients relapse or are refractory to these treatments. Several treatment shortcomings have been identified as research priorities, however rituximab resistance and refractory/relapsed disease represent major current and emerging challenges (53–55).

Thus, a plethora of new immunotherapeutic approaches to treat lymphoma have been ensued. The most exciting classes of immunotherapies comprise chimeric antigen receptor T-cells, bispecific antibodies, immune checkpoint inhibitors, and vaccines. The advent of such innovative therapies brought unique challenges that need to be considered, including the assessment of the appropriate timing of treatment, optimal patient population, duration of therapy, toxicity, and cost. Hence, future studies need to focus on the de-



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non– Hodgkin's Lymphoma in Comparative Oncology
- Current
- Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



velopment of new strategies, models and paths in order to optimize the drug development of novel immunotherapies for hNHL (56).

Owing to shared molecular, incidence, genetic, histopathologic and clinical features, cNHL has been proposed as a comparative animal model for the research of novel therapeutic agents and approaches for hNHL (22–24, 30). cNHL displays several histological subtypes and patients can manifest a wide range of symptoms. However, most suffer from generalized lymphadenopathy (multicentric form) and are diagnosed with intermediate to high-grade lymphoma, more commonly of B-cell origin. Without treatment, the disease has high mortality (28), requiring prompt chemotherapy to achieve temporary remission and prolonged survival. Chemotherapy still remains the mainstay for the treatment of cNHL and regardless of the numerous published chemotherapeutic protocols, it seems we have reached a stalemate concerning what this treatment modality has to offer in standard settings (57). Yet, cure is rarely achieved and the majority of dogs relapse with lethal, drug-resistant lymphoma. The 12 month median survival barrier and the 20 to 25% 2 years survival rates demonstrate an urgent and unmet need in veterinary medicine to develop new treatment strategies for refractory disease (58-61).

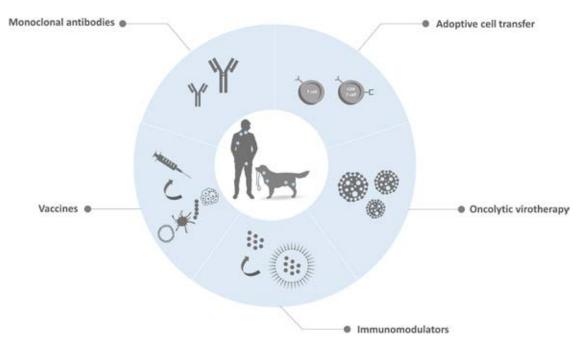


Figure 1. Schematic representation of available and under development immunotherapy strategies for cNHL. Currently, several research groups are actively investigating new immunotherapies that mobilize the patient's own immune system to treat NHL in both pets and pet owners. These treatment modalities include therapeutic mAbs that promote the direct or indirect death of cancer cells, adoptative cell transfer that uses a patient's own cells to induce antitumor activity, oncolytic virotherapy that involves the replication-competent virus in the elimination of cancer, immunomodulators that aim to enhance immune responses and tumor control and vaccines that stimulate a patient's own immune system against cancer cells.

Thus, immunotherapies for cNHL are a promising approach for the development of a new class of anti-cancer therapeutics, which will in many cases benefit humans and man's best friends. To demonstrate the potential of these strategies, available and under development immunotherapies for cNHL will be summarized below (**Figure 1**).



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



| Monoclonal antibodies therapy | Study | References |
|---|--------------------------|---------------------------|
| mAb 231 | Preclinical and clinical | (64–66) |
| Anti-HLA-DR (L243) | Preclinical and clinical | (67) |
| Anti-HLA-DR (IMMU-114) | Preclinical and clinical | (67) |
| Anti-CD20 (6C8) | Preclinical | (51) |
| Anti-CD20 (1E4-clgGB) | Preclinical and clinical | (68) |
| Anti-CD20 (NCD1.2) | Preclinical | (69) |
| Anti-CD20 (AT-004) | Preclinical and clinical | Aratana Therapeutics® |
| Anti-CD52 (AT-005) | Preclinical and clinical | Aratana Therapeutics® |
| Anti-CD20 (1E4-clgGB) plus CD47 blockade | Preclinical | (45) |
| Anti-CD20 (4E1-7-B_f) | Preclinical and clinical | (70) |
| Adoptive cell transfer therapy | Study | References |
| Autologous T cells | Preclinical and clinical | (71, 72) |
| Autologous T-cells | | Aurelius BioTherapeutics® |
| Autologous T-cells plus tumor vaccination | | Elias Animal Health® |
| CD20 CAR-T cells | Preclinical and clinical | (73) |
| Oncolytic virotherapy | Study | References |
| Canine distemper virus (pCDVeGFPN) | Preclinical and clinical | (74, 75) |
| Newcastle disease virus | Preclinical and clinical | (76, 77) |
| Reovirus (dearing strain) | Preclinical and clinical | (78, 79) |
| Immunomodulator therapy | Study | References |
| Autologous tumor antigen-coated microbeads with IL-2 and GM-CSF | Preclinical and clinical | (80) |
| Vaccine therapy | Study | References |
| Intralymphatic autologous tumor vaccine | Preclinical and clinical | (81–83) |
| Autologous CD-40-activated B-cells loaded with total RNA from autologous lymphoma cells | Preclinical and clinical | (84) |
| DNA-vaccine targeting canine telomerase reverse transcriptase | Preclinical and clinical | (85–87) |
| Autologous tumor heat shock proteins (APAVAC) | Preclinical and clinical | (88–90) |

Table 1. Immunotherapyapproaches developed and underdevelopment for cNHL.

Current Immunotherapies for Canine Non-Hodgkin's Lymphoma

After decades of weakening or even eliminating the patient's immune system with chemotherapy, now the trend is to harness the ability of the immune system to eradicate cancer (62). Over the past decades immunotherapy has moved into the forefront of cancer care due to unprecedented clinical success in a wide range of malignancies, sometimes even in late stages of disease (63). The field of veterinary immunotherapy holds similar promise for companion animals with cancer, and several efforts have been made in order to develop veterinary specific immunotherapies (**Table 1**). In the nearby future, it is hoped that tumor immunotherapy will become a valid therapeutic tool in veterinary oncology, along with chemotherapy, radiotherapy and surgery.

Monoclonal Antibodies

In cancer therapy, the main purpose of antibody treatment is to promote the direct or indirect death of cancer cells and a number of strategies have been successfully employed. MAbs can bind to target cancer cells and directly promote signaling-induced death or can mediate an anti-tumor immune response by promoting antibody-dependent cellular cytotoxicity (ADCC) and inducing complement-dependent cyto-



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



toxicity (CDC) (91). In the case of ADCC responses, mAbs bind to target tumor cells while the mAb Fc region engage with the FcyRs on the surface of effector cells, including natural killer cells and macrophages. These immune cells cause phagocytosis, apoptosis or lysis of the target cells. In CDC responses, mAbs promote directly target cell death through the development of a complement cascade membrane attack complex. Furthermore, mAb-based therapies can also block growth-promoting pathways, such as angiogenesis or can directly regulate the anti-tumoral activity of adaptive immune cells by blocking inhibitory signals responsible for limiting T cell activation (92). Most marketed mAbs consist of a full-length IgG molecule. By providing a long half-life and effector functions, these molecules have been presenting a guite successful application in therapeutics. However, this conventional antibody format present some drawbacks that limit their clinical use and there is a range of therapeutic applications in which other antibody formats may be more appropriate. To address these major issues, smaller antibody scaffolds such as the Fab or the single chain variable fragment (scFv) or single domain antibody are emerging as alternative therapeutic agents (93) (Figure 2).

MAbs are the most commonly used and approved cancer immunotherapy meth-

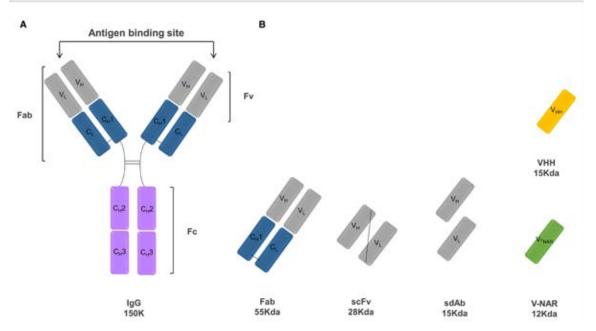


Figure 2. Schematic representation of various antibody formats including a conventional IgG antibody (**A**) and antibody fragments (**B**) of interest. (**A**) The basic unit of a conventional IgG antibody is a polypeptide consisting of a pair of identical heavy and light chains held together by disulfide bonds. Light chains are comprised of one constant domain (CL) and one variable domain (VL), whereas heavy chains are comprised of three constant domains (CH1, CH2, and CH3) and one variable domain (VH). The antigen-binding site is composed by the variable domains of both the heavy and light chains. In turn, the Fc constant region is responsible for the recruitment of the immune system effector functions. (**B**) Antibody fragments that can be engineered from a conventional IgG include: antigen-binding fragment (Fab), single-chain Fv fragment (scFv), heavy and light single domains antibodies (sdAbs) and natural camelid variable domain (VHH) and shark variable domains (V-NAR).

od in clinical practice (94). The use of an antibody targeting the human surface antigen CD20 (Rituximab®), expressed on B-lymphocytes has revolutionized the treatment of B-cell lymphoma (51, 52). Rituximab is a chimeric antibody and was the first US Food and Drug Administration (FDA) approved mAb for the treatment of human cancer, being used for the treatment of most B-cell NHL and subtypes of acute lymphocytic leukemia (95–97). This immunotherapy provided significant enhancements in the efficacy of treatment vs. existing non-mAb therapies, increasing the rate of durable remissions from 30 to 60% (51).

Even though immunotherapy has a crucial role in the treatment of B-cell malignancies in humans, its role in canine



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



lymphoma remains limited. Immunohistochemistry using mAbs that recognize the CD20 intracellular domains demonstrated the presence of CD20 in canine lymphoma tissue samples (98, 99). However, Rituximab[®] and other anti-human and anti-mouse antibodies that recognize the CD20 extracellular domains, failed to bind to canine CD20, even though the reported epitopes are conserved between human and canine CD20 (100). For that reason, it is evident that technology to speciate antibodies is essential when developing similar passive immunotherapy strategies for canine cancer patients.

Interestingly, in 1992, prior to FDA approval of Rituximab, the United States Department of Agriculture (USDA) approved the licensing of mAb 231 for use in cNHL. mAb 231 consists of a murine-derived mAb that showed both in vitro (64) and in vivo activity and served as adjuvant therapy following remission induction with chemotherapy (65, 66, 81). Unfortunately, subsequent clinical trials failed to confirm the initial study results and the antibody epitope was never identified, which culminated in its commercial suspension (65).

Since then, driven by the great potential of the canine lymphoma model for immunotherapeutic approaches, academic research groups and industry began exploiting the dual benefit approach of comparative medicine. One of the first examples was a pilot study that aimed to assess the suitability of the canine lymphoma model to evaluate endpoints with clinical relevance of anti-HLA-DR mAb treatment before proceeding to an extensive trial in pet dogs, and eventually human research. In vitro studies revealed that L243, a murine IgG1 anti-HLA-DR, binds to canine healthy lymphocytes and lymphoma cells, inducing apoptosis in cNHL cells. In turn, in vivo studies confirmed the L243 treatment safety in healthy dogs and dogs with lymphoma and its binding activity to lymphoma affected lymph node samples. Preliminary data also showed that a subset of patients with advanced lymphoma achieved transient disease stabilization after L243 treatment (67). Furthermore. this work also reported that hL243y4P (IMMU-114), a humanized IgG4 anti-HLA-DR, under preclinical evaluation for human trials, also bound to cNHL cells. Finally, the assessment of IMMU-114 treatment in healthy canine patients indicated a safety and pharmacokinetic profile similar to L243. Overall, these findings supported the use of cNHL in safety and efficacy studies of anti-HLA-DR mAbs for both veterinary and human medicine (67).

Advances in speciation technology has also led to several clinical trials in pet dogs since "caninization" of antibodies is crucial when approaching canine patients with cancer. With this in mind, research groups focused on the technique to generate caninized antibodies, which resulted in the development of a canine anti-EGFR (epidermal growth factor receptor) mAbs (101) and nowadays is also being offered as a service by companies (Creative Biolabs).

Considering the success achieved with Rituximab in human medicine, several studies also focused on developing canine anti-CD20 antibodies. An anti-canine CD20 mAb (6C8) that recognized the extracellular domain of canine CD20 and showed high-affinity binding to canine CD20 in solution and its native conformation on canine B-cells was developed. This mAb promoted phagocytosis of B-cell lymphoma cells by macrophages, but in its current framework did not induce direct cytotoxicity or CDC (51). In the same year, Rue et al. reported the development of an anti-canine CD20 antibody (1E4) and the generation of a canine chimeric molecule for therapeutic use. This clone bound a similar extracellular domain as rituximab, and flow cytometry analysis confirmed that 1E4-based chimeric versions were able to stain canine B cells and canine CD16a, a receptor that mediates ADCC responses. Moreover, the best chimeric mAb candidate depleted the number of circulating B cells in healthy beagles in an in vivo study. Though, the clinical efficacy in dogs with canine B cell lym-



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



phoma remains unknown (68). Likewise, a new anti-CD20 mAb (NCD1.2) that bound both human and canine CD20 has been developed, in order to strengthen human-canine comparative model. NCD1.2 bound to clinically derived canine cells including B-cells in peripheral blood and in different histologic types of B-cell lymphoma. Heavy chain and light chain genes from the NCD1.2 hybridomas were cloned and packaged as scFv into a phage-display library. Recombinant anti-CD20 scFv were identified and selected as a possible useful tool for evaluation in bioconjugate-directed anti-CD20 immunotherapies in comparative medicine (69). Although these works established several canine anti-CD20 mAbs candidates with high potential for therapeutic use, their clinical efficacy in dogs bearing B-cell lymphoma remains unknown.

A canine anti-CD20 mAb (AT-004) has been fully approved by USDA for clinical usage in dogs with B-cell Lymphoma and is currently being commercialized in the United States and Canada. Treatment with AT-004 (Aratana Therapeutics), an anti-canine CD20 was subject to a prospective randomized clinical trial and preliminary results suggested an improved median progression-free survival of dogs with B-cell lymphoma (102). Yet, these results were published in a conference abstract and peer-reviewed results are still lacking.

Another work evaluated the combination of CD47-blockade with 1E4-clgGB. a canine-specific antibody to CD20. Although 1E4-clgGB could elicit an in vivo therapeutic response against canine lymphoma as a single agent, superior responses were observed when combined with agents targeting CD47, an immune checkpoint that enables the evasion of tumor cells to phagocytosis promoted by therapeutic antibodies, such as anti-CD20 mAbs. The combination of CD47-blocking therapies with 1E4-clgGB resulted in synergic antitumoral effects in vitro and in vivo, eliciting cures in 100% of mice bearing canine lymphoma (45). However, there is no anti-CD20 antibody treatment for cNHL currently available. More recently a novel approach of developing an anti-canine CD20 monoclonal antibody using rats as a host species renewed hopes of finally obtaining an antibody-based therapy for cNHL. This work culminated in the generation of a mAb capable of inducing cell death of B cell lymphoma cell lines, however this mAb was incapable of eliciting CDC and ADCC responses. To tackle these limitations, this antibody was modified into a canine/rat anti-CD20 chimeric, which resulted in the alterations of its characteristics into a potent CDC and ADCC inducer. Furthermore, its defucosylation resulted in a 10-fold higher ADCC activity. The in vivo antitumor activity of this

improved mAb version was assessed, revealing a tumoral growth inhibition in a cNHL xenograft mouse model and a peripheral B cell depletion in healthy beagles (70). Finally, AT-005 (Aratana Therapeutics), a caninized mAb targeting CD52 on T cells, has obtained conditional USDA approval for the treatment of T-cell lymphoma and is currently being evaluated in clinical trials (62).

The success of mAbs in human medicine strongly encourages veterinary medicine to develop similar therapeutics for our pets. Regardless of their potential, little speciated mAbs have been established for veterinary application and fewer were investigated in clinical trials enrolling companion animals. Nonetheless, the approval of the first mAb by the European Union Agency for the treatment of atopic dermatitis in dogs—Lokivetmat, a caninized, anti-canine IL-31 mAb (103), highlighted the impact that biological therapies may have in veterinary practice. In the oncology setting, mAbs have the capacity to treat a diversity of hematological and solid malignancies, do not need to be a personalized product and manufacturing methods are well-established, minimizing the cost associated limitation. Hence, mAb-based therapy is one of the most promising immunotherapy strategies in veterinary settings (63).



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's
- Lymphoma
- Discussion
- Author Contributions
- Funding



Adoptive T-Cell Transfer

Adoptive cell therapy is a term that was first used to describe the infusion of lymphocytes to mediate rejection of organ allografts and to treat tumors (104). This immunotherapeutic option represents the most effective treatment for patients with metastatic melanoma inducing visible cancer regression in ~50% of patients. Adoptive cell therapy is also associated with clinical improvement in selected patients with post-transplant lymphoproliferative diseases caused by Epstein-Barr virus infection (105). More recently, gene transfer techniques developed in the 1990s allowed to convert normal lymphocytes into lymphocytes with anti-cancer activity by redirecting the specificity of T cells with the use of T-cell receptors or chimeric antigen receptor (CARs). CARs are engineered receptors that graft a defined specificity onto an immune effector cell, typically a T cell, resulting in the augment of T-cell function (104). This innovation represented a possibility of extending adoptive cell immunotherapy to patients with a large diversity of cancer types (105). In humans, treatment of advanced B-cell leukemia or lymphoma using CAR T-cells has demonstrated promising clinical responses, resulting in the approval of two autologous CAR T-cell therapies (Kymriah[™] and Yescarta[™]) by the FDA (106, 107). These therapies

are both genetically modified autologous T cells expressing a CD19-specific CAR, lysing CD19-positive targets (107).

By displaying an intact immune response and genetic similarities to humans, dogs may potentially inform the development of the later-stages of human clinical trials, while studying the use of adoptive cell therapy in veterinary malignancies, including hematologic neoplasias (71, 72). In fact, there is evidence that canine cancer. and specifically cNHL, respond to cellbased immunotherapy. Half a century ago, the Fred Hutchinson Cancer Center established hematopoietic cell transplantation for canine lymphoma (108). At first, the therapeutic value of this practice was solely associated with the administration of high-dose chemotherapy and radiation prior to the transplant. Yet, a larger retrospective study confirmed that, despite the use of the same chemotherapy and radiation protocols, dogs that received an allogeneic transplant from a littermate exhibited a significantly lower relapse rate, in contrast to dogs that received their own (autologous) bone marrow stem cells. This effect was later known as the "graft vs. leukemia/ tumor effect" and is mainly promoted by activated allogeneic T cells that recognize and react to antigen differences, and therefore also attack residual tumor cells (109).

Since then, few studies have focused on the scientific and clinical investigation of cell-based immunotherapies for canine patients. O'Connor et al. conducted a clinical trial to test non-specific autologous T cells isolated from dogs with NHL and expanded ex vivo using a novel artificial antigen presenting cell protocol (71, 72). Infused cells were detected in the blood for longer than 49 days and trafficked to secondary lymphoid organs, confirming the safety of adoptive transfer of autologous T cells in dogs. Furthermore, this adoptive immunotherapy demonstrated to be viable and effective in improving first remission and overall survival periods in dogs with multicentric lymphoma (71, 72).

Notably, a few biotech companies have emerged in the area of autologous T-cell based therapy for veterinary medicine. One example is Aurelius BioTherapeutics that provides a service that expands for 2–3 weeks autologous lymphocytes collected from dogs with canine lymphoma, in order to increase T cell numbers exponentially and to activate them to be responsive to antigens presented by the tumor cells before reinfusion. However, the methods used for the activation and expansion of dog's immune cells and the clinical benefit of this therapy are not disclosed. In turn, Elias Animal Health included a vaccination procedure prior to cell collection, aim-



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



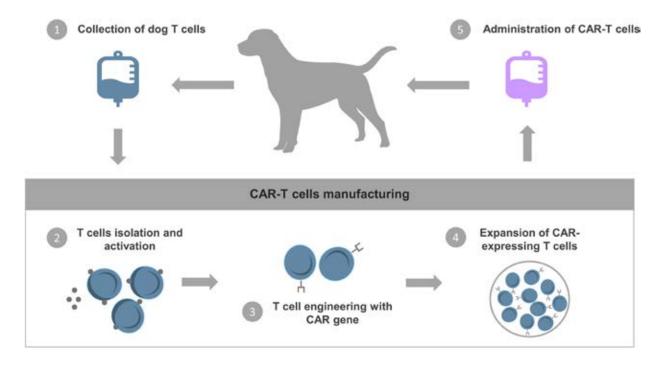


Figure 3. CAR-T cells therapy. The basic procedures for CAR-T cell therapy start with the collection and extraction of T cells from the pet's peripheral blood. The T cells are then genetically engineered in vitro to express chimeric antigen receptors (CARs) that can recognize specific tumor-associated antigens and activate self-proliferation and cytotoxicity. Finally, CAR-T cells are expanded and reinfused into the patient.

ing to improve cancer-cell specificity of their autologous T-cell therapy. The vaccine is obtained from the excised tumor material and is given through an intradermal route. Additionally, a brief cycle of chemotherapy may be administered prior to the infusion, which has shown to result in better acceptance of the lymphocyte therapy in humans. The preliminary results revealed that overall survival may be prolonged with this adoptive cell-based therapy, indicating that this immunotherapy prompts an antitumor vaccine-like effect that extends canine patients' lives, even when the disease is not fully eradicated. The holding company is pursuing regulato-

ry approval, which would qualify it as the first approved and commercialized cell therapy for dogs (106).

More recently, researchers have started to explore chimeric antigen receptor T-lymphocytes (CAR-T) cell therapy for dogs (**Figure 3**). CARs engineering consists of modifying T-cells to express artificial receptors formed by a tumor-antigen specific scFv linked to an intracellular signaling domain and co-stimulatory molecules. Because CARs work in a MHC independent manner, antigen presentation do not rely on patient antigen presenting cells. Moreover, CARs do not have to be syngeneic to the patient immune system (63). Canine T cells expressing a HER2 (human epidermal growth factor receptor 2)-specific CAR have been produced and showed anti-tumoral activity in vitro against canine osteosarcoma cells expressing HER2 (110). This work proved that a successful ex vivo expansion of HER2-CAR specific T-lymphocytes is possible. Yet, no canine patients have been treated. Ongoing studies aim to develop a canine CAR-T cells for the treatment of B-cell lymphomas and other malignancies (63). Importantly, protocols for the propagation of CD20 CAR-T cells have been reported (73, 111). Researchers transfected the CD20 CAR into the expanded T-cells us-



- Introduction
- Rationale for a Canine
 Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



ing electroporation of CAR mRNA. Unfortunately, even though this strategy allows to avert using retro or lentivirus, mRNA transfection results in variable efficiency and transient transcriptional activity that ceases following 24 to 48 h. It was reported the treatment of one dog diagnosed with lymphoma with these transfected T-cells, however it only presented a short-term partial response (73, 106). This limited clinical response can be due to the inability of these transfected cells to expand in vivo, considering that human studies demonstrated that in vivo expansion is a requirement for durable responses. Furthermore, this treatment protocol did not include chemotherapy sessions prior to the CAR-T cells infusion, a common practice used in the human treatment to deplete inhibitory immune cells that has shown to potentiate clinical efficacy. In the case of dogs, the addition of this procedure could also minimize the risk of triggering a canine anti-mouse antibody immune response, considering that most scFvs derived from murine mAbs, thereby increasing the risk for an anti-CAR T cell immune response. To conclude, reported data proved the feasibility of generating canine CAR-T cells, however the necessary logistics and expenses are expected to be considerable.

Oncolytic Virotherapy

Oncolytic virotherapy is a new concept of immunotherapy recently introduced that involves the replication-competent virus in the elimination of cancer. By infecting tumor cells, oncolytic virotherapy can stimulate de novo or enhance pre-existing native immune response. The majority of developed oncolytic virus are genetically altered to promote tumor tropism while reduce virulence against healthy host cells. Thereby, oncolytic virotherapy have the ability to promote a proinflammatory environment by improving antigen release/recognition and promoting immune activation, while reverting immunosuppression of tumor cells and improving the efficacy of other forms of immunotherapy (112, 113). Although several oncolytic virotherapies are being developed in preclinical and clinical settings, currently the only oncolytic viral therapy approved by FDA is talimogene laherparepvec (T-Vec or Imlygic) for advanced melanoma (114). In veterinary medicine, several studies evaluated natural and genetically modified oncolytic viruses for dogs diagnosed with cancer, showing some encouraging results. However, the majority of the developed research work focused on in vitro results. with a few reporting in vivo studies, of which most were isolated clinical case reports (115).

Regarding cNHL, a study reported that a recombinant strain of the canine distemper virus (CDV)—pCDVeGFP Δ N was capable of infecting cNHL cell lines in vitro, inducing significant apoptotic cell death. The pCDVeGFPAN strain also efficiently infected primary canine B and T-cell lymphoma cells, though its oncolytic efficacy was not proved (74). Another work evaluated the anti-tumoral effect of CDV infection using an attenuated strain in seven dogs with naturally occurring lymphoma. For this purpose, single or multiple doses of the virus were injected intratumorally. This study reported low toxicity with a severe fibrotic reaction in the injection site. Immunohistochemistry analysis revealed a variable positive detection of CDV antigen in treated lymph nodes, while co-culturing enabled virus isolation from treated lymph nodes, but not from distant nodes or from peripheral blood mononuclear cells (PBMCs). Furthermore, this treatment promoted a strong anti-CDV antibody response (75). However, one of the major drawbacks of this immunotherapy is that CDV belongs to the regular vaccination schedule in dogs and pre-existing antibodies can limit its efficacy (116). Another group explored the oncolytic properties of a vaccine strain of Newcastle disease virus, an attenuated lentogenic strain presenting low virulence, on a human large B-cell lymphoma cell



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's
- Lymphoma
- Discussion
- Author Contributions
- Funding



line and on primary canine B-cell lymphoma cells. The group used as controls healthy PBMCs from humans and dogs. Newcastle disease virus infection decreased cell viability in both human and dog lymphoma cells when compared to untreated controls, with minimal tropism toward healthy PBMCs. In the same work the authors reported the viral biodistribution in a canine patient diagnosed with T-cell lymphoma, 24 h following the virus intravenous injection. Immunohistochemistry and endpoint PCR demonstrated viral dissemination in the salivary gland, kidney, stomach and lung, but not in tumor samples, with no abnormal findings on the histopathological evaluation (76). Curiously, a complete and long-term clinical response was reported in a dog diagnosed with lymphoma resistant to chemotherapy (76, 77). Although these preliminary data revealed that Newcastle disease virus could represent a promising oncolytic virotherapy, future studies are required to determinate the best therapeutic regimen and define the proper safety protocol (117).

One of the oncolytic virotherapies that has gathered most interest amongst the scientific community, due to the promising results obtained in multiple phase I and II clinical trials, is the dearing strain of Reovirus (Reolysin®, from OncolyticsTM Biotech Inc., Calgary, AB, Canada) (118). In dogs, Reolysin® showed promising in vitro results for the treatment of a variety of malignancies, such as mastocytoma, lymphoma, mammary gland tumors and melanoma. In fact, in vitro studies showed apoptosis induction and a significant cell viability reduction in both T and B-cell lymphoma. Furthermore, a mouse xenograft model of canine T-cell lymphoma treated via intratumoral injection revealed significant tumor growth inhibition, compared to the control group treated with reovirus inactivated by ultraviolet (78). Notably, the safety profile of Reolysin[®] was proven in a clinical trial enrolling dogs with advanced cancer, including mastocytoma, lymphoma, oral melanoma and soft tissue sarcoma. In this work, dogs received virotherapy by intratumoral injection or intravenous injection daily for 5 days, during one or several treatment cycles. Live virus was only detected in the serum of one dog in the first chemotherapy cycle, but not in the subsequent treatment cycles. While all dogs exhibited an increase in the titer of anti-reovirus neutralizing antibodies. tumor volume reduction was observed in five dogs and six dogs presented alleviation of clinical manifestations. Furthermore, a subset of dogs revealed a good safety profile, as well as clinical response. Taking into account the experience gathered in human medicine, the combination of this immunotherapy with conventional therapies such as

chemotherapy, radiotherapy, or other could be investigated in dogs (79).

Overall, these studies provide preliminary results that support the development of oncolytic virotherapy as canine cancer therapy to benefit pets and pet-owners (115).

Immunomodulators

Cytokine therapy aims to enhance immune responses and tumor control in a variety of spontaneous oncologic diseases. In human medicine, modest success has been obtained with a low-dose IL-2 therapy delivered subcutaneously, with few side effects (119–124). Additionally, subcutaneous GM-CSF (Granulocyte-macrophage colony-stimulating factor) therapy boosts cell-mediated immune responses and improves anti-idiotype vaccines efficacy in human lymphoma (125). In canine patients, IL-2 delivered subcutaneously, intralesionally, by inhalation and via liposome-DNA complexes encoding IL-2 gene, as a monotherapy or in combination with other modalities, promoted regression in dogs with oral melanoma, soft tissue sarcoma, squamous cell carcinoma and pulmonary metastases from osteosarcoma (126–131). Likewise, in dogs with oral melanoma, combination therapy including GM-CSF delivered intralesionally, either via liposome-DNA complexes or via GM-CSF secreting transgenic xenogeneic



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



cells, resulted in regression (126, 132). Through the Comparative Oncology Trials Consortium, a Phase I safety/dose escalation study of human IL12 administered subcutaneously to dogs with melanoma was conducted. Data gathered from this study and other preclinical data allowed to inform the design of a Phase I clinical trial of IL12 in human cancer patients (133).

A phase I study enrolling 15 dogs with B-cell lymphoma tested a therapy with a combination of autologous tumor antigen-coated microbeads (large multivalent immunogen—LMI) with cytokine therapy including IL-2 and GM-CSF, following induction of remission with conventional chemotherapy. Results demonstrated no significant toxicity, no adverse effects in disease-free interval and half of the animals presented quantifiable delayed-type hypersensitivity reactions to intradermal LMI, suggestive of a specific cell-mediated immune response (80).

Although these studies show that human cytokines can be effectively used in dogs, the often-needed higher doses and the immunogenicity that they generate, limits their use. Nonetheless, the development of canine IL-15 has led to a renewed interest in cytokine therapy as an immunotherapy strategy for veterinary settings (134).

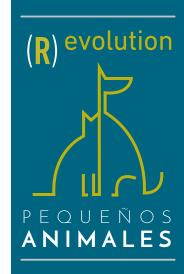
Vaccines

Therapeutic vaccines represent a viable and attractive cancer immunotherapy strategy that aim to treat late stage disease by stimulating a patient's own immune system against cancer cells (135).

Several attempts to use vaccines as a treatment for cNHL have been made. In the initial studies, Freund's adjuvant was added to lymphoma cell extracts lysates and used as a cancer vaccine strategy. Despite de fact that these early studies reported some treatment benefit (136), this was later attributed to the use of the Freund's adjuvant (137).

Later, Jeglum et al. described the use of an autologous tumor vaccine administrated via intralymphatic injection following remission induction with chemotherapy. However, results using this strategy have been conflicting (81–83).

In a clinical trial, autologous CD40-activated B-cells loaded with total RNA from autologous lymphoma cells were administered to 19 dogs with NHL as an adjuvant, following induction of a complete response with chemotherapy. Vaccination promoted an anti-tumor response and increased a lasting second remission rate, however median time to disease progression and overall survival did not show differences between groups (84). Moreover, a new approach targeting canine telomerase reverse transcriptase using a genetic vaccine, Tel-eVax, is reported. As telomerase confers immortality to cells, telomerase reverse transcriptase is overexpressed in cancer cell lines and in several tumors and undetectable in the majority of normal tissues, establishing a possible target for translational cancer immunotherapy. A DNA-vaccine targeting canine telomerase reverse transcriptase was able to prompt an immune response against telomerase in dogs diagnosed with multicentric lymphoma, and conventional chemotherapy seems not to alter the immunotherapy effects (85). The combination of this vaccine with chemotherapy using the cyclophosphamide, vincristine and prednisolone protocol resulted in a durable immune response, as well as prolonged survival in dogs with B-cell lymphoma (86). On other clinical study including 17 pet dogs, Tel-eVax in association with CHOP chemotherapy demonstrated to be safe and immunogenic and presented a significant impact on DLBCL canine patients' survival. Antibody response induced by Tel-eVax against telomerase reverse transcriptase (TERT) protein was also evaluated considering the potential that these anti-TERT antibodies may possess as surrogate biomarkers of the immune response in vaccinated dogs. Curiously, most dogs developed



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



a low but detectable seroconversion against the N-terminal of TERT protein (87).

More recently, an autologous vaccine APAVAC[®], comprised of hydroxylapatite ceramic powder with autologous heat shock proteins (HSP) purified from affected lymph node biopsy is currently available (88). HSPs resultant from tumor cells, including gp96, hsp90, hsp70, calreticulin, hsp110, and hsp170, present strong immunogenicity. Furthermore, the chaperone function of HSPs allows their combination with immunogenic tumor specific peptides (HSPPC), exposing the host to a large repertoire of tumor associated antigens for immunization. In addition, hydroxylapatite vehicles and HSPPCs functions as an adjuvant. In order to reproduce the tumor heterogeneity, each vaccine is produced for each dog. Vaccination protocol consists of four administrations within 4 weeks followed by one injection a month for 4 months in combination with dose-intense chemotherapy. In an initial phase, preliminary results showed that the administration of this autologous vaccine is effective in prolonging overall survival and the time to progression in dogs with DLBCL and multicentric indolent B-cell neoplasia, without increasing treatment toxicity (88, 89). Following, to better characterize the safety and efficacy of APAVAC®, and to find the best candidates for immunotherapy, a larger retrospective study was conducted, which included all dogs treated with chemo-immunotherapy to date. Overall, compared to dogs treated with chemotherapy only, dogs receiving the chemo-immunotherapy protocol survived significantly longer, regardless of histotype and evaluated prognostic factors. The study also confirmed the excellent tolerability of the vaccine in dogs diagnosed with B-cell lymphomas (90). Unfortunately, until now there is no information regarding the chemo-immunotherapy treatment response in T-cell lymphoma dogs.

Altogether these works clearly demonstrate the potential of the cNHL model to advance cancer vaccine strategies research to treat lymphoma both in humans and dogs.

Immune Checkpoint Blockade

Immune checkpoint inhibitors, such as those targeting CTLA-4 and the PD-1 (programmed-death 1)/PD-L1 (PD ligand 1) axis, have shown unprecedented and durable clinical effect in a wide range of malignancies and are rapidly transforming the practice of medical oncology in humans (138).

Tumor cells can successfully evade immunosurveillance and progress through different mechanisms, including activation of immune checkpoint pathways that hinder antitumor immune responses. By interrupting co-inhibitory signaling pathways, immune checkpoint inhibitors reestablish antitumor immune responses and promote immune-mediated elimination of malignant cells (139). Hematologic malignancies such as lymphoma are likely targets for this type of treatment. Several clinical trials of checkpoint blockade have been performed in hematological malignancies, with promising preliminary results, suggesting the therapeutic benefit of this approach. These results were specially promising regarding PD-1 blockade in Hodgkin lymphoma (140). To date, there are currently seven approved immune checkpoint inhibitors for the treatment of various cancers in human medicine.

Clinical trials using checkpoint inhibitors for the treatment of cNHL have yet to be conducted. Nevertheless, expression of canine PD-L1 has been reported on a variety of canine tumor types, including mastocytoma, melanoma and renal cell carcinoma (141). A preliminary study suggests that anti-PD-L1 might play a significant role in the treatment of dogs with tumors expressing PD-L1, by demonstrating that treatment of canine tumor infiltrating lymphocytes with this molecule improved interferon-y production (141). It was recently reported that PD-L1 is elevated in canine B cell lymphomas compared to normal B cells. Tumor cells from



- Introduction
- Rationale for a Canine
 Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



T-cell cNHL and healthy canine patients both showed low to negative expression of PD-1 and PD-L1. In addition, tumor infiltrating lymphocytes from both B-cell and T-cell lymphoma cells presented an increased expression of both PD-1 and PD-L1 expression compared to B and T cells from lymph nodes of healthy animals. In vitro, chemotherapy-resistant canine B-cell and T-cell lymphoma cell lines exhibited increases in both PD-1 and PD-L1 expression, compared to non-chemotherapy selected tumor cells (142). In line with this, a panel of 5 canine PD-1/PD-L1 mAbs were generated and are being studied for in vitro activity in T cell assays (143). Moreover, the immunomodulatory effects of c4G12, a canine-chimerised anti-PD-L1 mAb, were evaluated in vitro. demonstrating significantly enhanced cytokine production and proliferation of dog PBMCs. Then, a pilot clinical study was performed on seven dogs with oral malignant melanoma and two with undifferentiated sarcoma, revealing that this antibody can be a safe and effective treatment option for canine cancers (144).

Importantly, canine CTLA-4 (cytotoxic T-lymphocyte associated protein 4) has also been described and cloned (145). An agonistic recombinant canine CTLA has been efficiently used to promote tolerance in a transplant model (146), suggesting that the mechanism of action of CTLA-4 in dogs is similar to humans and that CTLA-4 checkpoint blockade could represent a novel immunotherapy for canine cancer. Importantly, Tagawa et al. (147) demonstrated an up-regulation expression of CTLA-4 on CD4+ T cells from peripheral blood obtained from dogs with B cell high grade lymphoma. CTLA-4 expression on T cells was also associated with a poor prognosis.

With the development of new checkpoint molecule targeted drugs for dogs, multiple opportunities emerge in which the dog model may provide relevant clinical information, especially regarding the rational combination of immunotherapies, including checkpoint inhibitors.

Discussion

The current landscape of cancer research is facing a profound transformation with the introduction of immune-oncology as the fourth pillar for cancer therapy. Not only have immunotherapies resulted in unprecedented clinical responses, rapid drug development and several first-in-class approvals from the FDA in the past few years, but the advent of such innovative therapies is also revolutionizing treatment paradigms and algorithms in current oncology and hemato-oncology practice (148). As a result, clinical and translational research need to adapt to a rapidly changing scenario to effectively translate novel concepts into sustainable and accessible therapeutic options for cancer patients (149). The complexities and challenges of the new era of immune-oncology strongly emphasize the need to identify new strategies, models and paths to develop fast, successful, and cost-effective therapies (13, 149). The inclusion of a canine model in the drug development path of cancer immunotherapies is being widely recognized as a valid solution to overcome several hurdles associated with conventional preclinical models (150). Dogs with naturally occurring tumors are highly translational models that represent an opportunity to investigate the clinical potential of novel immunotherapies in a comprehensive manner. By complementing murine studies and human clinical trials, dogs allow monitoring the "scaling up" effects of a therapeutic approach that depends on complex interactions between tumor and immune cells, while assessing long-term efficacy and toxicity (15). Taken together, these features may allow the establishment of solid foundations to rapidly translate the results obtained from canine patients to human patient management, with benefits for both species (151).

Importantly, the benefits of these collaborative studies can more easi-



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



lv translate into clinical success in emerging technologies, such as immune checkpoint inhibitors and CAR T cells therapy, where the rapid pace of its clinical applicability is proving critical challenges. In fact, a lot remains to be understood about patient selection, delivery, and off-target effects of emerging immunotherapy used alone or in combination. While clinicians have learned during the last decades to deal with clinical responses and toxicities related to the use of antibodies in cancer therapy, emerging therapies, such as those mentioned, are much less familiar to oncologists. Therefore, cancer research needs to develop better predictive clinical models to make these emerging immunotherapies universally available to those patients with cancer who need immune intervention in addition to other therapies (152).

However, the implementation of such canine clinical trials is far from being an easy quest. It requires multiple organized efforts to validate the canine model, which still lacks a thorough characterization of the canine immune system and its effector cells and molecules, the evaluation of common tumor epitopes, the development of canine-specific/ cross-reactive agents and the establishment of preclinical models for veterinary oncological settings (62, 153, 154). Furthermore, this also requires veterinary scientific community to join forces to implement diagnosis, staging and treatment response assessment optimization and standardization, to perform large and organized clinical trials and to achieve conformity when analyzing data (26).

Regardless of the challenges that implementing immunotherapies for cNHL lymphoma may pose, cNHL treatment is facing a paradigm shift. With several new immunotherapies emerging, it is expected that in the nearby future, immunotherapy will become a valid therapeutic tool, along with chemotherapy, radiotherapy and surgery. Furthermore, these advances also provide an integrated drug discovery platform that maximize interdisciplinary cooperation and leverage commonalities across humans and dogs, for the development of novel immunotherapies against NHL, benefiting both species.

Author Contributions

JD: writing—original draft preparation and visualization. AA: visualization and writing—review and editing. SA, SG, LT, and FA-d-S: writing—reviewing and editing. All authors contributed to the article and approved the submitted version.

Funding

This work was supported by the Portuguese Funding Agency, Fundação para a Ciência e Tecnologia, FCT IP, [SA-ICT/2017/32085 and Ph.D. fellowship SFRH/BD/131468/2017 to AA]. CIISA has provided support through Project UIDB/CVT/00276/2020, funded by FCT. Gilead Génese has provided support through Project PGG/050/2019.

Conflict of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for
 - Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



Acknowledgments

Some figures used images modified from Servier Medical Art. licensed under a Creative Common Attribution 3.0 Generic License. http://smart.servier. com/.

Abbreviations

ADCC, antibody-dependent cellular cytotoxicity; CAR, chimeric antigen receptor; CAR-T, chimeric antigen receptor T-lymphocytes; CDC, complement dependent cytotoxicity; CDV, canine distemper virus; CHOP, cyclophosphamide, doxorubicin, vincristine and prednisolone; cNHL, canine lymphoma; CTLA-4, cytotoxic T-lymphocyte associated protein 4; DLBCL, diffuse large B-cell lymphoma; EGFR, epidermal growth factor receptor; FDA, US Food and Drug Administration; GM-CSF, Granulocyte-macrophage colony-stimulating factor; HER2, human epidermal growth factor receptor 2; hNHL, human non-Hodgkin lymphoma; HSP, heat shock proteins; HSPPC, immunogenic tumor specific peptides; LMI, large multivalent immunogen; mAbs, monoclonal antibodies; NHL, non-Hodgkin lymphoma; PBMC, peripheral blood mononuclear cells; PD-1, programmed-death 1; PD-L1, PD ligand 1; scFv, single chain variable fragment; TERT, telomerase reverse transcriptase; USDA, United States Department of Agriculture.

References

- Bray F, Ferlay J, Soerjomataram I, Siegel 1. RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. (2018) 68:394-424. doi: 10.3322/ caac.21492
- 2. Fitzmaurice C. Allen C. Barber RM. Barregard L, Bhutta ZA, Brenner H, et al. Global, regional, and national cancer incidence, mortality, years of life lost, years lived with disability, and disabilityadjusted life-years for 32 cancer groups, 1990 to 2015: a systematic analysis for the global burden of disease study. JAMA Oncol. (2017) 3:524-48. doi: 10.1001/ jamaoncol.2016.5688
- 3. Torre LA, Siegel RL, Ward EM, Jemal A. Global cancer incidence and mortality rates and trends—an update. Cancer Epidemiol Prev Biomark. (2016) 25:16–27. doi: 10.1158/1055-9965.EPI-15-0578
- 4. Yang Y. Cancer immunotherapy: Harnessing the immune system to battle cancer. J Clin Invest. (2015) 125:3335-7. doi: 10.1172/JCI83871
- 5. Emens LA, Ascierto PA, Darcy PK, Demaria S, Eggermont AMM, Redmond WL, et al. Cancer immunotherapy: Opportunities and challenges in the rapidly evolving clinical landscape. Eur J Cancer. (2017) 81:116-29. doi: 10.1016/j. ejca.2017.01.035
- 6. Farkona S, Diamandis EP, Blasutig IM. Cancer immunotherapy: The beginning of the end of cancer? BMC Med. (2016) 14:73. doi: 10.1186/s12916-016-0623-5
- 7. Marks L. Engineering Health: How Biotechnology Changed Medicine. Royal Society of Chemistry (2017).

- 8. Freeman GJ. Long AJ. Iwai Y. Bourgue K. Chernova T. Nishimura H, et al. Engagement of the Pd-1 immunoinhibitory receptor by a novel B7 family member leads to negative regulation of lymphocyte activation. J Exp Med. (2000) 192:1027-34. doi: 10.1084/ iem.192.7.1027
- 9. Ishida Y, Agata Y, Shibahara K, Honjo T. Induced expression of PD-1, a novel member of the immunoglobulin gene superfamily, upon programmed cell death. EMBO J. (1992) 11:3887-95. doi: 10.1002/j.1460-2075.1992.tb05481.x
- 10. Iwai Y, Terawaki S, Honjo T. PD-1 blockade inhibits hematogenous spread of poorly immunogenic tumor cells by enhanced recruitment of effector T cells. Int Immunol. (2005) 17:133-44. doi: 10.1093/ intimm/dxh194
- 11. Kwon ED, Hurwitz AA, Foster BA, Madias C, Feldhaus AL, Greenberg NM, et al. Manipulation of T cell costimulatory and inhibitory signals for immunotherapy of prostate cancer. Proc Natl Acad Sci USA. (1997) 94:8099-103. doi: 10.1073/ pnas.94.15.8099
- 12. Leach DR. Krummel MF. Allison JP. Enhancement of antitumor immunity by CTLA-4 blockade. Science. (1996) 271:1734-6. doi: 10.1126/ science.271.5256.1734
- 13. Ventola CL. Cancer immunotherapy, Part 3: challenges and future trends. Pharm Ther. (2017) 42:514-21.
- 14. Klevorn LE, Teague RM. Adapting cancer immunotherapy models for the real world. Trends Immunol. (2016) 37:354-63. doi: 10.1016/j.it.2016.03.010



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment **Options for Non-**Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



- Park JS, Withers SS, Modiano JF, Kent MS, Chen M, Luna JI, et al. Canine cancer immunotherapy studies: linking mouse and human. J Immunother Cancer. (2016) 4:97. doi: 10.1186/s40425-016-0200-7
- Malaney P, Nicosia SV, Davé V. One mouse, one patient paradigm: new avatars of personalized cancer therapy. Cancer Lett. (2014) 344:1–12. doi: 10.1016/j. canlet.2013.10.010
- 17. Biemar F, Foti M. Global progress against cancer—challenges and opportunities. Cancer Biol Med. (2013) 10:183–6. doi: 10.7497/j.issn.2095-3941.2013.04.001
- Kola I, Landis J. Can the pharmaceutical industry reduce attrition rates? Nat Rev Drug Discov. (2004) 3:711–6. doi: 10.1038/ nrd1470
- 19. Kohnken R, Porcu P, Mishra A. Overview of the use of murine models in leukemia and lymphoma research. Front Oncol. (2017) 7:22. doi: 10.3389/fonc.2017.00022
- 20. Henry C, Bryan J. Not lost in translation: how study of diseases in our pets can benefit them and us. Mo Med. (2013) 110:216–9.
- Porrello A, Cardelli P, Spugnini EP. Oncology of companion animals as a model for humans. An overview of tumor histotypes. J Exp Clin Cancer Res CR. (2006) 25:97–105.
- 22. Gardner HL, Fenger JM, London CA. Dogs as a model for cancer. Annu Rev Anim Biosci. (2016) 4:199–222. doi: 10.1146/ annurev-animal-022114-110911
- 23. Pinho SS, Carvalho S, Cabral J, Reis CA, Gärtner F. Canine tumors: a spontaneous animal model of human carcinogenesis. Transl Res J Lab Clin Med. (2012) 159:165–72. doi: 10.1016/j. trsl.2011.11.005

- Ranieri G, Gadaleta CD, Patruno R, Zizzo N, Daidone MG, Hansson MG, et al. A model of study for human cancer: spontaneous occurring tumors in dogs. Biological features and translation for new anticancer therapies. Crit Rev Oncol Hematol. (2013) 88:187–97. doi: 10.1016/j. critrevonc.2013.03.005
- Rowell JL, McCarthy DO, Alvarez CE. Dog models of naturally occurring cancer. Trends Mol Med. (2011) 17:380–8. doi: 10.1016/j.molmed.2011.02.004
- 26. Marconato L, Polton GA, Sabattini S, Dacasto M, Garden OA, Grant I, et al. Conformity and controversies in the diagnosis, staging and follow-up evaluation of canine nodal lymphoma: A systematic review of the last 15 years of published literature. Vet Comp Oncol. (2017) 15:1029–40. doi: 10.1111/ vco.12244
- 27. Richards KL, Suter SE. Man's best friend: What can pet dogs teach us about non-Hodgkin lymphoma? Immunol Rev. (2015) 263:173–91. doi: 10.1111/imr.12238
- Marconato L, Gelain ME, Comazzi S. The dog as a possible animal model for human non-Hodgkin lymphoma: A review. Hematol Oncol. (2013) 31:1–9. doi: 10.1002/hon.2017
- 29. Ito D, Frantz AM, Modiano JF. Canine lymphoma as a comparative model for human non-Hodgkin lymphoma: recent progress and applications. Vet Immunol Immunopathol. (2014) 159:192–201. doi: 10.1016/j.vetimm.2014.02.016
- Khanna C, Lindblad-Toh K, Vail D, London C, Bergman P, Barber L, et al. The dog as a cancer model. Nat Biotechnol. (2006) 24:1065–6. doi: 10.1038/nbt0906-1065b

- Kisseberth WC, Nadella MVP, Breen M, Thomas R, Duke SE, Murahari S, et al. A novel canine lymphoma cell line: a translational and comparative model for lymphoma research. Leuk Res. (2007) 31:1709–20. doi: 10.1016/j. leukres.2007.04.003
- 32. Ponce, Marchal T, Magnol JP, Turinelli V, Ledieu D, Bonnefont C, et al. A morphological study of 608 cases of canine malignant lymphoma in France with a focus on comparative similarities between canine and human lymphoma morphology. Vet Pathol. (2010) 47:414–33. doi: 10.1177/0300985810363902
- Rütgen BC, Hammer SE, Gerner W, Christian M, de Arespacochaga AG, Willmann M, et al. Establishment and characterization of a novel canine B-cell line derived from a spontaneously occurring diffuse large cell lymphoma. Leuk Res. (2010) 34:932–8. doi: 10.1016/j. leukres.2010.01.021
- Rütgen BC, Willenbrock S, Reimann-Berg N, Walter I, Fuchs-Baumgartinger A, Wagner S, et al. Authentication of primordial characteristics of the CLBL-1 cell line prove the integrity of a canine B-cell lymphoma in a murine in vivo model. PloS ONE. (2012) 7:e40078. doi: 10.1371/journal.pone.0040078
- Hahn K, Bravo L, Adams WH, Frazier DL. Naturally occurring tumors in dogs as comparative models for cancer therapy research. Vivo Athens Greece. (1994) 8:133–43.
- Vail DM, MacEwen EG. Spontaneously occurring tumors of companion animals as models for human cancer. Cancer Invest. (2000) 18:781–92. doi: 10.3109/07357900009012210



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



23 de 72

- Hansen K, Khanna C. Spontaneous and genetically engineered animal models; use in preclinical cancer drug development. Eur J Cancer Oxf Engl. (2004) 40:858–80. doi: 10.1016/j. ejca.2003.11.031
- Seelig DM, Avery AC, Ehrhart EJ, Linden MA. The comparative diagnostic features of canine and human lymphoma. Vet Sci. (2016) 3:11. doi: 10.3390/vetsci3020011
- Swerdlow SH. WHO classification of tumours of haematopoietic and lymphoid tissues. WHO Classif Tumours. (2008) 22008:439.
- Lindblad-Toh K, Wade CM, Mikkelsen TS, Karlsson EK, Jaffe DB, Kamal M, et al. Genome sequence, comparative analysis and haplotype structure of the domestic dog. Nature. (2005) 438:803–19. doi: 10.1038/nature04338
- Pastor M, Chalvet-Monfray K, Marchal T, Keck G, Magnol JP, Fournel-Fleury C, et al. Genetic and environmental risk indicators in canine non-Hodgkin's lymphomas: breed associations and geographic distribution of 608 cases diagnosed throughout France over 1 year. J Vet Intern Med Am Coll Vet Intern Med. (2009) 23:301–10. doi: 10.1111/j.1939-1676.2008.0255.x
- 42. Gamlem H, Nordstoga K, Glattre E. Canine neoplasia–introductory paper. APMIS Suppl. (2008) 116:5–18. doi: 10.1111/j.1600-0463.2008.125m2.x
- Modiano JF, Breen M, Burnett RC, Parker HG, Inusah S, Thomas R, et al. Distinct B-cell and T-cell lymphoproliferative disease prevalence among dog breeds indicates heritable risk. Cancer Res. (2005) 65:5654–61. doi: 10.1158/0008-5472.CAN-04-4613

- 44. Paoloni M, Khanna C. Comparative oncology today. Vet Clin North Am Small Anim Pract. (2007) 37:1023–32; v. doi: 10.1016/j.cvsm.2007.08.003
- 45. Weiskopf K, Anderson KL, Ito D, Schnorr PJ, Tomiyasu H, Ring AM, et al. Eradication of canine diffuse large B-cell lymphoma in a murine xenograft model with CD47 blockade and anti-CD20. Cancer Immunol Res. (2016) 4:1072–87. doi: 10.1158/2326-6066.CIR-16-0105
- 46. Fisher SG, Fisher RI. The epidemiology of non-Hodgkin's lymphoma. Oncogene. (2004) 23:6524–34. doi: 10.1038/ sj.onc.1207843
- 47. Kong Y, Barisone GA, Sidhu RS, O'Donnell RT, Tuscano JM. Efficacy of combined histone deacetylase and checkpoint kinase inhibition in a preclinical model of human Burkitt lymphoma. Mol Med. (2015) 21:824–32. doi: 10.2119/ molmed.2015.00032
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. (2021) 71:209–49. doi: 10.3322/caac.21660
- 49. Shankland KR, Armitage JO, Hancock BW. Non-Hodgkin lymphoma. Lancet. (2012) 380:848–57. doi: 10.1016/S0140-6736(12)60605-9
- 50. Ansell SM. Non-Hodgkin lymphoma: diagnosis and treatment. Mayo Clin Proc. (2015) 90:1152–63. doi: 10.1016/j. mayocp.2015.04.025

- 51. Ito D, Brewer S, Modiano JF, Beall MJ. Development of a novel anti-canine CD20 monoclonal antibody with diagnostic and therapeutic potential. Leuk Lymph. (2015) 56:219–25. doi: 10.3109/10428194.2014.914193
- 52. Motta G, Cea M, Moran E, Carbone F, Augusti V, Patrone F, et al. Monoclonal antibodies for non-Hodgkin's lymphoma: state of the art and perspectives. Clin Dev Immunol. (2011) 2010:428253. doi: 10.1155/2010/428253
- 53. Molina A. A decade of rituximab: improving survival outcomes in non-Hodgkin's lymphoma. Annu Rev Med. (2008) 59:237–50. doi: 10.1146/annurev. med.59.060906.220345
- 54. Zappasodi R, de Braud F, Di Nicola M. Lymphoma immunotherapy: current status. Front Immunol. (2015) 6:448. doi: 10.3389/fimmu.2015.00448
- 55. Chao MP. Treatment challenges in the management of relapsed or refractory non-Hodgkin's lymphoma – novel and emerging therapies. Cancer Manag Res. (2013) 5:251–69. doi: 10.2147/CMAR. S34273
- 56. Heyman B, Yang Y. New developments in immunotherapy for lymphoma. Cancer Biol Med. (2018) 15:189–209. doi: 10.20892/j.issn.2095-3941.2018.0037
- 57. Zandvliet M. Canine lymphoma: a review. Vet Q. (2016) 36:76–104. doi: 10.1080/01652176.2016.1152633
- 58. Flory A b, Rassnick K m, Al-Sarraf R, Bailey D b., Balkman C e., Kiselow M a., et al. Combination of CCNU and DTIC chemotherapy for treatment of resistant lymphoma in dogs. J Vet Intern Med. (2008) 22:164–71. doi: 10.1111/j.1939-1676.2007.0005.x



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



- Fahey CE, Milner RJ, Barabas K, Lurie D, Kow K, Parfitt S, et al. Evaluation of the University of Florida lomustine, vincristine, procarbazine, and prednisone chemotherapy protocol for the treatment of relapsed lymphoma in dogs: 33 cases (2003–2009). J Am Vet Med Assoc. (2011) 239:209–15. doi: 10.2460/javma.239.2.209
- 60. Back AR, Schleis SE, Smrkovski OA, Lee J, Smith AN, Phillips JC. Mechlorethamine, vincristine, melphalan and prednisone (MOMP) for the treatment of relapsed lymphoma in dogs. Vet Comp Oncol. (2015) 13:398–408. doi: 10.1111/ vco.12055
- 61. Zandvliet M, Rutteman GR, Teske E. Prednisolone inclusion in a firstline multidrug cytostatic protocol for the treatment of canine lymphoma does not affect therapy results. Vet J. (2013) 197:656–61. doi: 10.1016/j. tvjl.2013.04.022
- 62. Klingemann H. Immunotherapy for dogs: running behind humans. Front Immunol. (2018) 9:133. doi: 10.3389/ fimmu.2018.00133
- 63. Anderson KL, Modiano JF. Progress in adaptive immunotherapy for cancer in companion animals: success on the path to a cure. Vet Sci. (2015) 2:363–87. doi: 10.3390/vetsci2040363
- 64. Rosales C, Jeglum KA, Obrocka M, Steplewski Z. Cytolytic activity of murine anti-dog lymphoma monoclonal antibodies with canine effector cells and complement. Cell Immunol. (1988) 115:420–8. doi: 10.1016/0008-8749(88)90194-3
- Jeglum KA. The history and future of canine lymphoma monoclonal antibody 231. Cancer Ther. (2009) 7:59–61. doi: 10.1016/s0195-5616(96)50007-0

- 66. Jeglum KA. Chemoimmunotherapy of canine lymphoma with adjuvant canine monoclonal antibody 231. Vet Clin North Am Small Anim Pract. (1996) 26:73–85. doi: 10.1016/S0195-5616(96)50007-0
- 67. Stein R, Balkman C, Chen S, Rassnick K, McEntee M, Page R, et al. Evaluation of anti-human leukocyte antigen-DR monoclonal antibody therapy in spontaneous canine lymphoma. Leuk Lymph. (2011) 52:273–84. doi: 10.3109/10428194.2010.535182
- Rue SM, Eckelman BP, Efe JA, Bloink K, Deveraux QL, Lowery D, et al. Identification of a candidate therapeutic antibody for treatment of canine B-cell lymphoma. Vet Immunol Immunopathol. (2015) 164:148– 59. doi: 10.1016/j.vetimm.2015.02.004
- 69. Jain S, Aresu L, Comazzi S, Shi J, Worrall E, Clayton J, et al. The development of a recombinant scFv monoclonal antibody targeting canine CD20 for use in comparative medicine. PloS ONE. (2016) 11:e0148366. doi: 10.1371/journal. pone.0148366
- Mizuno T, Kato Y, Kaneko MK, Sakai Y, Shiga T, Kato M, et al. Generation of a canine anti-canine CD20 antibody for canine lymphoma treatment. Sci Rep. (2020) 10:11476. doi: 10.1038/s41598-020-68470-9
- 71. O'Connor CM, Wilson-Robles H. Developing T cell cancer immunotherapy in the dog with lymphoma. ILAR J Natl Res Counc Inst Lab Anim Resour. (2014) 55:169–81. doi: 10.1093/ilar/ilu020
- 72. O'Connor CM, Sheppard S, Hartline CA, Huls H, Johnson M, Palla SL, et al. Adoptive T-cell therapy improves treatment of canine non-Hodgkin lymphoma post chemotherapy. Sci Rep. (2012) 2:249. doi: 10.1038/srep00249

- 73. Panjwani MK, Smith JB, Schutsky K, Gnanandarajah J, O'Connor CM, Powell DJ, et al. Feasibility and safety of RNAtransfected CD20-specific chimeric antigen receptor t cells in dogs with spontaneous B cell lymphoma. Mol Ther. (2016) 24:1602–14. doi: 10.1038/ mt.2016.146
- 74. Suter SE, Chein MB, von Messling V, Yip B, Cattaneo R, Vernau W, et al. In vitro canine distemper virus infection of canine lymphoid cells: a prelude to oncolytic therapy for lymphoma. Clin Cancer Res. (2005) 11:1579–87. doi: 10.1158/1078-0432.CCR-04-1944
- 75. Henson M, Suter S, Vonmessling V, Cattaneo R, Fielding A. 803. The effects of intratumoral injection of a replicating morbillivirus in a canine model of naturally occurring lymphoma. Mol Ther. (2005) 11:312. doi: 10.1016/j. ymthe.2005.07.340
- 76. Sanchez D, Pelayo R, Sarmiento RE, Medina LA, Cesarman-Maus GN, Nuñez L, et al. in vitro and in vivo oncolytic activity of Lasota strain of Newcastle disease virus on a lymphoma B-cell line and a canine cutaneous T-cell lymphoma. Blood. (2014) 124:5504. doi: 10.1182/blood. V124.21.5504.5504
- Sánchez D, Pelayo R, Medina LA, Vadillo E, Sánchez R, Núñez L, et al. Newcastle disease virus: potential therapeutic application for human and canine lymphoma. Viruses. (2015) 8:3. doi: 10.3390/v8010003
- 78. Hwang CC, Umeki S, Igase M, Coffey M, Noguchi S, Okuda M, et al. The effects of oncolytic reovirus in canine lymphoma cell lines. Vet Comp Oncol. (2016) 14(Suppl. 1):61–73. doi: 10.1111/ vco.12124



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



- 79. Hwang CC, Igase M, Sakurai M, Haraguchi T, Tani K, Itamoto K, et al. Oncolytic reovirus therapy: pilot study in dogs with spontaneously occurring tumours. Vet Comp Oncol. (2018) 16:229–38. doi: 10.1111/vco.12361
- Henson MS, Curtsinger JM, Larson VS, Klausner JS, Modiano JF, Mescher MF, et al. Immunotherapy with autologous tumour antigen-coated microbeads (large multivalent immunogen), IL-2 and GM-CSF in dogs with spontaneous B-cell lymphoma. Vet Comp Oncol. (2011) 9:95–105. doi: 10.1111/j.1476-5829.2010.00234.x
- Jeglum KA, Winters WD, Young KM. In vitro immune monitoring of antibody response in dogs given chemoimmunotherapy for lymphoma. Am J Vet Res. (1989) 50:488– 92.
- 82. Jeglum KA, Young KM, Barnsley K, Whereat A. Chemotherapy versus chemotherapy with intralymphatic tumor cell vaccine in canine lymphoma. Cancer. (1988) 61:2042–50.
- Jeglum KA, Young KM, Barnsley K, Whereat A, McGrath D, Hutson C. Intralymphatic autochthonous tumor cell vaccine in canine lymphoma. J Biol Response Mod. (1986) 5:168–75.
- 84. Sorenmo K, Krick E, Coughlin CM, Overley B, Gregor TP, Vonderheide RH, et al. CD40activated B cell cancer vaccine improves second clinical remission and survival in privately owned dogs with non-Hodgkin's lymphoma. PloS ONE. (2011) 6:e24167. doi: 10.1371/journal.pone.0024167

- Peruzzi D, Gavazza A, Mesiti G, Lubas G, Scarselli E, Conforti A, et al. A vaccine targeting telomerase enhances survival of dogs affected by B-cell lymphoma. Mol Ther J Am Soc Gene Ther. (2010) 18:1559–67. doi: 10.1038/mt.2010.104
- 86. Gavazza A, Lubas G, Fridman A, Peruzzi D, Impellizeri JA, Luberto L, et al. Safety and efficacy of a genetic vaccine targeting telomerase plus chemotherapy for the therapy of canine B-cell lymphoma. Hum Gene Ther. (2013) 24:728–38. doi: 10.1089/hum.2013.112
- Impellizeri JA, Gavazza A, Greissworth E, Crispo A, Montella M, Ciliberto G, et al. Tel-eVax: a genetic vaccine targeting telomerase for treatment of canine lymphoma. J Transl Med. (2018) 16:349. doi: 10.1186/s12967-018-1738-6
- 88. Marconato L, Frayssinet P, Rouquet N, Comazzi S, Leone VF, Laganga P, et al. Randomized, placebo-controlled, doubleblinded chemoimmunotherapy clinical trial in a pet dog model of diffuse large b-cell lymphoma. Clin Cancer Res. (2014) 20:668–77. doi: 10.1158/1078-0432.CCR-13-2283
- 89. Marconato L, Stefanello D, Sabattini S, Comazzi S, Riondato F, Laganga P, et al. Enhanced therapeutic effect of APAVAC immunotherapy in combination with dose-intense chemotherapy in dogs with advanced indolent B-cell lymphoma. Vaccine. (2015) 33:5080–6. doi: 10.1016/j. vaccine.2015.08.017

- 90. Marconato L, Aresu L, Stefanello D, Comazzi S, Martini V, Ferrari R, et al. Opportunities and challenges of active immunotherapy in dogs with B-cell lymphoma: a 5-year experience in two veterinary oncology centers. J Immunother Cancer. (2019) 7:146. doi: 10.1186/s40425-019-0624-y
- 91. Makkouk A, Weiner G. Cancer immunotherapy and breaking immune tolerance-new approaches to an old challenge. Cancer Res. (2015) 75:5–10. doi: 10.1158/0008-5472.CAN-14-2538
- 92. Vacchelli E, Pol J, Bloy N, Eggermont A, Cremer I, Fridman WH, et al. Trial watch: tumor-targeting monoclonal antibodies for oncological indications. Oncoimmunology. (2015) 4:e985940. doi: 10.4161/2162402X.2014.985940
- 93. Aguiar S, Dias J, Manuel AM, Russo R, Gois PMP, da Silva FA, et al. Chimeric small antibody fragments as strategy to deliver therapeutic payloads. Adv Protein Chem Struct Biol. (2018) 112:143–82. doi: 10.1016/bs.apcsb.2018.03.002
- 94. Kimiz-Gebologlu I, Gulce-Iz S, Biray-Avci C. Monoclonal antibodies in cancer immunotherapy. Mol Biol Rep. (2018) 45:2935–40. doi: 10.1007/s11033-018-4427-x
- 95. Chames P, Van Regenmortel M, Weiss E, Baty D. Therapeutic antibodies: successes, limitations and hopes for the future. Br J Pharmacol. (2009) 157:220–33. doi: 10.1111/j.1476-5381.2009.00190.x



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



- 96. Maloney DG, Liles TM, Czerwinski DK, Waldichuk C, Rosenberg J, Grillo-Lopez A, et al. Phase I clinical trial using escalating single-dose infusion of chimeric anti-CD20 monoclonal antibody (IDEC-C2B8) in patients with recurrent B-cell lymphoma. Blood. (1994) 84:2457–66. doi: 10.1182/ blood.V84.8.2457.2457
- Waldmann TA. Immunotherapy: past, present and future. Nat Med. (2003) 9:269–77. doi: 10.1038/nm0303-269
- Jubala CM, Wojcieszyn JW, Valli VEO, Getzy DM, Fosmire SP, Coffey D, et al. CD20 expression in normal canine B cells and in canine non-Hodgkin lymphoma. Vet Pathol. (2005) 42:468–76. doi: 10.1354/ vp.42-4-468
- 99. Kano R, Inoiue C, Okano H, Yamazaki J, Takahashi T, Watari T, et al. Canine CD20 gene. Vet Immunol Immunopathol. (2005) 108:265–8. doi: 10.1016/j. vetimm.2005.05.011
- 100. Impellizeri JA, Howell K, McKeever KP, Crow SE. The role of rituximab in the treatment of canine lymphoma: an ex vivo evaluation. Vet J Lond Engl. (2006) 171:556–8. doi: 10.1016/j.tvjl.2005.03.005
- 101. Singer J, Fazekas J, Wang W, Weichselbaumer M, Matz M, Mader A, et al. Generation of a canine anti-EGFR (ErbB-1) antibody for passive immunotherapy in dog cancer patients. Mol Cancer Ther. (2014) 13:1777–90. doi: 10.1158/1535-7163.MCT-13-0288
- 102. Killick DR, Stell AJ, Catchpole B. Immunotherapy for canine cancer–is it time to go back to the future? J Small Anim Pract. (2015) 56:229–41. doi: 10.1111/jsap.12336

- 103. Gonzales AJ, Fleck TJ, Humphrey WR, Galvan BA, Aleo MM, Mahabir SP, et al. IL-31-induced pruritus in dogs: a novel experimental model to evaluate antipruritic effects of canine therapeutics. Vet Dermatol. (2016) 27:34–e10. doi: 10.1111/ vde.12280
- 104. June CH, Sadelain M. Chimeric Antigen Receptor Therapy. N Engl J Med. (2018) 379:64–73. doi: 10.1056/NEJMra1706169
- 105. Rosenberg SA, Restifo NP, Yang JC, Morgan RA, Dudley ME. Adoptive cell transfer: a clinical path to effective cancer immunotherapy. Nat Rev Cancer. (2008) 8:299–308. doi: 10.1038/nrc2355
- 106. Addissie S, Klingemann H. Cellular immunotherapy of canine cancer. Vet Sci. (2018) 5:100. doi: 10.3390/vetsci5040100
- 107. Zheng P-P, Kros JM, Li J. Approved CAR T cell therapies: ice bucket challenges on glaring safety risks and longterm impacts. Drug Discov Today. (2018) 23:1175–82. doi: 10.1016/j. drudis.2018.02.012
- 108. Storb R, Epstein RB, Rudolph RH, Thomas ED. Allogeneic canine bone marrow transplantation following cyclophosphamide. Transplantation. (1969) 7:378. doi: 10.1097/00007890-196905000-00007
- 109. Kolb H-J. Graft-versus-leukemia effects of transplantation and donor lymphocytes. Blood. (2008) 112:4371–83. doi: 10.1182/ blood-2008-03-077974
- 110. Mata M, Vera JF, Gerken C, Rooney CM, Miller T, Pfent C, et al. Toward immunotherapy with redirected T cells in a large animal model: ex vivo activation, expansion, and genetic modification of canine T cells. J Immunother Hagerstown Md. (2014) 37:407–15. doi: 10.1097/ CJI.000000000000052

111. Mason N, Powell D, Panjwani MK, Smith J. Treatment of a Canine cd20 Positive Disease or Condition Using a Canine cd20-Specific Chimeric Antigen Receptor. W02017011316A1 (2017).

Google Scholar

- 112. Marchini A, Daeffler L, Pozdeev VI, Angelova A, Rommelaere J. Immune conversion of tumor microenvironment by oncolytic viruses: the protoparvovirus H-1PV case study. Front Immunol. (2019) 10:1848. doi: 10.3389/fimmu.2019.01848
- 113. Raja J, Ludwig JM, Gettinger SN, Schalper KA, Kim HS. Oncolytic virus immunotherapy: future prospects for oncology. J Immunother Cancer. (2018) 6:140. doi: 10.1186/s40425-018-0458-z
- 114. Conry RM, Westbrook B, McKee S, Norwood TG. Talimogene laherparepvec: first in class oncolytic virotherapy. Hum Vaccines Immunother. (2018) 14:839–46. doi: 10.1080/21645515.2017.1412896
- 115. Sánchez D, Cesarman-Maus G, Amador-Molina A, Lizano M. Oncolytic viruses for canine cancer treatment. Cancers. (2018) 10:401. doi: 10.3390/cancers10110404
- 116. Gentschev I, Patil SS, Petrov I, Cappello J, Adelfinger M, Szalay AA. Oncolytic virotherapy of canine and feline cancer. Viruses. (2014) 6:2122–37. doi: 10.3390/ v6052122
- 117. Naik S, Galyon GD, Jenks NJ, Steele MB, Miller AC, Allstadt SD, et al. Comparative oncology evaluation of intravenous recombinant oncolytic vesicular stomatitis virus therapy in spontaneous canine cancer. Mol Cancer Ther. (2018) 17:316–26. doi: 10.1158/1535-7163.MCT-17-0432



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



- 118. Gong J. Sachdev E. Mita AC. Mita MM. Clinical development of reovirus for cancer therapy: an oncolytic virus with immune-mediated antitumor activity. World J Methodol. (2016) 6:25-42. doi: 10.5662/wjm.v6.i1.25
- 119. Atzpodien J, Kirchner H. The out-patient use of recombinant human interleukin-2 and interferon alfa-2b in advanced malignancies. Eur J Cancer Oxf Engl. (1991) 27(Suppl. 4):S88-91; discussion S92. doi: 10.1016/0277-5379(91)90586-3
- 120. Burns LJ, Weisdorf DJ, DeFor TE, Repka TL, Ogle KM, Hummer C, et al. Enhancement of the anti-tumor activity of a peripheral blood progenitor cell graft by mobilization with interleukin 2 plus granulocyte colony-stimulating factor in patients with advanced breast cancer. Exp Hematol. (2000) 28:96-103. doi: 10.1016/ S0301-472X(99)00129-0
- 121. Burns LJ, Weisdorf DJ, DeFor TE, Vesole DH, Repka TL, Blazar BR, et al. IL-2based immunotherapy after autologous transplantation for lymphoma and breast cancer induces immune activation and cytokine release: a phase I/II trial. Bone Marrow Transplant. (2003) 32:177-86. doi: 10.1038/sj.bmt.1704086
- 122. Dudek AZ, Mescher MF, Okazaki I, Math VT, Luo X, Curtsinger JM, et al. Autologous large multivalent immunogen vaccine in patients with metastatic melanoma and renal cell carcinoma. Am J Clin Oncol. (2008) 31:173-81. doi: 10.1097/ COC.0b013e3181573e6b

- 123. Lissoni P. Barni S. Tancini G. Ardizzoia A. Ricci G. Aldeghi R. et al. A randomised study with subcutaneous low-dose interleukin 2 alone vs interleukin 2 plus the pineal neurohormone melatonin in advanced solid neoplasms other than renal cancer and melanoma. Br J Cancer. (1994) 69:196-9. doi: 10.1038/bic.1994.34
- 124. Miller JS, Tessmer-Tuck J, Pierson BA, Weisdorf D, McGlave P, Blazar BR, et al. Low dose subcutaneous interleukin-2 after autologous transplantation generates sustained in vivo natural killer cell activity. Biol Blood Marrow Transplant. (1997) 3:34-44.
- 125. Bendandi M, Gocke CD, Kobrin CB, Benko FA, Sternas LA, Pennington R, et al. Complete molecular remissions induced by patient-specific vaccination plus granulocyte-monocyte colony-stimulating factor against lymphoma. Nat Med. (1999) 5:1171-7. doi: 10.1038/13928
- 126. Dow S, Elmslie RE, Willson AP, Roche L, Gorman C, Potter TA. In vivo tumor transfection with superantigen plus cytokine genes induces tumor regression and prolongs survival in dogs with malignant melanoma. J Clin Invest. (1998) 101:2406–14. doi: 10.1172/JCI510
- 127. Dow S, Elmslie R, Kurzman I, MacEwen G, Pericle F, Liggitt D. Phase I study of liposome-DNA complexes encoding the interleukin-2 gene in dogs with osteosarcoma lung metastases. Hum Gene Ther. (2005) 16:937-46. doi: 10.1089/hum.2005.16.937
- 128. Khanna C. Anderson PM. Hasz DE. Katsanis E. Neville M. Klausner JS. Interleukin-2 liposome inhalation therapy is safe and effective for dogs with spontaneous pulmonary metastases. Cancer. (1997) 79:1409-21.

- 129. Moore AS. Theilen GH. Newell AD. Madewell BR. Rudolf AR. Preclinical study of sequential tumor necrosis factor and interleukin 2 in the treatment of spontaneous canine neoplasms. Cancer Res. (1991) 51:233-8.
- 130. Otter WD, Cadée J, Gavhumende R, De Groot CJ, Hennink WE, Stewart R. Effective cancer therapy with a single injection of interleukin-2 at the site of the tumour. Cancer Immunol Immunother CII. (1999) 48:419-20. doi: 10.1007/s002620050595
- 131. Thamm DH, Kurzman ID, Macewen EG, Feinmehl R, Towell TL, Longhofer SL, et al. Intralesional lipid-complexed cytokine/ superantigen immunogene therapy for spontaneous canine tumors. Cancer Immunol Immunother CII. (2003) 52:473-80. doi: 10.1007/s00262-003-0387-6
- 132. Finocchiaro LME. Fiszman GL. Karara AL, Glikin GC. Suicide gene and cytokines combined nonviral gene therapy for spontaneous canine melanoma. Cancer Gene Ther. (2008) 15:165-72. doi: 10.1038/sj.cgt.7701096
- 133. Paoloni M, Mazcko C, Selting K, Lana S, Barber L, Phillips J, et al. Defining the pharmacodynamic profile and therapeutic index of NHS-IL12 immunocytokine in dogs with malignant melanoma. PLoS ONE. (2015) 10:e0129954. doi: 10.1371/ journal.pone.0129954
- 134. Lee S-H, Shin D-J, Kim S-K. Generation of recombinant canine interleukin-15 and evaluation of its effects on the proliferation and function of canine NK cells. Vet Immunol Immunopathol. (2015) 165:1–13. doi: 10.1016/j. vetimm.2015.04.002



- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment **Options for Non-**Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



- 135. Guo C, Manjili MH, Subjeck JR, Sarkar D, Fisher PB, Wang X-Y. Therapeutic cancer vaccines: past, present and future. Adv Cancer Res. (2013) 119:421–75. doi: 10.1016/B978-0-12-407190-2.00007-1
- 136. Crow SE, Theilen GH, Benjamini E, Torten M, Henness AM, Buhles WC. Chemoimmunotherapy for canine lymphosarcoma. Cancer. (1977) 40:2102– 8.

Google Scholar

- 137. Weller RE, Theilen GH, Madewell BR, Crow SE, Benjamini E, Villalobos A. Chemoimmunotherapy for canine lymphosarcoma: a prospective evaluation of specific and nonspecific immunomodulation. Am J Vet Res. (1980) 41:516–21.
- 138. Jenkins RW, Barbie DA, Flaherty KT. Mechanisms of resistance to immune checkpoint inhibitors. Br J Cancer. (2018) 118:9–16. doi: 10.1038/bjc.2017.434
- 139. Darvin P, Toor SM, Nair VS, Elkord E. Immune checkpoint inhibitors: recent progress and potential biomarkers. Exp Mol Med. (2018) 50:1–11. doi: 10.1038/ s12276-018-0191-1
- 140. Armand P. Immune checkpoint blockade in hematologic malignancies. Blood. (2015) 125:3393–400. doi: 10.1182/ blood-2015-02-567453
- 141. Maekawa N, Konnai S, Ikebuchi R, Okagawa T, Adachi M, Takagi S, et al. Expression of PD-L1 on canine tumor cells and enhancement of ifn-γ production from tumor-infiltrating cells by PD-L1 blockade. PLoS ONE. (2014) 9:e98415. doi: 10.1371/journal.pone.0098415

- 142. Hartley G, Elmslie R, Dow S, Guth A. Checkpoint molecule expression by B and T cell lymphomas in dogs. Vet Comp Oncol. (2018) 16:352–60. doi: 10.1111/ vco.12386
- 143. Choi JW, Withers SS, Sciammas R, Rebhun RB, McSorley SJ. PD-1/PD-L1 monoclonal antibody development for canine cancer therapy. J Immunol. (2018) 200:59.28.

Google Scholar

- 144. Maekawa N, Konnai S, Takagi S, Kagawa Y, Okagawa T, Nishimori A, et al. A canine chimeric monoclonal antibody targeting PD-L1 and its clinical efficacy in canine oral malignant melanoma or undifferentiated sarcoma. Sci Rep. (2017) 7:8951. doi: 10.1038/s41598-017-09444-2
- 145. Shin I-S, Choi E-W, Chung J-Y, Hwang C-Y, Lee C-W, Youn H-Y. Cloning, expression and bioassay of canine CTLA4Ig. Vet Immunol Immunopathol. (2007) 118:12–8. doi: 10.1016/j.vetimm.2007.03.013
- 146. Graves SS, Stone D, Loretz C, Peterson L, McCune JS, Mielcarek M, et al. Establishment of long-term tolerance to SRBC in dogs by recombinant canine CTLA4-Ig. Transplantation. (2009) 88:317– 22. doi: 10.1097/TP.0b013e3181ae3285
- 147. Tagawa M, Kurashima C, Takagi S, Maekawa N, Konnai S, Shimbo G, et al. Evaluation of costimulatory molecules in dogs with B cell high grade lymphoma. PLoS ONE. (2018) 13:e0201222. doi: 10.1371/journal.pone.0201222
- 148. Kelly PN. The cancer immunotherapy revolution. Science. (2018) 359:1344–5. doi: 10.1126/science.359.6382.1344

- 149. Golan T, Milella M, Ackerstein A, Berger R. The changing face of clinical trials in the personalized medicine and immuno-oncology era: report from the international congress on clinical trials in Oncology & Hemato-Oncology (ICTO 2017). J Exp Clin Cancer Res CR. (2017) 36:192. doi: 10.1186/s13046-017-0668-0
- 150. Decker WK, da Silva RF, Sanabria MH, Angelo LS, Guimarães F, Burt BM, et al. Cancer immunotherapy: historical perspective of a clinical revolution and emerging preclinical animal models. Front Immunol. (2017) 8:829. doi: 10.3389/ fimmu.2017.00829
- 151. Barutello G, Rolih V, Arigoni M, Tarone L, Conti L, Quaglino E, et al. Strengths and weaknesses of pre-clinical models for human melanoma treatment: dawn of dogs' revolution for immunotherapy. Int J Mol Sci. (2018) 19:799. doi: 10.3390/ ijms19030799
- 152. Whiteside TL, Demaria S, Rodriguez-Ruiz ME, Zarour HM, Melero I. Emerging opportunities and challenges in cancer immunotherapy. Clin Cancer Res. (2016) 22:1845–55. doi: 10.1158/1078-0432.CCR-16-0049
- 153. Dias JNR, André AS, Aguiar SI, Ministro J, Oliveira J, Peleteiro C, et al. Establishment of a bioluminescent canine B- cell lymphoma xenograft model for monitoring tumor progression and treatment response in preclinical studies. PLoS ONE. (2018) 13:e0208147. doi: 10.1371/journal.pone.0208147



Tabla de contenido:

- Introduction
- Rationale for a Canine Model of Lymphoma
- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



29 de 72

154. Dias JNR, Lopes M, Peleteiro C, Vicente G, Nunes T, Mateus L, et al. Canine multicentric lymphoma exhibits systemic and intratumoral cytokine dysregulation. Vet Immunol Immunopathol. (2019) 218:109940. doi: 10.1016/j. vetimm.2019.109940

Citation: Dias JNR, André AS, Aguiar SI, Gil S, Tavares L and Aires-da-Silva F (2021) Immunotherapeutic Strategies for Canine Lymphoma: Changing the Odds Against Non-Hodgkin Lymphoma. Front. Vet. Sci. 8:621758. doi: 10.3389/ fvets.2021.621758 Received: 26 October 2020; Accepted: 27 July 2021; Published: 26 August 2021.

Edited by: Steven E. Suter, North Carolina State University, United States

Reviewed by: Maria Gärtner, University of Porto, Portugal Jillian M. Richmond, University of Massachusetts Medical School, United States

Copyright © 2021 Dias, André, Aguiar, Gil, Tavares and Aires-da-Silva. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

*Correspondence: Frederico Aires-da-Silva, fasilva@fmv.ulisboa.pt

Tabla de contenido:

- Introduction
- Rationale for a Canine Model of Lymphoma

PEQUEÑOS

ANIMALES

(**R**) evolution

- A Critical Unmet Need for Novel Treatment Options for Non-Hodgkin's Lymphoma in Comparative Oncology
- Current Immunotherapies for Canine Non-Hodgkin's Lymphoma
- Discussion
- Author Contributions
- Funding



Comparte tu opinión con nosotros



https://axoncomunicacion.net/estrategiasinmunoterapeuticas-para-el-linfoma-canino-cambiarlas-probabilidades-contra-el-linfoma-no-hodgkin/

Por el bienestar de tu paciente elige LASERVET

No invasivo

Sin dolor

Tiempos de recuperación reducidos o nulos

Mas información en: info@laservet-iberia.com





de 30 vídeos

Por el bienestar de tu paciente elige LASERVET

No invasivo

Sin dolor

-LASERVET EL LÁSER PARA EL VETERINARIO

Formación: Accede a más de 30 vídeos:

https:// vimeo.com/ showcase/ 9972082



Tiempos de recuperación reducidos o nulos

Mas información en: info@laservet-iberia.com



Animal models of cancer metastasis to the bone

Modelos animales de metástasis del cáncer al hueso

https://www.frontiersin.org/journals/oncology/articles/10.3389/ fonc.2023.1165380/full

Yihan Yu^{1†}, Kanglu Li^{1†}, Yizhong Peng^{1†}, Wei Wu¹, Fengxia Chen^{2*}, Zengwu Shao^{1*} and Zhicai Zhang^{1*}

1. Department of Orthopedics, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, Hubei, China

2. Department of Radiation and Medical Oncology, Zhongnan Hospital, Wuhan University, Wuhan, Hubei, China

Palabras clave:

metástasis óseas, modelos animales, cáncer de mama, cáncer de próstata, líneas celulares

Keywords: bone metastases, animal models, breast cancer, prostate cancer, cell lines



Tabla de contenido:

- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References



33 de 72

a metástasis del cáncer es una causa importante de mortalidad por varios tumores, incluidos los de mama, próstata y glándula tiroides. Dado que el tejido

óseo es uno de los sitios más comunes de metástasis. el tratamiento de las metástasis óseas es crucial para la curación del cáncer. Por lo tanto, se deben desarrollar modelos de enfermedad para comprender el proceso de metástasis ósea con el fin de diseñar terapias para ello. Se han desarrollado varios modelos traslacionales de diferentes tumores metastásicos óseos, incluidos modelos animales, modelos de invección de líneas celulares, modelos de implantes óseos y modelos de xenoinjertos derivados de pacientes. Sin embargo, actualmente no se dispone de un compendio sobre diferentes cánceres metastásicos óseos. Aquí, hemos recopilado varios modelos animales derivados de experimentos actuales sobre metástasis óseas, principalmente con cáncer de mama y próstata, para mejorar el desarrollo de modelos preclínicos y promover el tratamiento de la metástasis ósea.

ancer metastasis is a maior cause of mortality from several tumors, including those of the breast, prostate, and the thyroid gland. Since bone tissue is one of the most common sites of metastasis, the treatment of bone metastases is crucial for the cure of cancer. Hence, disease models must be developed to understand the process of bone metastasis in order to devise therapies for it. Several translational models of different bone metastatic tumors have been developed, including animal models, cell line injection models, bone implant models, and patient-derived xenograft models. However, a compendium on different bone metastatic cancers is currently not available. Here, we have compiled several animal models derived from current experiments on bone metastasis, mostly involving breast and prostate cancer, to improve the development of preclinical models and promote the treatment of bone metastasis.

Introduction

Metastasis is a frequent malignant manifestation of cancer in the mid to late stages of tumor progression. Metastasis to the bone, one of the most common sites, occurs when cancer cells migrate from the original site and invade bone tissue. It indicates adverse

prognosis, and can cause severe pain, fractures, impaired mobility, and death. The invasion of cancer cells into target sites involves several stages. Initially, they invade the surroundings of the original site, breaching the vasculature and entering the circulation. Then, depending on molecular signals on cell membranes or in their microenvironment, they invade a particular target organ along their path of circulation (1, 2). Although the precise process has not been elucidated yet, the invasion appears to last many months if not vears (3). Once a bulk of invasive cancer cells agglomerate into a mass, metastasis begins. Cancer cells modify the surrounding tissues and vasculature to favor their growth. Cancer treatment often involves a combination of radiation, chemotherapy, and medications to reduce the pain and inflammation.

Breast cancer, one of the most prevalent malignant tumors, exhibits a 40% likelihood to eventually develop bone metastases (4, 5). Bone tissue is the most common target site of breast cancer. Bone metastasis reflects potential skeletal-related events and poor clinical results. To improve the current therapies for bone-metastasized breast cancer, animal models that mimic the human tumor microenvironment have been used in preclinical experiments (6). Prostate cancer is the second most frequently occurring cancer in men.



- Tabla de contenido:
- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References



It preferentially metastasizes to the bone, and presents a worse prognosis at the metastatic stage. Rarely lethal when restricted to its primary site, the 5-year-survival rate of prostate cancer decreases by 29.8% when it metastasizes to the bone, explaining its rank as the fifth leading cause of tumor-related mortality in males (7). Antimetastatic agents need to be urgently developed and the prognosis following bone metastasis must be improved.

Multiple animal models have been used in clinical research to explore the mechanisms and prognosis of tumor metastasis. Translational models have been used to study the advanced stages of tumor metastases, reveal potential protein targets, and develop metastasis-related treatments. However, fully reproducing human bone metastases in animal models is difficult. Nevertheless, by selecting different cell lines, animal strains, and tumor transplantation methods, animal models can be constructed to answer various questions.

In this review, we have discussed the animal models of bone metastasis most commonly used in preclinical experiments and their underlying mechanisms. No single model can represent all the genetic mechanisms of bone metastasis, which requires whole-body organisms. Here, we have compiled a

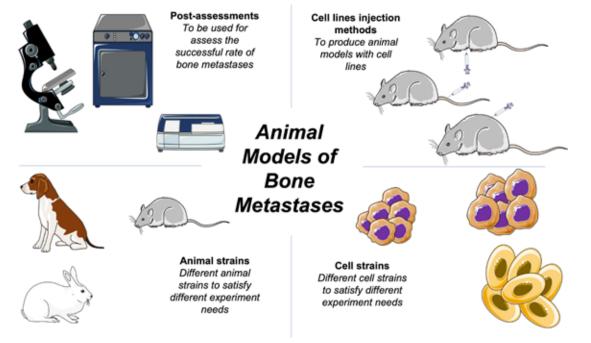


Figure 1 Schematic of basic bone metastases animal models methods.

selection of animal models to assist in future studies (**Figure 1**).

Commonly used animals in building animal models

Basing animal models of bone metastasis on general disease models is unreliable. Because the etiology of bone metastasis of human and animal cancers is different, different cancers have different metastatic targets. For example, mouse breast cancer may preferentially metastasize to the lung, while human breast cancer mainly metastasizes to the bone (2). Lung tumors may specifically metastasize to the vertebral column (8, 9). Hence, researchers are required to modify the animal models based on their experiments. The mouse is the most common animal of choice to construct bone metastasis models.

Breast cancer

Animal models based on human breast cancer cells are commonly constructed using rodents, such as mice or rats, and used in preclinical experiments (10). Both immunodeficient and immunocompetent animals are used. Nude mice of the Balb/c background are frequently used because they are



- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References



susceptible to both human and rodent breast cancer cell lines (2). Due to the lack of a thymus, immune responses are hardly generated in most of these mice following the injection of cancer cells, which significantly improves the success rate of model construction. Non-obese diabetic/severe combined immunodeficiency (NOD/SCID) mice are immunodeficient mice commonly used in xenograft experiments. Disabilities in the immune system of NOD/ SCID mice affect the growth of lymph cells as well as immune signaling. Yin's team used NOD/SCID mice paired with the MDA-MB-231 cell line to investigate how runt-related transcription factor 2, an osteogenesis-related factor, promotes breast cancer and bone metastasis (11).

The demand for crossbred or genetically engineered mice has also increased to better meet experimental needs (12–16). Mice that have been crossed and repeatedly backcrossed can offer an in vivo environment better suited to investigate the mechanism of breast cancer bone metastasis (13). In Laura's experiment, Col1a-Krm2 mice were backcrossed with NOD/SCID/IL-2rynull (NSG) mice for 10 generations to introduce an immunocompromised background (13). They found that cancer metastasis to other organs like the spine may be prevented in rather young animals. By modifying the animal mod-

el into adult mice and backcrossing over 10 generations, they could focus on the early stages of human breast cancer metastasis. Devignes' team also backcrossed Floxed mice bred in previous experiments with FVB/n wildtype mice for 10 generations to achieve genetic reconstitution consistent with experimental requirements. their Based on whether the HIF gene was expressed, mice were divided into two groups to verify whether the HIF signaling pathway in osteoblasts could promote breast cancer cell invasion and bone metastasis (14).

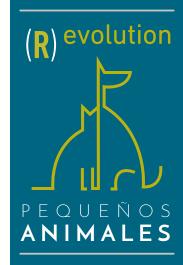
Unlike these experiments, Mercatali's team used zebrafish as a special model to study bone metastasis (17). Visualizing zebrafish embryos and easy genetic manipulation provide researchers with a new method of studying cancer progression.

Prostate cancer

The first model of prostate cancer – the Dunning rat – exhibits a spontaneous development of the disease (7). However, this model did not show a tendency for bone metastasis, and R-3327 cells derived from the Dunning rat can only metastasize to the lymph nodes. Dogs are also listed as candidate animal models, but they rarely develop prostate cancer due to the lack of androgen receptors on their cell membranes (7). The internal organization of mice femur includes a high-woven bone structure that is less fibrolamellar in nature, providing conditions amenable for bone metastasis (10, 18).

Transgenic mouse models have the advantage of lacking immune responses to injected cells or xenografts (19). Transgenic adenocarcinoma of the mouse prostate (TRAMP) is one of the most famous transgenic models, exhibiting metastases to the lung and lymph nodes rather than the bone (19, 20). The promoters expressed in neuroendocrine cells, such as the probasin promoter in TRAMP, drive transgenic oncogene expression. NOD/SCID mouse is one of the most used immunodeficient animal models in prostate cancer bone metastasis experiments (21–25). Landgraf created a new model for studying prostate cancer bone metastasis by modifying NSG mice with a humanized tissue-engineered bone construct (hTEBC), which facilitates cancer cell growth (23). Ganguly's team injected PC3 cells into the tibia of 6-week-old NSG mice to explore whether NOTCH3 induces tumor-specific elevation and secretion via MMP-3 (21).

However, the existing models are still limited to some of the detectable cancer-related factors, and cannot provide a comprehensive or linear picture of bone metastasis.



- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References



Cancer cell lines

Both patient-derived cancer tissues and immortalized cancer cell lines are used for transplantation. Patient-derived cancer tissues show genetic concordance between the clinic and the animal models, and help to establish consistent animal models specific to particular cancer cell lines. However, these models may face obstacles in the form of ethics and tissue availability. Cell lines, after several passages, can generate stable primary or secondary cancer sites. Moreover, researchers can genetically edit cell lines by using luciferase genes or knocking out certain genes (26-28).

Breast cancer

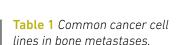
Immortalized human breast cancer cell lines, such as MDA-MB-231, 4T1, and MCF-7, are more easily available than patient-derived tissues. They possess obvious breast cancer target characteristics, and can also exhibit a tendency for bone metastasis after multiple passages (**Table 1**) (2, 5, 11, 51). They can help restore human bone metastasis in animal models. The bone-homing capabilities of MDA-MB-231 sub-lines can be enhanced via generation injections, and up to 90% of MDA-MB-231bone cells can form neoplasms (52-54). Using 5–8-week-old mice is vital to achieve bone metastasis via intracardiac, intra-arterial, or intravenous injections. Farhoodi injected 4T1 cells into

the mammary fat pad of Balb/c mice, and then examined their legs for bone metastases. Once its incidence was confirmed, the mice were sacrificed to collect the metastatic tumor cells from the leg bones. These cells were cultivated to purify tumor cells with bone-metastatic tendencies (51). They purified their experimental cells to improve the success rate.

Different pairs of cell lines can also be combined to test certain concepts. Yin's team compared MCF-7 and HCC1954 to validate whether KRT13, a protein from the keratin family, promotes stemness, metastasis, and cellular invasiveness (55). Han's group estimated the metastatic rate of different cell lines (56). They found that the proliferation of

| Cancer | Cell Lines | Origin | Model System | Metastases Preference |
|--------|---|---|--------------------------------|---|
| BCa | BCa MDA-MB- Human mammary adenocarcinoma from a 51-year-old 231 Caucasian female | | Balb/c nude, MF1 nude, NSG | Mouse long bones, spine and jaw (29-34) |
| | MCF-7 Human mammary adenocarcinoma from a 69-year-old Caucasian female | | Balb/c nude, NOD/SCID | Mouse long bones (32-34) |
| | T47D Human mammary ductal carcinoma isolated from a pleu effusion | | Balb/c nude, NOD/SCID | Mouse long bones (35, 36) |
| | 4T1 | Stage IV mammary tumor from a female Balb/c cfC3H mouse | Balb/c cfC3H | Mouse long bones, Spine, jaw, lungs, and spleen (37-40) |
| PCa | PC3 | Bone metastases from a 62-year-old white man | Balb/c nude, NOD/SCID, NSG | Mouse long bones, spine (33, 41-45) |
| | LNCaP | Supraclavicular lymph node from a 50-year-old white man | Balb/c nude, SCID | Mouse long bones, spine (29, 46-48) |
| | DU145 | Brain metastases from a 69-year-old white man | Balb/c nude, Ncr nu/nu, NOD | Mouse long bones (25, 45, 47, 49, 50) |

BCa, breast cancer; PCa, prostate cancer.





- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References

MDA-MB-453, UACC-893, and HCC-202 cells increased in the eighth week. while MDA-MB-361, UACC-812, BT-474, and ZR-75-1 cells exhibited moderate proliferation but obvious migration. Using HCC-2218 and HCC1419 cells, tumors did not form, suggesting that both lack the ability to metastasize to the bone. The tumors formed by HCC-202 and MDA-MB-361 cells decreased in size after the sixth week, indicating that these two cell lines may not survive long-term metastasis (56). Eckhardt et al. also tested several cell lines, and NSG mice were used in xenograft studies involving MDA-MB-231 and SUM159 cells (37).

Prostate cancer

Like other cancer cell lines, those of prostate cancer also originate from both humans and animals (Table 1). R-3327, derived from the Dunning rat, has been used to investigate human prostate cancer due to its spontaneous neoplasm development (57). Other animal-derived cell lines, such as PA-III or AT6-1, naturally form osteolytic and osteoblastic lesions similar to human bone metastases in animal models (57–59). RM1. derived from the mouse prostate, is a highly metastatic cell line, but does not metastasize to the bone (60). Although it can induce consistent bone lesions in mouse models, it is a transformed cell line, not a natural one.

PC3, DU145, and LNCaP are patient-derived cell lines commonly used in prostate cancer animal models. They are easily available and possess the basic prostate cancer cell targets. PC3, derived from the bone metastases of a 62-year-old white man, was selected by isolating highly invasive cells from bone metastatic lesions. Landgraf implanted an hTEBC structure based on the bone-homing properties of PC3 cells, followed by an intracardiac injection of Luc-transfected cancer cells. facilitating the construction of models for transferring the human osteoblast line PC3 to hTEBC and the murine femur (23). Studies on LNCaP, PC3, and DU145 cells. all of which differ in their sensitivity to androgens, showed that prostate cancer-secreted growth differentiation factor 15 modulates the potential for bone remodeling in metastatic bone lesions (49, 61). Lang's team grouped five common prostate cancer cell lines to verify whether PCAT7, a bone metastasis-related long non-coding RNA, activates the transforming growth factor- β /suppressor of mothers against decapentaplegic signaling pathway by upregulating transforming growth factor- β receptor 1. Its negative correlation with miR-324-5p was also investigated (62). Sohn's team tried to intracardiacally inject LNCaP cell lines grouped with CD133+. The overexpression of CD133+ in LNCaP cells enhanced their cancer

stem cell-like characteristics in terms of colony formation, migration, etc. The CD133+ group exhibited a bone metastasis rate of 80%, compared with 20% in the Vec group. Moreover, the CD133+ group showed a significant violation of the diffuse osteolytic characteristics of the spinal cord and the vertebral bodies (29).

Preparation of cell lines for transplantation

Orthotopic inoculation of cells

In situ injection of cancer cells best reproduces the process of cancer metastasis in the human body. Injected into mouse mammary fat pads, tumor cells can be seeded through the vasculature towards the target organs - a method that achieves 40-60% of bone metastases in breast cancer animal models. (63). To study the function of TIE2, a tyrosine kinase receptor, in osteolytic bone metastasis. Drescher's team administered both bilateral mammary fat pad injections and left ventricular injections to the grouped mice. The correlation between carcinoma in situ and bone metastasis was evaluated to determine whether TIE2 inhibition stimulates the dormant breast cancer cells and promotes bone metastasis (34). Likewise, Spadazzi's team injected MCF-7 cells into the left ventricle and



- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References



mammary fat pads of NSG mice to investigate whether trefoil factor-1 could exert estrogen-induced effects (64).

However, this method suffers from a considerable variation in metastatic tumor growth, besides the comorbidity caused by development of the tumor (Table 2) (73). In addition, it poses the problem of small bone metastases while the primary tumor has grown beyond an ethically reasonable size (5), which seriously compromises the detection of stimulated bone metastases.

Some scientists have also suggested subcutaneous allografts to model bone metastasis. Peiffer's team provided a detailed protocol of resecting subcutaneous prostate cancer allografts from immunocompetent mice (65). Bone metastases, abdominal cavity metastases, and local invasion all occurred in eight mice. This study demonstrated that resection of subcutaneous allografts from mice can lead to the development of metastasis; however, the duration of the experiment was extended by the removal of the prostate gland and precise operations.

Intravascular injection

Intravascular injection is a way of inoculating cells into the blood circulation. Unlike in orthotopic or ectopic inoculation, tumor cells injected via this method can localize to the target site through the intravascular circulation (Table 2) (66). Intra-arterial injections are usually administered to the left ventricle, limiting the clearance of cells that occurs when they pass through the lung capillaries (10, 53, 67). Tail vein injection, which is the more common intravenous injection today, effectively increases the rate of bone metastasis while also increasing the rate of mortality in mice (51).

Animal models currently rely on intracardiac injections to realize the pro-

cess of bone metastasis. Tumor cells are injected into the circulation through the left ventricle of mice, after which they go through the processes of adhesion, degradation, and migration to finally cause metastases in different organs, thereby simulating the process of bloodway metastasis of tumors. Using intracardiac injections to probe the role of cancer-associated factors in the regulation of tumor bone metastasis has become the preferred modeling approach (44-46). Zheng et al. used this method to prove that osteoblastic Niche-derived Jagged1 sensitizes bone metastases (15). Wang's team showed that the bone sialoprotein- $\alpha v\beta 3$ integrin axis functioned significantly more efficiently in cancer cell bone metastasis when integrin was overexpressed. For comparison, stained specimens of the brain, lung, tibia, and femur were collected after left ventricular injection in nude mice (52). Although the postop-

 Table 2 Implantation methods for

bone metastases models.



- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References

| Cell Injection Methods | Module of metastases studied | Advantages | Disadvantages |
|---------------------------|---|--|--|
| Orthotopic Inoculation | Primary tumor and invasively distant metastases | Study of tumor growth <i>in situ</i> and distant metastases | Unstable bone metastasis success rate (65-67) |
| Intracardiac | Circulation and metastases | Easily producing metastases | Requiring sophisticated skills (68-70) |
| Caudal Vessels | Circulation and metastases | More visualization of circulation inoculation | Potential lung metastases (7, 24, 51) |
| Intraosseous | Bone metastases | Most convenient and successful method for bone metastases models | Not reflecting the complete course of tumor metastasis (71) |
| Allografts/Xenografts | Depend on location | Reflecting natural heritability and cellular heterogeneity | Usually requiring immunodeficient mice and high maintenance (23, 72) |

| 1 | |
|--------------|-----|
| - በ | |
| الم | 'n |
| \mathbf{X} | -) |
| - λ | |

erative mortality is relatively high, the survival rate can still exceed 90% with practice.

Caudal vessel injection can produce a higher rate of metastasis to the leg bone than to other vital organs. This method offers better accuracy than intracardiac injection because the visibility of tail vessels enables researchers to observe the flow of cancer cell fluids within (74). Caudal vascular injections can either be intravenous or arterial. Injecting through the tail artery will reduce the elimination of tumor cells in pulmonary capillaries and improve the success rate of colonization to the bone. while tail vein injection will promote tumor metastasis to the lung (2, 51, 74). In Farhoodi's experiments, the 4T1 cell model tail artery injection mice showed a significant number of tumor cells localized to the subinguinal fat pad and the leg bone (51). Tumor cells were found in the leg bones of all 32 mice injected through the tail artery, and the rate of bone metastasis following complete tail veil injection was greater than 90% as well. Metastases were also detected in 70% of other target locations 2 weeks post-injection. Hamaidi et al. determined the effect of Lim1 on the adhesion, epithelial-mesenchymal transition, invasion, and metastatic progression of cancer cell surface targets after injection of the renal carcinoma cell line Caki2/786 through the lateral

caudal vein of nude mice (75). However, caudal vein injection also resulted in metastatic foci in the lungs of mice.

Multiple factors affect the success of experiments involving vascular injection. Operator skill gaps, standard cell operation procedures, and pressure within the caudal vessels can all influence the growth rate and success of tumor bone metastasis (51). Dilation of the caudal vessels prior to injection or the use of fluorescein to reveal vessel flow can improve the effectiveness of the injection. Non-directed intracardiac injection is still associated with a risk of thrombosis due to the procoagulant activity of tumor cells after accurate completion. The mortality of post-inoculation animal models may be reduced by injecting low-molecular weight heparin into the tail vein 10 minutes before inoculation (76).

Intraosseous injection

Metastatic tumors can bypass the pre-metastatic process if they are directly ectopically implanted into the bone. The growth of tumor cells inside the bone depends on their interaction with bone cells and the bone microenvironment (Table 2) (77, 78). Therefore, while intraosseous injection can help examine local tumor behavior within the bone microenvironment, it cannot be used to study the early stages of bone metastasis (79). Researchers

typically inject 50,000-100,000 cancer cells directly into the tibia or femurs of mice, avoiding the possible comorbidity of the animals' primary tumor (80, 81). Chen et al. observed that Brachyury, one gene affects tail length in mice, was expressed at a low level in the highly metastatic MDA-MB-231 cell line while it was highly expressed in the poorly metastatic T47D cell line when breast cancer cells were injected into the top anterior condylar region of the right tibia of mice. Nude mice showed significant swelling at the injection site 4 weeks post-injection, and X-ray revealed tumor-induced osteolytic lesions (35). After injecting prostate cancer cells into the left tibia of Balb/c nude mice. Thulin's team performed bone tumor development status assays using peripheral quantitative computed tomography (CT) and microCT to investigate the effect of signal transducer and activator of transcription 3 (STAT3) inhibitors on STAT3-regulated prostate cancer bone metastasis. The STAT3 inhibitor treatment resulted in an intact tibial bone microenvironment with no tumor formation or sclerotic response in mice, whereas the VCaP group showed sclerotic bone tumor response up to 85% (48).

Allograft and xenograft models

Transplanting allogeneic or xenogeneic tissues into animal models is a com-



- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References



mon way of modeling bone metastasis (Table 2). Since animals with different genetic backgrounds respond to allogeneic tissues differently, selecting the appropriate tissue source is especially important. In the case of xenografts, patient-derived tumor tissues can better reflect the biological characteristics of tumor bone metastasis in humans (82). Patient-derived xenografts aim to directly transplant human tumor tissue into immunodeficient mice, which represents natural heritability and cellular heterogeneity in human cancer better than simple cell-transplantation models (83). Among animal models, xenografts can only be performed in immunocompromised or immunodeficient animals. Aoki et al. first grew tumor tissue from bone metastases by intraperitoneally injecting it into male thymus-free nu/nu nude mice (42). The tumors were surgically processed to 1-mm3 fragments to be implanted into the proximal left tibia of the nude mice when they reached 10 mm in diameter. They observed tumor growth in all eight mice. Landgraf's hTEBC model is likewise based on the low immune response of NSG mice to xenografts, while adding humanized components to mimic human tumor bone metastasis as satisfyingly as possible in mice (23).

Assessment of animal models of bone metastasis

After injecting cancer cells into mice, bone lesions develop quickly, necessitating researchers to detect physiological conditions, bone changes, and tumor lesions in a timely manner.

Establishing bone metastasis models using luciferase or fluorescent protein-labeled cell lines allows researchers to monitor tumor development in the bones of living animals (15, 39-41). Oliemuller et al. studied the effects of SOX11 on cell invasion and bone metastasis using DCIS-Luc cells, generated by transducing the cells with luciferase 2 lentiviral particles (84). Arriaga's team bred NPKEYFP mice by crossing NPK mice with the Rosa-CAG-LSL-EY-FP-WPRE reporter allele, facilitating in vivo fluorescence visualization and quantification of YFP-positive prostate tumors and metastases (85).

In turn, instrumentation such as the IVIS system can provide more accurate quantitative indicators through fluorescent or bioluminescent readings obtained from tumors (76–78). Typically, tumor growth in the bone is measured once or twice a week. The area of osteolytic lesions and abnormal bone remodeling can be assessed visually by X-ray or in vivo microCT (45–47, 85). Hinz's team then used the IVIS system. After injecting MDA-MB-231 cells into the left ventricle of NSG mice, they performed IVIS bioluminescence assays weekly to assess osteolytic lesions caused by bone metastasis from triple-negative breast cancer. The inoculation of AKT3-knockout 231-B0 cells into NSG mice resulted in enhanced bone metastases (86). Another team validated the effect of intracardiacally injecting MDA-MB-231-derived osteotropic cells into nude mice by examining osteolytic lesions in their hind tibia and femurs by microCT. MicroCT images showed that NKX2-8-silenced cell lines were more likely to produce earlier bone metastases, while its overexpression delayed the appearance of metastases, inhibited osteoclast activity, and reduced bone metastatic lesions (87).

At the end of the animal test, the mice should be examined simultaneously for extraosseous metastases. All relevant organs and metastases are fixed in 10% formalin for analysis. For histological studies, samples are fixed in paraformaldehyde for 24-48 hours and then decalcified in paraformaldehvde/ethvlenediaminetetraacetic acid solution for 2 weeks. The decalcified paraffin-embedded bone should be sectioned for hematoxylin and eosin staining and evaluated using image analysis software. Bone conversion-related growth factors in the serum can also be assayed (88, 89). Metastases from the lung, liver, and brain tissue



- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References



can likewise be analyzed and studies investigating the correlation between the area and the number of bone metastases can be performed (90).

Conclusion

Bone metastasis is a common manifestation of cancer deterioration in the mid and late stages of the disease. Much research has been done on the invasion of cancer cells, from migration to the bone tissue and beyond; however, much needs to be understood yet. Animal models are vital tools in preclinical metastatic experiments that can help identify the key steps in bone metastasis. Here, we have summarized the experimental animals, cell lines, cell implantation techniques, and evaluation methods used while studying common breast and prostate cancer bone metastases. For preclinical animal testing, immunodeficient animals are used to achieve xenograft growth without eliciting a host immune response. In preclinical studies, many investigators have successfully improved the success of tumor cell colonization to the bone by backcrossing cell lines and transgenic mice. More importantly, most animal tests related to cancer bone metastasis have been performed using cancer cell line injection models. Although the early stages of bone metastasis cannot be studied, these models are effective

for studying the interaction between cancer cells and the bone microenvironment.

However, using mice to study human tumor immunity has its limitations. The differences in bone metastasis pathways between humans and animal models can explain why the success of preclinical treatments is not perfectly reproduced in humans. The inability to present a complete and comprehensive picture of the whole process of bone metastasis is also a problem that needs to be addressed while engineering animal models today.

Author contributions

All authors contributed equally to this work. All authors contributed to the article and approved the submitted version.

Funding

National Natural Science Foundation of China.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Zhang L, Gaskins K, Yu Z, Xiong Y, Merino MJ, Kebebew E. An in vivo mouse model of metastatic human thyroid cancer. Thyroid Off J Am Thyroid Assoc (2014) 24(4):695– 704. doi: 10.1089/thy.2013.0149
- 2. Tulotta C, Groenewoud A, Snaar-Jagalska BE, Ottewell P. Animal models of breast cancer bone metastasis. Methods Mol Biol Clifton NJ (2019) 1914:309–30. doi: 10.1007/978-1-4939-8997-3_17
- Gawrzak S, Rinaldi L, Gregorio S, Arenas EJ, Salvador F, Urosevic J, et al. MSK1 regulates luminal cell differentiation and metastatic dormancy in ER+ breast cancer. Nat Cell Biol (2018) 20(2):211–21. doi: 10.1038/s41556-017-0021-z



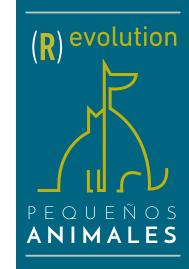
- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References



- Horas K, Zheng Y, Zhou H, Seibel MJ. Animal models for breast cancer metastasis to bone: Opportunities and limitations. Cancer Invest. (2015) 33(9):459–68. doi: 10.3109/07357907.2015.1065500
- 6. Deasy SK, Erez N. A glitch in the matrix: Organ-specific matrisomes in metastatic niches. Trends Cell Biol (2022) 32(2):110– 23. doi: 10.1016/j.tcb.2021.08.001
- Berish RB, Ali AN, Telmer PG, Ronald JA, Leong HS. Translational models of prostate cancer bone metastasis. Nat Rev Urol. (2018) 15(7):403–21. doi: 10.1038/ s41585-018-0020-2
- Liang H, Zhou L, Hu Z, Ge Y, Zhang T, Chen Q, et al. Siglec15 checkpoint blockade for simultaneous immunochemotherapy and osteolysis inhibition in lung adenocarcinoma spinal metastasis via a hollow nanoplatform. Small Weinh Bergstr Ger (2022) 18(29):e2107787. doi: 10.1002/ smll.202107787
- Wang K, Jiang L, Hu A, Sun C, Zhou L, Huang Y, et al. Vertebral-specific activation of the CX3CL1/ICAM-1 signaling network mediates non-small-cell lung cancer spinal metastasis by engaging tumor cell-vertebral bone marrow endothelial cell interactions. Theranostics (2021) 11(10):4770–89. doi: 10.7150/thno.54235
- Haq M, Goltzman D, Tremblay G, Brodt P. Rat prostate adenocarcinoma cells disseminate to bone and adhere preferentially to bone marrow-derived endothelial cells. Cancer Res (1992) 52(17):4613–9. doi: 10.1038/s41467-022-30105-0

- 11. Yin X, Teng X, Ma T, Yang T, Zhang J, Huo M, et al. RUNX2 recruits the NuRD(MTA1)/ CRL4B complex to promote breast cancer progression and bone metastasis. Cell Death Differ (2022) 29(11):2203–17. doi: 10.1038/s41418-022-01010-2
- 12. Zuo H, Yang D, Wan Y. Fam20C regulates bone resorption and breast cancer bone metastasis through osteopontin and BMP4. Cancer Res (2021) 81(20):5242–54. doi: 10.1158/0008-5472.CAN-20-3328
- Brylka L, Jähn-Rickert K, Baranowsky A, Neven M, Horn M, Yorgan T, et al. Spine metastases in immunocompromised mice after intracardiac injection of MDA-MB-231-SCP2 breast cancer cells. Cancers (2022) 14(3):556. doi: 10.3390/ cancers14030556
- Devignes CS, Aslan Y, Brenot A, Devillers A, Schepers K, Fabre S, et al. HIF signaling in osteoblast-lineage cells promotes systemic breast cancer growth and metastasis in mice. Proc Natl Acad Sci USA (2018) 115(5):E992–1001. doi: 10.1073/pnas.1718009115
- Zheng H, Bae Y, Kasimir-Bauer S, Tang R, Chen J, Ren G, et al. Therapeutic antibody targeting tumor- and osteoblastic niche-derived Jagged1 sensitizes bone metastasis to chemotherapy. Cancer Cell (2017) 32(6):731–747.e6. doi: 10.1016/j. ccell.2017.11.002
- 16. Ouarné M, Bouvard C, Boneva G, Mallet C, Ribeiro J, Desroches-Castan A, et al. BMP9, but not BMP10, acts as a quiescence factor on tumor growth, vessel normalization and metastasis in a mouse model of breast cancer. J Exp Clin Cancer Res CR. (2018) 37:209. doi: 10.1186/s13046-018-0885-1

- 17. Mercatali L, La Manna F, Groenewoud A, Casadei R, Recine F, Miserocchi G, et al. Development of a patient-derived xenograft (PDX) of breast cancer bone metastasis in a zebrafish model. Int J Mol Sci (2016) 17(8):1375. doi: 10.3390/ ijms17081375
- Kerschnitzki M, Wagermaier W, Roschger P, Seto J, Shahar R, Duda GN, et al. The organization of the osteocyte network mirrors the extracellular matrix orientation in bone. J Struct Biol (2011) 173(2):303–11. doi: 10.1016/j. jsb.2010.11.014
- 19. Kang J, La Manna F, Bonollo F, Sampson N, Alberts IL, Mingels C, et al. Tumor microenvironment mechanisms and bone metastatic disease progression of prostate cancer. Cancer Lett (2022) 530:156–69. doi: 10.1016/j.canlet.2022.01.015
- Gingrich JR, Barrios RJ, Morton RA, Boyce BF, DeMayo FJ, Finegold MJ, et al. Metastatic prostate cancer in a transgenic mouse. Cancer Res (1996) 56(18):4096– 102.
- 21. Ganguly SS, Hostetter G, Tang L, Frank SB, Saboda K, Mehra R, et al. Notch3 promotes prostate cancer-induced bone lesion development via MMP-3. Oncogene (2020) 39(1):204–18. doi: 10.1038/s41388-019-0977-1
- 22. Polavaram NS, Dutta S, Islam R, Bag AK, Roy S, Poitz D, et al. Tumor- and osteoclast-derived NRP2 in prostate cancer bone metastases. Bone Res (2021) 9(1):24. doi: 10.1038/s41413-021-00136-2



- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References



43 de 72

- 23. Landgraf M, Lahr CA, Sanchez-Herrero A, Meinert C, Shokoohmand A, Pollock PM, et al. Humanized bone facilitates prostate cancer metastasis and recapitulates therapeutic effects of zoledronic acid in vivo. Bone Res (2019) 7:31. doi: 10.1038/ s41413-019-0072-9
- 24. Hu CY, Chen J, Qin XH, You P, Ma J, Zhang J, et al. Long non-coding RNA NORAD promotes the prostate cancer cell extracellular vesicle release via microRNA-541-3p-regulated PKM2 to induce bone metastasis of prostate cancer. J Exp Clin Cancer Res CR. (2021) 40(1):98. doi: 10.1186/s13046-021-01891-0
- Li L, Ameri AH, Wang S, Jansson KH, Casey OM, Yang Q, et al. EGR1 regulates angiogenic and osteoclastogenic factors in prostate cancer and promotes metastasis. Oncogene (2019) 38(35):6241–55. doi: 10.1038/s41388-019-0873-8
- 26. Luo A, Xu Y, Li S, Bao J, Lü J, Ding N, et al. Cancer stem cell property and gene signature in bone-metastatic breast cancer cells. Int J Biol Sci (2020) 16(14):2580–94. doi: 10.7150/ijbs.45693
- 27. Jenkins DE, Hornig YS, Oei Y, Dusich J, Purchio T. Bioluminescent human breast cancer cell lines that permit rapid and sensitive in vivo detection of mammary tumors and multiple metastases in immune deficient mice. Breast Cancer Res BCR (2005) 7(4):R444–454. doi: 10.1186/ bcr1026
- Sawada Y, Kikugawa T, Iio H, Sakakibara I, Yoshida S, Ikedo A, et al. GPRC5A facilitates cell proliferation through cell cycle regulation and correlates with bone metastasis in prostate cancer. Int J Cancer (2020) 146(5):1369–82. doi: 10.1002/ ijc.32554

- 29. Sohn HM, Kim B, Park M, Ko YJ, Moon YH, Sun JM, et al. Effect of CD133 overexpression on bone metastasis in prostate cancer cell line LNCaP. Oncol Lett (2019) 18(2):1189–98. doi: 10.3892/ ol.2019.10443
- 30. Sung NJ, Kim NH, Surh YJ, Park SA. Gremlin-1 promotes metastasis of breast cancer cells by activating STAT3-MMP13 signaling pathway. Int J Mol Sci (2020) 21(23):9227. doi: 10.3390/ijms21239227
- Marino S, de Ridder D, Bishop RT, Renema N, Ponzetti M, Sophocleous A, et al. Paradoxical effects of JZL184, an inhibitor of monoacylglycerol lipase, on bone remodelling in healthy and cancer-bearing mice. EBioMedicine (2019) 44:452–66. doi: 10.1016/j.ebiom.2019.05.048
- 32. Zhang Z, Xu Q, Song C, Mi B, Zhang H, Kang H, et al. Serum- and glucocorticoidinducible kinase 1 is essential for osteoclastogenesis and promotes breast cancer bone metastasis. Mol Cancer Ther (2020) 19(2):650–60. doi: 10.1158/1535-7163.MCT-18-0783
- 33. Kim B, Kim H, Jung S, Moon A, Noh DY, Lee ZH, et al. A CTGF-RUNX2-RANKL axis in breast and prostate cancer cells promotes tumor progression in bone. J Bone Miner Res (2020) 35(1):155–66. doi: 10.1002/ jbmr.3869
- 34. Drescher F, Juárez P, Arellano DL, Serafín-Higuera N, Olvera-Rodriguez F, Jiménez S, et al. TIE2 induces breast cancer cell dormancy and inhibits the development of osteolytic bone metastases. Cancers (2020) 12(4):868. doi: 10.3390/ cancers12040868

- 35. Chen M, Zou S, He C, Zhou J, Li S, Shen M, et al. Transactivation of SOX5 by brachyury promotes breast cancer bone metastasis. Carcinogenesis (2020) 41(5):551–60. doi: 10.1093/carcin/bgz142
- 36. Zhao C, Cai X, Wang Y, Wang D, Wang T, Gong H, et al. NAT1 promotes osteolytic metastasis in luminal breast cancer by regulating the bone metastatic niche via NF- B/IL-1B signaling pathway. Am J Cancer Res (2020) 10(8):2464–79.
- Eckhardt BL, Cao Y, Redfern AD, Chi LH, Burrows AD, Roslan S, et al. Activation of canonical BMP4-SMAD7 signaling suppresses breast cancer metastasis. Cancer Res (2020) 80(6):1304–15. doi: 10.1158/0008-5472.CAN-19-0743
- 38. Zhang D, Iwabuchi S, Baba T, Hashimoto S, Mukaida N, Sasaki S. Involvement of a transcription factor, Nfe2, in breast cancer metastasis to bone. Cancers (2020) 12(10):3003. doi: 10.3390/ cancers12103003
- 39. Sasaki S, Zhang D, Iwabuchi S, Tanabe Y, Hashimoto S, Yamauchi A, et al. Crucial contribution of GPR56/ADGRG1, expressed by breast cancer cells, to bone metastasis formation. Cancer Sci (2021) 112(12):4883–93. doi: 10.1111/cas.15150
- 40. Sun J, Huang J, Lan J, Zhou K, Gao Y, Song Z, et al. Overexpression of CENPF correlates with poor prognosis and tumor bone metastasis in breast cancer. Cancer Cell Int (2019) 19:264. doi: 10.1186/ s12935-019-0986-8
- 41. Connelly ZM, Jin R, Zhang J, Yang S, Cheng S, Shi M, et al. FOXA2 promotes prostate cancer growth in the bone. Am J Transl Res (2020) 12(9):5619–29. doi: 10.1038/ s41413-021-00178-6



- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References



- 42. Aoki Y, Masaki N, Tome Y, Kubota Y, Aoki Y, Bouvet M, et al. Non-invasively imageable tibia-tumor-fragment implantation experimental-bone-metastasis mouse model of GFP-expressing prostate cancer. Vivo Athens Greece (2022) 36(4):1647–50. doi: 10.21873/invivo.12876
- 43. Shin SH, Kim I, Lee JE, Lee M, Park JW. Loss of EGR3 is an independent risk factor for metastatic progression in prostate cancer. Oncogene (2020) 39(36):5839–54. doi: 10.1038/s41388-020-01418-5
- 44. Huang S, Wa Q, Pan J, Peng X, Ren D, Li Q, et al. Transcriptional downregulation of miR-133b by REST promotes prostate cancer metastasis to bone via activating TGF- β signaling. Cell Death Dis (2018) 9(7):779. doi: 10.1038/s41419-018-0807-3
- 45. Meng X, Vander Ark A, Daft P, Woodford E, Wang J, Madaj Z, et al. Loss of TGF-β signaling in osteoblasts increases basic-FGF and promotes prostate cancer bone metastasis. Cancer Lett (2018) 418:109– 18. doi: 10.1016/j.canlet.2018.01.018
- 46. Li Q, Wang M, Hu Y, Zhao E, Li J, Ren L, et al. MYBL2 disrupts the hippo-YAP pathway and confers castration resistance and metastatic potential in prostate cancer. Theranostics (2021) 11(12):5794–812. doi: 10.7150/thno.56604
- 47. Zhao Z, Li E, Luo L, Zhao S, Liu L, Wang J, et al. A PSCA/PGRN-NF-κB-Integrin-α4 axis promotes prostate cancer cell adhesion to bone marrow endothelium and enhances metastatic potential. Mol Cancer Res MCR (2020) 18(3):501–13. doi: 10.1158/1541-7786.MCR-19-0278

- 48. Thulin MH, Määttä J, Linder A, Sterbova S, Ohlsson C, Damber JE, et al. Inhibition of STAT3 prevents bone metastatic progression of prostate cancer in vivo. Prostate (2021) 81(8):452–62. doi: 10.1002/pros.24125
- 49. Siddiqui JA, Seshacharyulu P, Muniyan S, Pothuraju R, Khan P, Vengoji R, et al. GDF15 promotes prostate cancer bone metastasis and colonization through osteoblastic CCL2 and RANKL activation. Bone Res (2022) 10(1):6. doi: 10.1038/s41413-021-00178-6
- 50. Zhang B, Li Y, Wu Q, Xie L, Barwick B, Fu C, et al. Acetylation of KLF5 maintains EMT and tumorigenicity to cause chemoresistant bone metastasis in prostate cancer. Nat Commun (2021) 12(1):1714. doi: 10.1038/s41467-021-21976-w
- 51. Farhoodi HP, Segaliny AI, Wagoner ZW, Cheng JL, Liu L, Zhao W. Optimization of a syngeneic murine model of bone metastasis. J Bone Oncol (2020) 23:100298. doi: 10.1016/j.jbo.2020.100298
- 52. Wang L, Song L, Li J, Wang Y, Yang C, Kou X, et al. Bone sialoprotein- $\alpha v\beta 3$ integrin axis promotes breast cancer metastasis to the bone. Cancer Sci (2019) 110(10):3157–72. doi: 10.1111/cas.14172
- 53. Wetterwald A, van der Pluijm G, Que I, Sijmons B, Buijs J, Karperien M, et al. Optical imaging of cancer metastasis to bone marrow: A mouse model of minimal residual disease. Am J Pathol (2002) 160(3):1143–53. doi: 10.1016/S0002-9440(10)64934-6

- 54. Nutter F, Holen I, Brown HK, Cross SS, Evans CA, Walker M, et al. Different molecular profiles are associated with breast cancer cell homing compared with colonisation of bone: Evidence using a novel bone-seeking cell line. Endocr Relat Cancer (2014) 21(2):327–41. doi: 10.1530/ ERC-13-0158
- 55. Yin L, Li Q, Mrdenovic S, Chu GCY, Wu BJ, Bu H, et al. KRT13 promotes stemness and drives metastasis in breast cancer through a plakoglobin/c-myc signaling pathway. Breast Cancer Res BCR (2022) 24(1):7. doi: 10.1186/s13058-022-01502-6
- 56. Han Y, Azuma K, Watanabe S, Semba K, Nakayama J. Metastatic profiling of HER2-positive breast cancer cell lines in xenograft models. Clin Exp Metastasis (2022) 39(3):467–77. doi: 10.1007/s10585-022-10150-1
- 57. Liepe K, Geidel H, Haase M, Hakenberg OW, Runge R, Kotzerke J. New model for the induction of osteoblastic bone metastases in rat. Anticancer Res (2005) 25(2A):1067– 73.
- 58. Blouin S, Baslé MF, Chappard D. Rat models of bone metastases. Clin Exp Metastasis (2005) 22(8):605–14. doi: 10.1007/s10585-006-9002-5
- 59. Lamoureux F, Ory B, Battaglia S, Pilet P, Heymann MF, Gouin F, et al. Relevance of a new rat model of osteoblastic metastases from prostate carcinoma for preclinical studies using zoledronic acid. Int J Cancer (2008) 122(4):751–60. doi: 10.1002/ ijc.23187
- 60. Power CA, Pwint H, Chan J, Cho J, Yu Y, Walsh W, et al. A novel model of bone-metastatic prostate cancer in immunocompetent mice. Prostate (2009) 69(15):1613–23. doi: 10.1002/pros.21010



- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References



- 61. Wang W, Yang X, Dai J, Lu Y, Zhang J, Keller ET. Prostate cancer promotes a vicious cycle of bone metastasis progression through inducing osteocytes to secrete GDF15 that stimulates prostate cancer growth and invasion. Oncogene (2019) 38(23):4540–59. doi: 10.1038/s41388-019-0736-3
- 62. Lang C, Dai Y, Wu Z, Yang Q, He S, Zhang X, et al. SMAD3/SP1 complex-mediated constitutive active loop between lncRNA PCAT7 and TGF-β signaling promotes prostate cancer bone metastasis. Mol Oncol (2020) 14(4):808–28. doi: 10.1002/1878-0261.12634
- 63. Lee JH, Kim B, Jin WJ, Kim JW, Kim HH, Ha H, et al. Trolox inhibits osteolytic bone metastasis of breast cancer through both PGE2-dependent and independent mechanisms. Biochem Pharmacol (2014) 91(1):51–60. doi: 10.1016/j. bcp.2014.06.005
- 64. Spadazzi C, Mercatali L, Esposito M, Wei Y, Liverani C, De Vita A, et al. Trefoil factor-1 upregulation in estrogen-receptor positive breast cancer correlates with an increased risk of bone metastasis. Bone (2021) 144:115775. doi: 10.1016/j. bone.2020.115775
- 65. Peiffer LB, Hicks J, Sosa RY, De Marzo AM, Sfanos KS, Maynard JP. Modeling human prostate cancer metastasis in mice via resection of subcutaneous allografts. Front Oncol (2022) 12:877536. doi: 10.3389/ fonc.2022.877536
- 66. Yip RKH, Rimes JS, Capaldo BD, Vaillant F, Mouchemore KA, Pal B, et al. Mammary tumour cells remodel the bone marrow vascular microenvironment to support metastasis. Nat Commun (2021) 12(1):6920. doi: 10.1038/s41467-021-26556-6

- 67. Chang J, Sun X, Ma X, Zhao P, Shi B, Wang Y, et al. Intra-cardiac injection of human prostate cancer cells to create a bone metastasis xenograft mouse model. J Vis Exp JoVE (2022) 189). doi: 10.3791/64589
- 68. Chen M, Wu C, Fu Z, Liu S. ICAM1 promotes bone metastasis via integrin-mediated TGF-β/EMT signaling in triple-negative breast cancer. Cancer Sci (2022) 113(11):3751–65. doi: 10.1111/cas.15532
- 69. Labanca E, Yang J, Shepherd PDA, Wan X, Starbuck MW, Guerra LD, et al. Fibroblast growth factor receptor 1 drives the metastatic progression of prostate cancer. Eur Urol Oncol (2022) 5(2):164–75. doi: 10.1016/j.euo.2021.10.001
- 70. Capietto AH, Lee S, Clever D, Eul E, Ellis H, Ma CX, et al. Effective treatment of established bone metastases can be achieved by combinatorial osteoclast blockade and depletion of granulocytic subsets. Cancer Immunol Res (2021) 9(12):1400–12. doi: 10.1158/2326-6066. CIR-21-0232
- Engelmann J, Zarrer J, Gensch V, Riecken K, Berenbrok N, Luu TV, et al. Regulation of bone homeostasis by MERTK and TYRO3. Nat Commun (2022) 13(1):7689. doi: 10.1038/s41467-022-33938-x
- 72. Boudreau MW, Duraki D, Wang L, Mao C, Kim JE, Henn MA, et al. A small-molecule activator of the unfolded protein response eradicates human breast tumors in mice. Sci Transl Med (2021) 13(603):eabf1383. doi: 10.1126/scitranslmed.abf1383
- 73. Bibby MC. Orthotopic models of cancer for preclinical drug evaluation: Advantages and disadvantages. Eur J Cancer Oxf Engl 1990 (2004) 40(6):852–7. doi: 10.1186/ s13046-018-0813-4

- 74. Kuchimaru T, Kataoka N, Nakagawa K, Isozaki T, Miyabara H, Minegishi M, et al. A reliable murine model of bone metastasis by injecting cancer cells through caudal arteries. Nat Commun (2018) 9:2981. doi: 10.1038/s41467-018-05366-3
- 75. Hamaidi I, Coquard C, Danilin S, Dormoy V, Béraud C, Rothhut S, et al. The Lim1 oncogene as a new therapeutic target for metastatic human renal cell carcinoma. Oncogene (2019) 38(1):60–72. doi: 10.1038/s41388-018-0413-y
- 76. Stocking KL, Jones JC, Everds NE, Buetow BS, Roudier MP, Miller RE. Use of lowmolecular-weight heparin to decrease mortality in mice after intracardiac injection of tumor cells. Comp Med (2009) 59(1):37–45.
- 77. Robinson BD, Sica GL, Liu YF, Rohan TE, Gertler FB, Condeelis JS, et al. Tumor microenvironment of metastasis in human breast carcinoma: A potential prognostic marker linked to hematogenous dissemination. Clin Cancer Res Off J Am Assoc Cancer Res (2009) 15(7):2433–41. doi: 10.1158/1078-0432.CCR-08-2179
- Coleman RE, Gregory W, Marshall H, Wilson C, Holen I. The metastatic microenvironment of breast cancer: Clinical implications. Breast Edinb Scotl (2013) 22 Suppl 2:S50–56. doi: 10.1016/j. breast.2013.07.010
- 79. Halpern J, Lynch CC, Fleming J, Hamming D, Martin MD, Schwartz HS, et al. The application of a murine bone bioreactor as a model of tumor: bone interaction. Clin Exp Metastasis (2006) 23(7–8):345–56. doi: 10.1007/s10585-006-9044-8



- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References



46 de 72

- Zhu W, Sheng D, Shao Y, Zhang Q, Peng Y. Neuronal calcitonin gene-related peptide promotes prostate tumor growth in the bone microenvironment. Peptides (2021) 135:170423. doi: 10.1016/j. peptides.2020.170423
- Zheng Y, Seibel MJ, Zhou H. Methods in bone biology: Cancer and bone. In: Duque G, Watanabe K, editors. Osteoporosis Res: Animal models. London: Springer London (2011). p. 83–91. doi: 10.1007/978-0-85729-293-3_7
- 82. Holzapfel BM, Thibaudeau L, Hesami P, Taubenberger A, Holzapfel NP, Mayer-Wagner S, et al. Humanised xenograft models of bone metastasis revisited: Novel insights into species-specific mechanisms of cancer cell osteotropism. Cancer Metastasis Rev (2013) 32(1–2):129–45. doi: 10.1007/s10555-013-9437-5
- Lin D, Xue H, Wang Y, Wu R, Watahiki A, Dong X, et al. Next generation patientderived prostate cancer xenograft models. Asian J Androl. (2014) 16(3):407–12. doi: 10.4103/1008-682X.125394
- 84. Oliemuller E, Newman R, Tsang SM, Foo S, Muirhead G, Noor F, et al. SOX11 promotes epithelial/mesenchymal hybrid state and alters tropism of invasive breast cancer cells. eLife (2020) 9. doi: 10.7554/ eLife.58374
- 85. Arriaga JM, Panja S, Alshalalfa M, Zhao J, Zou M, Giacobbe A, et al. A MYC and RAS co-activation signature in localized prostate cancer drives bone metastasis and castration resistance. Nat Cancer (2020) 11):1082–96. doi: 10.1038/s43018-020-00125-0

- 86. Hinz N, Baranowsky A, Horn M, Kriegs M, Sibbertsen F, Smit DJ, et al. Knockdown of AKT3 activates HER2 and DDR kinases in bone-seeking breast cancer cells, promotes metastasis In vivo and attenuates the TGF β /CTGF axis. Cells (2021) 10(2):430. doi: 10.3390/ cells10020430
- Abudourousuli A, Chen S, Hu Y, Qian W, Liao X, Xu Y, et al. NKX2-8/PTHrP axis-mediated osteoclastogenesis and bone metastasis in breast cancer. Front Oncol (2022) 12:907000. doi: 10.3389/fonc.2022.907000
- 88. Maimon A, Levi-Yahid V, Ben-Meir K, Halpern A, Talmi Z, Priya S, et al. Myeloid cell-derived PROS1 inhibits tumor metastasis by regulating inflammatory and immune responses via IL-10. J Clin Invest (2021) 131(10):e126089. doi: 10.1172/JCI126089
- 89. Cai WL, Huang WD, Li B, Chen TR, Li ZX, Zhao CL, et al. microRNA-124 inhibits bone metastasis of breast cancer by repressing interleukin-11. Mol Cancer (2018) 17(1):9. doi: 10.1186/s12943-017-0746-0
- 90. He Y, Luo W, Liu Y, Wang Y, Ma C, Wu Q, et al. IL-20RB mediates tumoral response to osteoclastic niches and promotes bone metastasis of lung cancer. J Clin Invest (2022) 132(20):e157917. doi: 10.1172/ JCI157917

Keywords: bone metastases, animal models, breast cancer, prostate cancer, cell lines

Citation: Yu Y, Li K, Peng Y, Wu W, Chen F, Shao Z and Zhang Z (2023) Animal models of cancer metastasis to the bone. Front. Oncol. 13:1165380. doi: 10.3389/fonc.2023.1165380 Received: 14 February 2023; Accepted: 10 March 2023; Published: 05 April 2023.

Edited by:Wenwen Zhang, Nanjing Medical University, China

Reviewed by:Qiuyu Liu, Henan Provincial People's Hospital, ChinaYanping Yang, Shanghai University of Traditional Chinese Medicine, China

Copyright © 2023 Yu, Li, Peng, Wu, Chen, Shao and Zhang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

*Correspondence: Fengxia Chen, fengxiachen@whu.edu.cn; Zengwu Shao, 1985XH0536@hust.edu.cn; Zhicai Zhang, zhicaizhang@126.com

†These authors have contributed equally to this work

This article is part of the Research Top-ic

Diagnosis and Treatment of Bone Metastases



- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References





Comparte tu opinión con nosotros



https://axoncomunicacion.net/modelos-animales-demetastasis-del-cancer-al-hueso/





- Introduction
- Commonly used animals in building animal models
- Cancer cell lines
- Preparation of cell lines for transplantation
- Assessment of animal models of bone metastasis
- Conclusion
- Author contributions
- Funding
- Conflict of interest
- Publisher's note
- References





ROYAL CANIN® Urinary es una gama diseñada para el manejo nutricional de los problemas del tracto urinario inferior

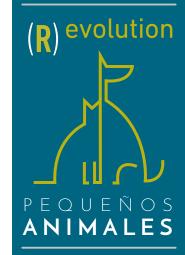
Incluye dietas con enfoques nutricionales diferentes en función del tipo de cálculo.

La presentación húmeda es especialmente interesante, ya que su alto contenido de humedad favorece la dilución urinaria y es la primera elección en caso de cistitis idiopática felina.

Gemma Baciero

Veterinaria, Acre. GTNC AVEPA. Comunicación Científica Royal Canin.







¿Cómo definirías la gama Urinary de Royal Canin?

ROYAL CANIN[®] Urinary es una gama formulada específicamente para el manejo dietético de las patologías de tracto urinario inferior en el gato y el perro, como urolitiasis y cistitis.

Incluye dietas con enfoques nutricionales diferentes en función del tipo de cálculo. Por un lado, para estruvita y oxalato, las urolitiasis más comunes tanto para gatos como perros, están las dietas Urinary S/O que provocan una baja sobresaturación relativa (SSR) para ambos, y que se caracterizan por su capacidad de disolver la estruvita, su efecto acidificante de la orina y su aporte controlado de los precursores como el magnesio. Al mismo tiempo, favorece la dilución urinaria y es indicado para ayudar a reducir la reaparición de los cálculos de oxalato, que no se pueden disolver.

Por otro lado, está la dieta Urinary UC, con un enfoque totalmente diferente, alcalinizante, con un contenido controlado de proteína y seleccionada por dejar menos residuos purínicos, indicada en el caso de los llamados cálculos metabólicos: urato, cistina y xantina.

¿Hay mucha diferencia entre los productos para perro y los productos para gato?

La estrategia nutricional de las dietas Urinary S/O es la misma para gatos y perros: baja SSR, dilución urinaria, efecto acidificante y control de precursores. La diferencia está en que en cada caso están ajustadas a las necesidades de cada especie.





¿En qué formatos se presenta? ¿Qué opciones hay disponibles?

La gama Urinary se presenta en formato seco y húmedo, disponible este último en diferentes texturas: bocaditos en salsa para gatos, en finas láminas en salsa para perros y paté para ambos, con el fin de satisfacer las distintas preferencias.

En el caso de las patologías de tracto urinario inferior, la presentación húmeda es especialmente interesante, ya que su alto contenido de humedad favorece la dilución urinaria y es la primera elección en caso de cistitis idiopática felina.





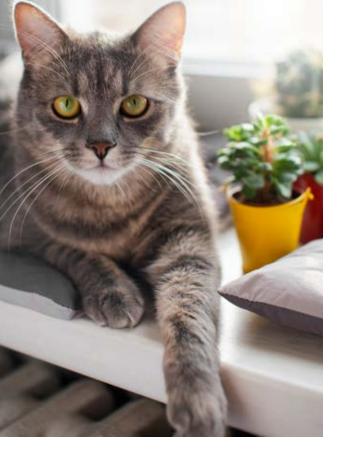
Además, es importante destacar que dentro de las dietas Urinary S/O disponemos de distintas opciones para dar una respuesta más precisa según el caso: si se trata de perros pequeños (de menos de 10kg), Urinary S/O Small Dogs; para perros mayores Urinary S/O Ageing 7+, especialmente adaptado a los cambios de las necesidades asociados a la edad y para aquellos gatos o perros con tendencia a engordar o un ligero sobrepeso, Urinary S/O Moderate Calorie.

¿Qué tipo de patologías podemos abordar con el uso de la gama Urinary?

La gama Urinary nos permite ofrecer un manejo nutricional específico en el caso de cálculos de estruvita y oxalato con las dietas Urinary S/O, también indicada como apoyo en casos de cistitis bacterianas en el perro y cistitis idiopática felina. La dieta Urinary UC es la opción para los cálculos de urato, cistina y xantina.







¿Cuáles son los tipos de cálculos más frecuentes en el perro y en el gato?

En ambas especies, las urolitiasis más comunes son las de estruvita y oxalato, seguidas de lejos por las de urato.

Aunque los datos pueden variar según el estudio que se consulte, los cálculos de estruvita en el perro suponen en torno al 35% en perros y el 40% en gatos y los de oxalato, aproximadamente un 45% y un 50% respectivamente. Los de urato son los siguientes, pero mucho menos frecuentes, con cerca de un 3% y un 4%. ¿Existe algún tipo de variabilidad dependiendo de la edad del animal? ¿Y de la raza? Nos referimos a si hay razas más predispuestas que otras.

Existen factores de riesgo de aparición de cálculos urinarios como la edad, la raza y el sexo, que dependen de cada tipo de urolito. En general, sabemos que los perros de razas pequeñas son más predispuestos a la formación de cálculos, que en las perras es más frecuente la estruvita porque presentan más infecciones urinarias y que en cuanto a los cálculos de urato, los más propensos son los Dálmata.

¿Qué otro tipo de cuidados serían complementarios para nuestro animal mientras usamos la gama Urinary? ¿Recomendáis una visita periódica al veterinario? ¿Cada cuánto tiempo?

Cuando se inicia el manejo nutricional con las dietas Urinary S/O es muy importante que la dieta sea el único alimento que reciba el animal, ya que es la manera en que la dieta puede ser eficaz. Si se mezcla con otros productos o se le añaden alimentos como comida de la mesa, trocitos de carne u otros complementos, la dieta perderá efectividad.

Si se quiere dar algo más que el alimento seco, siempre se puede combinar con la versión equivalente de la dieta en presentación húmeda, que además de mantener el mismo enfoque nutricional, aporta gran cantidad de agua que favorece la dilución urinaria.

Por supuesto, siempre tiene que haber un seguimiento por parte del veterinario, mientras se está intentando la disolución de los cálculos y también después para controlar que no reaparezcan de nuevo, detectando los primeros signos lo antes posible. Según el caso el veterinario indicará la frecuencia de las visitas

Por último, ¿dónde podemos encontrar esta línea de productos?

ROYAL CANIN[®] Urinary S/O solo debe utilizarse bajo prescripción del veterinario, por lo que podrán encontrarse en clínicas veterinarias.







TODOS LOS PROBLEMAS URINARIOS MERECEN SER SOLUCIONADOS

La enfermedad del tracto urinario inferior abarca una gran variedad de afecciones, se manifiesta a través de diversos signos y puede estar causada por múltiples problemas o la comorbilidad de diversas patologías.

Gracias a más de 50 años de ciencia, a una observación meticulosa y a la colaboración con veterinarios, sabemos que una nutrición especializada puede tener un papel fundamental en la recuperación de pacientes con problemas urinarios, así como en la salud general de los animales.

Por eso, disponemos de una amplia gama de soluciones nutricionales a medida para los problemas específicos del sistema urinario, ahora con innovaciones de última generación.







Animal models in osteosarcoma

Modelos animales en osteosarcoma

Maria V. Guijarro*, Steven C. Ghivizzani and C. Parker Gibbs

Department of Orthopaedics and Rehabilitation, University of Florida, Gainesville, FL, USA

https://www.frontiersin.org/journals/oncology/articles/10.3389/fonc.



Palabras clave:

osteosarcoma, modelos condicionales de ratón, modelos de ratón de línea germinal, modelos animales, p53, RB

Keywords:

osteosarcoma, conditional mouse models, germ-line mouse models, animal models, p53, RB



Tabla de contenido:

- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation Studies
- Genetically
- Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References



2014.00189/full

osteosarcoma (SG) es el tumor óseo primario no hematológico más frecuente en niños y adultos. La quimioterapia citotóxica en dosis altas y la resección quirúrgica han mejorado el pronóstico, con una supervivencia a largo plazo para la enfermedad no metastásica cercana al 70%. Sin embargo, la mayoría de los tumores de SG son de alto grado y tienden a desarrollar rápidamente metástasis pulmonares. A pesar de los avances clínicos, los pacientes con enfermedad metastásica o recaída tienen un mal pronóstico. Para una mejor comprensión de la patogénesis molecular de la SG humana, se han desarrollado varios modelos de ratón con SG modificados genéticamente y se revisarán aquí.

steosarcoma (OS) is the most common non-hematologic primary tumor of bone in children and adults. Highdose cytotoxic chemotherapy and surgical resection have improved prognosis, with long-term survival for non-metastatic disease approaching 70%. However, most OS tumors are high grade and tend to rapidly develop pulmonary metastases. Despite clinical advances, patients with metastatic disease or relapse have a poor prognosis. Toward a better understanding of the molecular pathogenesis of human OS, several genetically modified OS mouse models have been developed and will be reviewed here. However, better animal models that more accurately recapitulate the natural progression of the disease are needed for the development of improved prognostic and diagnostic markers as well as targeted therapies for both primary and metastatic OS.

Introduction

Osteosarcoma (OS) is a highly malignant form of bone cancer characterized by osteoid production. Although OS comprises <1% of cancers diagnosed in the United States, it is the most common primary malignancy of the bone (1, 2). It occurs predominantly after the first decade of life during periods of skeletal growth, with a second peak incidence in the geriatric patient population (1, 3). The vast majority of OS in children, adolescents, and young adults is high grade and begins in the intramedullary space of metaphyseal locations in long bones of the lower extremity. This suggests a relationship with active growth plates. After a low incidence in individuals between 25 and 59 years of age, the incidence of OS rises again in individuals over 60 years of age, and is most often associated with Paget's disease or radiation exposure (1, 2). This may suggest that the underlying pathogenesis is not identical in young and older patients. Conventional OS presents in three major subtypes based on histological classification: osteoblastic, fibroblastic, and chondroblastic. Osteoblastic is the most common (around 60%) with fibroblastic and chondroblastic being equally represented (4).

Osteosarcoma is characterized by a local invasion of bone and soft tissue, loss of the function of the affected extremity, and distant metastasis, most often to the lung (90%). Metastases are also found in bone (8–10%) and rarely in lymph nodes (5). Treatment involves aggressive removal of the primary tumor to afford local control via limb sparing surgery or amputation. Systemic chemotherapy (both prior to and after tumor removal) is used to suppress development of metastasis and effect cure. The most common chemotherapy



- Tabla de contenido:
- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation Studies
- Genetically
- Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References



regimens comprise the drugs, cisplatin, doxorubicin, and high-dose methotrexate in combination (6-8). Although chemotherapy slows tumor growth, it can induce cardiomyopathy, hearing loss, and risk of secondary malignancy (8, 9). In patients without metastases at the time of diagnosis (80-90%), surgical treatment in combination with chemotherapy has resulted in long-term survival rates that approach 70%. In contrast, for patients with established metastases there is currently no reliable therapeutic option to provide longterm tumor control. Despite intensive efforts to improve both chemotherapeutics and surgical management, 40% of all OS patients succumb to the disease. Specifically, the clinical outcome for metastatic OS remains poor; fewer than 30% of patients who present metastases survive 5 years after initial diagnosis. Therefore, there is an urgent need for the development of novel therapeutics for OS agents with increased capacity to eliminate systemic tumor burden as well as reduced toxicity in healthy tissues.

Etiology of OS

Osteosarcoma is characterized by a complex karyotype and a lack of recurrent translocations. Genetic approaches have identified several genes of potential importance in the development and progression of the disease (10-12). However, the widespread chromosomal alterations of the OS genome have limited the interpretation of these findings. Genetic alterations of OS are usually sporadic though genetic predisposition has been documented in patients with Li-Fraumeni and retinoblastoma syndrome. Somatic deletions and point mutations in P53 occur in approximately 50% of human OS (13-16) and half of those mutations are associated with loss of the remaining allele (14). Additionally, almost 70% of OS have at least one RB allele alteration (17, 18). Homozygous deletions of RB are seen in 23% of tumors, while point mutations appear in 6% (18, 19). In addition, numerous alterations that disrupt the RB pathway have also been reported; for example, the loss of function at the INK4a/ARF locus and the amplification of CDK4 have been found to occur (one or the other) in 22% of OS (20-22). The prevalence of these alterations would suggest that the deregulation of both G1/S and G2/M checkpoint in the cell cycle are a common event in OS.

For this, a tumor of unknown origin, chaotic genetics, early onset, and aggressive behavior, there is a need for more representative models to learn more about the biology of OS.

Animal Models in OS

Animal models hold significant promise in increasing our understanding of the genetic basis of OS and more importantly, in advancing preclinical studies aimed to the rational development of new therapeutic approaches as well as their validation prior to clinical trials.

In order for any animal model of human disease to be useful and informative. it is preferable to accurately recapitulate the natural course of the disease. Unfortunately, the etiology and pathogenesis of OS are not completely understood: therefore, the establishment and induction of representative experimental models are challenging and incomplete. Currently, there is not a robust animal model of OS that fully represents its biological and clinical features. The ideal would be one in which there was a naturally occurring primary bone lesion and spontaneous pulmonary metastases. To date, the major species used to generate OS models are mouse and rat; however, OS arising in dogs is also of note as a validated model of spontaneous OS.

Many aspects of the biology of the disease have been determined from a variety of animal model approaches. Genetically modified mouse models of OS have given the field much insight. However, spontaneous OS, secondary OS as a consequence of animals receiving



- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation Studies
- Genetically
- Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References



radiation, human and murine OS cell lines, and xenotransplantation studies are also important to understand the biology of this malignancy.

Canine Models

Spontaneous OS is much more common in large dogs than in humans, making the dog an attractive candidate model to study human disease (23). Canine OS is indistinguishable from human tumors at the histological and gene expression levels (24-27). The primary differences between the two are the age of development and the prevalence of the disease. In dogs, OS is a disease of older, large breed dogs (6–12 years of age), and it is estimated that over 10.000 cases occur annually in the United States. The median disease-free interval following surgery alone is 4 months, and after surgery with chemotherapy, 13 months. This high prevalence and the relatively rapid rate of disease progression provide the opportunity to model metastasis development and progression and evaluate novel treatment options in a relatively short period of time (28-32). Many of the genes involved in human OS pathogenesis appear to participate in canine OS, including P53, RB, and PTEN (33-36).

Although canine OS serves as an excellent comparative tumor model for human OS, there are some limitations to be considered. First, OS affects skeletally mature, geriatric dogs, which is different from humans where the peak of incidence occurs during adolescence. Second, some breeds have specific heritable germ-line mutations in certain genes that may influence OS biology, progression, and response to treatment without driving the initiation of the disease (37).

Secondary OS after Radiation

The development of rodent OS models began with the exposure of rats and mice to chemical and radioactive carcinogens (38–40). Of note, among those was the development of OS in rats treated with P32-orthophosphate, which resulted in a high incidence (41). These models yielded tumors that histologically resembled the human cancer and produced cell lines that complement human OS studies (42). Despite the high penetrance of the models, their relevance remains unclear since the majority of OS in humans is sporadic, while the carcinogen-induced murine model is more representative of a therapy induced disease.

Xenotransplantation Studies

There is a significant amount of literature related to the development and use of xenograft and allograft models of human and murine OS cells injected into immunocompromised mice. The injected cells form a solid tumor locally grown within days or weeks after implantation (42, 43). The use of these systems has become a prominent tool in current oncological research due to the quick onset, its affordable cost, and ease of handling and maintenance. In addition, OS donor-derived cells may metastasize to the lungs, providing an opportunity to investigate primary and secondary tumor growth. The principal limitation is that the approach uses fully developed OS cells and therefore does not provide information about the initiation of the tumor and its etiology. Furthermore, since the tumor microenvironment can contribute significantly to the tumor behavior, such interactions may be lost when establishing the disease by direct introduction into a recipient animal (44–46). In certain circumstances, the injected cell line may not be metastatic in the rodent context, making it impossible to study the dissemination of the disease. Despite these limitations, many groups have successfully used this model to identify factors involved in OS migration (47, 48) and more importantly for screening drugs with tumoricidal potential (49). Distinct advantages of the



- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation Studies
- Genetically
- Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References



subcutaneous cell suspension injection model are high rate of incidence and reproducibility that allows for accurate titration of cell numbers in the inoculum to quantify tumorigenic potential of the injected cells.

A variation of injecting cell suspensions into recipient animals is to transplant pieces of tumor directly harvested from the patient. The advantage is that the human malignant cells can grow in its native environment maintaining the heterogeneity that may be required for their proliferation, which in some reports has been shown to enhance tumor growth and metastasis. With the use of cell suspension and transplants, murine host cells can infiltrate the tumor, possibly influencing the activities of the tumor cells, and in some cases, cells of the rodent host can overgrow the human cell population (50). Alternatively orthotopic, intratibial implantation of OS cells has been shown to induce OS at local and metastatic sites (proximal tibia and lung) (43, 51–53). This approach allows the study of primary tumor formation within a more native context as well as the early stages of metastatic progression of OS, thereby reconstituting the entire metastatic process. Its use, however, is limited by a lack of reproducibility due in part to the technical skill required to perform the implantation and the associated lack of quantifiable inoculum.

Genetically Engineered Mouse Models

Of the sarcomas with complex karyotypes, OS is one of the most well-studied as exemplified by the development of numerous mouse models available for this disease. The ability to alter specifically the expression of individual genes (by loss or gain of function) became available in the mouse with the evolution of gene targeting technologies (54, 55).

Many murine OS models have been developed to recapitulate the P53 and RB mutations in hereditary and sporadic human OS. Germ-line deletion of P53 resulted in an OS incidence of 4% in homozygous P53 null mice (56) and 25% in heterozygous P53 mice (57), underlying the importance of altered P53 in driving OS. This unexpected ratio of tumor formation, though, is likely due to the early lethality seen in the homozygous null population. Further, the rapid development, the higher incidence of other tumors (mostly lymphomas), and the long latency of OS (58) necessitate the sacrifice of the mice before OS onset, hampering in many cases the utility of these models. The role of P53 was further highlighted by tumor analysis of P53 knock-in mice containing a mutant copy of P53R172H (corresponding to the R175H hot-spot mutation in humans) that not only develop primary tumors but also metastasize to the lungs as well as other organs (59, 60). Conversely, mice with germ-line deletions of Rb did not develop OS: homologous deletion of Rb is embryonic lethal and the heterozygotes are not predisposed to OS (61, 62).

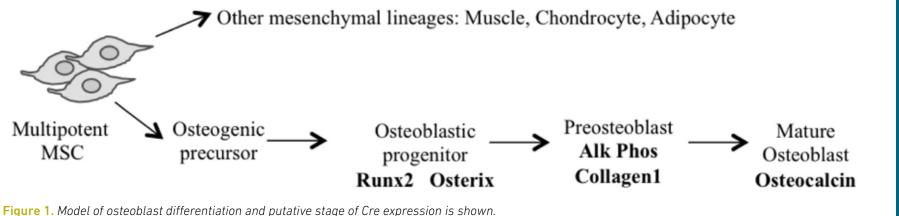
The application of conditional gene regulation and the availability of tissue specific Cre expressing mouse lines (63) have greatly enhanced our ability to generate specific models of mesenchymal osteogenic lineage that more faithfully resemble human OS (55, 64). The majority of these models have used the loss of P53 with or without the disruption of the Rb pathway to generate penetrant OS models (54). They use conditional gene deletion approaches restricted to multipotent mesenchymal progenitors, early committed osteoblasts (pre-osteoblasts) and the osteoblast population (Figure 1) (Table 1).

Using Cre recombinase activated by the gene promoter of Paired related homebox 1 (Prx1-Cre) (72) that deletes LoxP flanked alleles in the early limb mesenchyme (multipotential cells), 22% of mice with P53-mediated heterozygosity developed OS. Not surprisingly, homozygous deletion of P53 had a threefold increase in OS incidence over the heterozygous animals. In contrast, the deletion of Rb in the mesenchymal Prx expressing progenitors did not produce any OS tumors (65, 66). Interestingly,



- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation Studies
- Genetically
 - Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References





the highest incidence (92%) of OS occurred with the combined deletion of one allele of Rb with homozygous P53 deletion (66). Homozygous deletion of both genes resulted in more non-specific tumor formation with only 18% OS tumors and the remainder being poorly differentiated soft tissue sarcomas (PD-STS) and lymphoma (65, 66).

For a more restricted deletion of genes in the osteoblast lineage, promoters of genes ranging from those expressed early in the commitment of progenitors as Osterix 1 and Collagen1 α 1–3.6 to those expressed in more lineage-restricted osteoblast precursors such as Collagen1 α 1–2.3 and osteocalcin (Og2) have been used. Development of OS with a penetrance of 100% (67, 68) has been observed following osteoblast specific deletion of P53 using Osterix-mediated Cre expression (Osx-Cre) (73). As with mesenchymal pro-

genitors, Rb deletions have no effect and combined deletion of Rb and P53 in osteoblasts once again generated fibroblastic or undifferentiated OS with high penetrance (100%) (67, 68). Potential translational utility is the existence of short-latency spontaneous metastatic OS similar to human tumors in which cells are arrested in their differentiation (67, 68). Although the greatest proportion of tumors was OS when P53 was conditionally deleted, neuroendocrine tumors and hibernomas were also reported to be generated in several mice (67, 68). However, Walkley et al. enriched the C57BL/6 background of the mouse strain and the percentage of hibernomas was reduced, suggesting a possible impact of mouse strains in the phenotype observed (69). A recent study in mice that expressed SV40 T/t antigen (Tag) in mature osteoblasts under the Og2 (74) showed OS with complete penetrance (71) and 90% incidence of lung metastases. Further analysis of the tumors derived from this model revealed a recurrent genomic deletion of the Prkar1a gene in a specific subset also in human OS. Transgenic shR-NA has been used to specifically knock down P53 (rather than delete) using the Osx-Cre transgene (69). These mice develop osteoblastic OS with a 100% penetrance, and although they have a longer latency to tumor onset, they more often develop in long bones and are highly metastatic (lung and liver), features similar to human OS. This model has not developed any non-OS tumors.

Independent of the stage of development in which Cre becomes active, the latency of OS is essentially the same when comparing either P53 alone or in combination with Rb. The use of Cre in more primitive cells (Prx), however, leads to the development of tumors of other mesenchymal lineages at higher frequency.



- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation Studies
- Genetically
 - Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References



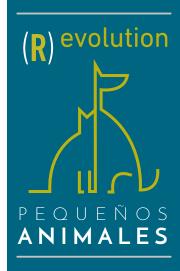
| Cell | Cre | Gene | OS penetrance (%) | Other tumors | Metastatic disease |
|-----------------|------------|---|-------------------------|--|----------------------|
| MSC/skeletal | Prx-1 | p53 ^{fl/+} | 22 (65) | | |
| Progenitors | | p53 ^{fl/fl} | 61 (65); 62(66) | PDS (32%), LY (3%), LPS (3%); RMS (15%), PDS (12%) | Yes (24%) |
| | | p53 ^{fl/fl} -Rb ^{fl/+} | 92 (66) | RMS (9%), PDS (18%), HIB (4%) | |
| | | p53 ^{fl/fl} -Rb ^{fl/fl} | 18 (65); 29 (66) | PDS (57%), LY (14%); RMS (12%), | |
| | | | | PDS (3%), HIB (91%) | |
| Pre-osteoblasts | Osx | p53 ^{fl/fl} | 100 (67); 100 (68) | | Yes (32%); yes (40%) |
| | | p53 ^{fl/fl} -Rb ^{fl/+} | 53 (67); 100 (68) | | |
| | | p53 ^{fl/fl} -Rb ^{fl/fl} | 72 (67); 100 (68) | Multiple tumors per animal; concurrent HIB (20–25%) | Yes (37%) |
| | | shp53 | 100 (69) | 0% | Yes (83.33%) |
| | | shp53-Rb ^{fl/+} | 100 (69) | 0% | Yes (58.82%) |
| | | shp53-Rb ^{fl/fl} | 100 (69) | 0% | Yes (85.72%) |
| | Col1a1–3.6 | p53 ^{fl/fl} | 60 (70) | | |
| Osteoblasts | Col1a1–2.3 | p53 ^{fl/fl} | 85 (65) | | |
| | Og2 | SV40 Tag | 100 (71) | | Yes (90%) |

LPS, liposarcoma; LY, lymphoma; RMS, rhabdomyosarcoma; PDS, poorly differentiated sarcoma; HIB, hibernomas.

Table 1. Summary of genetically modified OS murine models.

Possibly providing insight into the initiating events of OS (70), a prominent cellular feature of conditional inactivation of P53 in osteoblastic progenitors is the hyperproliferation of osteoblasts prior to tumor formation. Rb has been proposed to have a role in influencing late osteoblast differentiation by interacting with Runx2 (75). However, a number of independent studies have shown that the removal of Rb alone is not sufficient to induce OS. The different experimental approaches strongly suggest that mutation in the p53 pathway can serve as an initiating event in OS, with a subsequent mutation in the Rb pathway strongly accelerating tumor development.

These engineered mouse models of OS reproduce many features of human OS including similar gene-transcription signatures (76) and cytogenetic complexity. However, the sites of primary tumor formation in Cre–loxP mice do not recapitulate the spontaneous human disease. The majority of lesions (85%) arise in axial skeletal sites (mandibule, maxilla, rib/vertebra, skull, sternum) while on 13.6% of tumors developed from the appendicular skeleton (hind leg, front leg) (68). This contrasts with the anatomic distribution of OS diagnosed in humans, with the distal femur, proximal tibia, and proximal humerus being the most common sites involved and only 10% develop in the



- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation
 Studies
- Genetically
 - Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References



axial skeleton, most commonly the pelvis (5). Only in one study (69) did the tumor arise primarily in long bones. In addition, the observed frequency of distant metastases was comparatively low when compared to human disease except for the P53 knockdown model (69). As opposed to a complete deletion of P53, the primary tumor cells proliferated slower and the animals did not have to be sacrificed for local tumor size prior to completion of the metastatic process. Furthermore, the primary site of metastases in human OS is predominantly the lung parenchyma while in Cre-loxP mice, sites of metastases were more diverse with both the lung and liver being affected in almost equal proportions.

Other genes such as C-FOS (77, 78), TWIST (79), p14ARF (80), p16INK4a (81), PRKAR1A (71), and p21CIP (82) have also been implicated in OS pathogenesis based on studies of human OS samples. Their mutation appears to complement the defects in the P53 and RB pathways, and their involvement in osteosarcomagenesis is also demonstrated from genetically engineered mouse models. They provide important information regarding the genetics of OS, but the long latency combined with low penetrance makes utilization of these models less practical.

Targeted Therapies in OS

Osteosarcoma is very resistant to therapy and therefore there is an urgent need to effectively treat affected patients. The emergence of new anti-cancer drugs and the small number of patients eligible for early-phase clinical trials present another challenge in the clinical testing of novel compounds for OS treatment. As discussed earlier. xenotransplantation models have provided the greatest utility for preclinical screening of drugs with tumoricidal potential. To this end, the National Cancer Institute (NCI) has implemented the Pediatric Preclinical Testing Program (PPTP), a consortium of institutions across the United States and in Australia. Its objective is to identify agents with significant activity in panels of mouse xenograft models representing the most common pediatric cancers including OS (83). The program has been successful, leading to Phase I and II clinical trials for cixutumumab, sorafenib, and rapamycin for OS treatment. (84–86). In each case, these agents demonstrated high levels of response in the PPTP and were well-tolerated with promising anti-tumor activity in some adult and pediatric patients.

The use of spontaneous and transgenic OS models for high throughput screening of anti-OS drugs is hampered due to practical considerations associated

with the cost and time of generating sufficient numbers of animals for statistically meaningful data. This is due to variations in disease onset as well as tumor heterogeneity, incidence, and progression. However, the recent generation of transgenic animals expressing shRNAs to knock down P53 (69) represents a potential breakthrough with respect to preclinical screening. Unlike conventional Cre-mediated gene deletion approaches, P53 knock down mice exhibited 100% penetrance for osteoblastic OS (the most common form of the disease). Moreover, the tumors were most frequently present in long bones and preferentially disseminated to the lungs, consistent with human OS.

Another consideration for preclinical testing in in vivo models is the accurate measurement of the disease burden at non-accessible sites. The use of in vivo imaging offers the opportunity to detect and monitor the development and progression of the disease. However, imaging systems are costly and not always widely accessible for many researchers. OS has the advantage that the primary tumor in genetically engineered mouse models appears in long bones and is therefore more accessible than abdominal tumors. The monitoring/visualization of micrometastases represents a greater challenge due to their small size. Inaccurate evaluation of metastatic spread in preclinical studies potentially



- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation Studies
- Genetically
- Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References



leads to disappointing results in clinical trials. Consequently, there is great interest in refining the methods to enable reproducible and ultrasensitive detection of metastases at the single cell level. The main focus therefore is on techniques, which allow the detection of tumor cells in vivo, such as microcomputer tomography (micro-CT), positron emission tomography (PET), bioluminescence, or fluorescence imaging.

Conclusion

Our understanding of human OS biology is hindered by its rapid onset, low prevalence, and absence of predisposing conditions or precursor lesions. With limited human tissue available for study, animal models provide a valuable tool to investigate the underlying mechanisms driving tumor initiation, progression, metastatic events, and therapeutic interventions. While these models have yet to faithfully recapitulate all aspects of OS, there is no doubt that the study of OS animal models has enabled insight into the genetics of tumor initiation as well as the cellular and molecular profiles of tumor growth and metastasis. In particular, gene knockout studies have been instrumental in identifying genetic mutations that promote OS tumor initiation (P53), as well as co-operative mutations that increase disease incidence (RB, c-FOS).

With the use of cell lineage specific markers, it is now possible to introduce genetic mutations by sequential targeting from early precursor (multipotent mesenchymal cell) to more mature osteoblastic cells (osteoblast to osteocyte) to investigate OS incidence and tumor pathology. With this strategy, Prx1 and Osx have been used to identify mesenchymal and osteoprogenitor cells, respectively, following conditional mutation of P53. It remains to be seen, however, whether these populations are truly distinct, as Prx1 could be coexpressed with Osx in a certain subpopulation of cells. Another consideration particularly relevant in OS is its tumor heterogeneity among patients, which suggests that multiple cell types could act as cell of origin. Additionally, this concept of heterogeneity calls into question the utility of models exploiting single gene manipulation. Its consideration may permit a more systematic analysis of the genetic lesions involved in OS initiation and progression and could serve as a platform for the identification of early disease biomarkers. Cell of origin identification may also have important implications in the prevention of relapse and elucidate key molecular pathways and driver mutations that could lead to new therapeutic approaches to prevent the disease.

Thus, although for now, conventional orthotopic and subcutaneous trans-

plantation models will remain indispensable to continue the study of OS in vivo, new models of spontaneous OS need to be developed to further our understanding of OS biology. Models that accurately reproduce the establishment of spontaneous micrometastases are necessary to investigate novel antimetastatic agents, as this clinical scenario is most often the lethal event for patients with this form of cancer.

Conflict of Interest Statement

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

- Mirabello L, Troisi RJ, Savage SA. International osteosarcoma incidence patterns in children and adolescents, middle ages and elderly persons. Int J Cancer (2009) 125(1):229–34. doi: 10.1002/ ijc.24320
- Mirabello L, Troisi RJ, Savage SA. Osteosarcoma incidence and survival rates from 1973 to 2004: data from the Surveillance, Epidemiology, and End Results Program. Cancer (2009) 115(7):1531–43. doi:10.1002/cncr.24121
- McKenna RJ, Schwinn CP, Higinbotham NL. Osteogenic sarcoma in children. CA Cancer J Clin (1966) 16(1):26–8.



- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation Studies
- Genetically
- Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References



- 4. Gorlick R. Current concepts on the molecular biology of osteosarcoma. Cancer Treat Res (2009) 152:467–78. doi:10.1007/978-1-4419-0284-9_27
- 5. Luetke A, Meyers PA, Lewis I, Juergens H. Osteosarcoma treatment – where do we stand? A state of the art review. Cancer Treat Rev (2014) 40(4):523–32. doi:10.1016/j.ctrv.2013.11.006
- 6. Janeway KA, Grier HE. Sequelae of osteosarcoma medical therapy: a review of rare acute toxicities and late effects. Lancet Oncol (2010) 11(7):670–8. doi:10.1016/S1470-2045(10)70062-0
- Picci P, Ferrari S, Bacci G, Gherlinzoni F. Treatment recommendations for osteosarcoma and adult soft tissue sarcomas. Drugs (1994) 47(1):82–92. doi:10.2165/00003495-199447010-00006
- Sloet van Oldruitenborgh-Oosterbaan MM, Klein WR, Misdorp W. Differential diagnosis of non-healing 'fungal' patches in horses. Tijdschr Diergeneeskd (1994) 119(24):756– 9.
- Savage SA, Mirabello L, Wang Z, Gastier-Foster JM, Gorlick R, Khanna C, et al. Genome-wide association study identifies two susceptibility loci for osteosarcoma. Nat Genet (2013) 45(7):799–803. doi:10.1038/ng.2645
- Sandberg AA, Bridge JA. Updates on the cytogenetics and molecular genetics of bone and soft tissue tumors: osteosarcoma and related tumors. Cancer Genet Cytogenet (2003) 145(1):1–30. doi:10.1016/S0165-4608(02)00848-8

- Pasic I, Shlien A, Durbin AD, Stavropoulos DJ, Baskin B, Ray PN, et al. Recurrent focal copy-number changes and loss of heterozygosity implicate two noncoding RNAs and one tumor suppressor gene at chromosome 3q13.31 in osteosarcoma. Cancer Res (2010) 70(1):160–71. doi:10.1158/0008-5472.CAN-09-1902
- Fletcher JA, Gebhardt MC, Kozakewich HP. Cytogenetic aberrations in osteosarcomas. Nonrandom deletions, rings, and double-minute chromosomes. Cancer Genet Cytogenet (1994) 77(1):81–8. doi:10.1016/0165-4608(94)90154-6
- Overholtzer M, Rao PH, Favis R, Lu XY, Elowitz MB, Barany F, et al. The presence of p53 mutations in human osteosarcomas correlates with high levels of genomic instability. Proc Natl Acad Sci U S A (2003) 100(20):11547–52. doi:10.1073/ pnas.1934852100
- 14. Miller CW, Aslo A, Tsay C, Slamon D, Ishizaki K, Toguchida J, et al. Frequency and structure of p53 rearrangements in human osteosarcoma. Cancer Res (1990) 50(24):7950–4.
- Miller CW, Aslo A, Won A, Tan M, Lampkin B, Koeffler HP. Alterations of the p53, Rb and MDM2 genes in osteosarcoma. J Cancer Res Clin Oncol (1996) 122(9):559– 65. doi:10.1007/BF01213553
- Toguchida J, Yamaguchi T, Ritchie B, Beauchamp RL, Dayton SH, Herrera GE, et al. Mutation spectrum of the p53 gene in bone and soft tissue sarcomas. Cancer Res (1992) 52(22):6194–9.

- Hansen MF, Koufos A, Gallie BL, Phillips RA, Fodstad O, Brogger A, et al. Osteosarcoma and retinoblastoma: a shared chromosomal mechanism revealing recessive predisposition. Proc Natl Acad Sci U S A (1985) 82(18):6216–20. doi:10.1073/pnas.82.18.6216
- Wadayama B, Toguchida J, Shimizu T, Ishizaki K, Sasaki MS, Kotoura Y, et al. Mutation spectrum of the retinoblastoma gene in osteosarcomas. Cancer Res (1994) 54(11):3042–8.
- 19. Friend SH, Bernards R, Rogelj S, Weinberg RA, Rapaport JM, Albert DM, et al. A human DNA segment with properties of the gene that predisposes to retinoblastoma and osteosarcoma. Nature (1986) 323(6089):643–6. doi:10.1038/323643a0
- 20. Maitra A, Roberts H, Weinberg AG, Geradts J. Loss of p16(INK4a) expression correlates with decreased survival in pediatric osteosarcomas. Int J Cancer (2001) 95(1):34–8. doi:10.1002/1097-0215(20010120)95:1<34::AID-IJC1006>3.0.CO;2-V
- 21. Miller CW, Aslo A, Campbell MJ, Kawamata N, Lampkin BC, Koeffler HP. Alterations of the p15, p16, and p18 genes in osteosarcoma. Cancer Genet Cytogenet (1996) 86(2):136–42. doi:10.1016/0165-4608(95)00216-2
- Wei G, Lonardo F, Ueda T, Kim T, Huvos AG, Healey JH, et al. CDK4 gene amplification in osteosarcoma: reciprocal relationship with INK4A gene alterations and mapping of 12q13 amplicons. Int J Cancer (1999) 80(2):199–204. doi:10.1002/ (SICI)1097-0215(19990118)80:2<199::AID-IJC7>3.0.C0;2-4
- 23. Misdorp W. Skeletal osteosarcoma. Animal model: canine osteosarcoma. Am J Pathol (1980) 98(1):285–8.



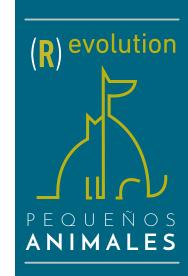
- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation
 Studies
- Genetically
- Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References



- 24. Selvarajah GT, Kirpensteijn J, van Wolferen ME, Rao NA, Fieten H, Mol JA. Gene expression profiling of canine osteosarcoma reveals genes associated with short and long survival times. *Mol Cancer* (2009) 8:72. doi:10.1186/1476-4598-8-72
- Paoloni M, Davis S, Lana S, Withrow S, Sangiorgi L, Picci P, et al. Canine tumor cross-species genomics uncovers targets linked to osteosarcoma progression. *BMC Genomics* (2009) 10:625. doi:10.1186/1471-2164-10-625
- Mueller F, Fuchs B, Kaser-Hotz B. Comparative biology of human and canine osteosarcoma. *Anticancer Res* (2007) 27(1A):155–64.
- Rankin KS, Starkey M, Lunec J, Gerrand CH, Murphy S, Biswas S. Of dogs and men: comparative biology as a tool for the discovery of novel biomarkers and drug development targets in osteosarcoma. *Pediatr Blood Cancer* (2012) 58(3):327–33. doi:10.1002/pbc.23341
- 28. Kirpensteijn J, Teske E, Kik M, Klenner T, Rutteman GR. Lobaplatin as an adjuvant chemotherapy to surgery in canine appendicular osteosarcoma: a phase II evaluation. *Anticancer Res* (2002) 22(5):2765–70.
- Kirpensteijn J, Timmermans-Sprang EP, van Garderen E, Rutteman GR, Lantingavan Leeuwen IS, Mol JA. Growth hormone gene expression in canine normal growth plates and spontaneous osteosarcoma. *Mol Cell Endocrinol* (2002) 197(1–2):179– 85. doi:10.1016/S0303-7207(02)00269-1
- Kirpensteijn J, Kik M, Rutteman GR, Teske E. Prognostic significance of a new histologic grading system for canine osteosarcoma. *Vet Pathol* (2002) 39(2):240– 6. doi:10.1354/vp.39-2-240

- 31. Lascelles BD, Dernell WS, Correa MT, Lafferty M, Devitt CM, Kuntz CA, et al. Improved survival associated with postoperative wound infection in dogs treated with limb-salvage surgery for osteosarcoma. *Ann Surg Oncol* (2005) 12(12):1073–83. doi:10.1245/ AS0.2005.01.011
- Kirpensteijn J, Kik M, Teske E, Rutteman GR. TP53 gene mutations in canine osteosarcoma. *Vet Surg* (2008) 37(5):454– 60. doi:10.1111/j.1532-950X.2008.00407.x
- 33. van Leeuwen IS, Cornelisse CJ, Misdorp W, Goedegebuure SA, Kirpensteijn J, Rutteman GR. P53 gene mutations in osteosarcomas in the dog. *Cancer Lett* (1997) 111(1–2):173–8. doi:10.1016/ S0304-3835(96)04529-6
- 34. Johnson AS, Couto CG, Weghorst CM. Mutation of the p53 tumor suppressor gene in spontaneously occurring osteosarcomas of the dog. *Carcinogenesis* (1998) 19(1):213–7. doi:10.1093/ carcin/19.1.213
- 35. Levine RA, Fleischli MA. Inactivation of p53 and retinoblastoma family pathways in canine osteosarcoma cell lines. *Vet Pathol* (2000) 37(1):54–61. doi:10.1354/vp.37-1-54
- 36. Levine RA, Forest T, Smith C. Tumor suppressor PTEN is mutated in canine osteosarcoma cell lines and tumors. *Vet Pathol* (2002) 39(3):372–8. doi:10.1354/ vp.39-3-372
- 37. Liao AT, McMahon M, London CA. Identification of a novel germline MET mutation in dogs. *Anim Genet* (2006) 37(3):248–52. doi:10.1111/j.1365-2052.2006.01415.x

- Martin TJ, Ingleton PM, Underwood JC, Michelangeli VP, Hunt NH, Melick RA. Parathyroid hormone-responsive adenylate cyclase in induced transplantable osteogenic rat sarcoma. *Nature* (1976) 260(5550):436–8. doi:10.1038/260436a0
- Ingleton PM, Coulton LA, Preston CJ, Martin TJ. Alkaline phosphatase in serum and tumour of rats bearing a hormoneresponsive transplantable osteogenic sarcoma. *Eur J Cancer* (1979) 15(5):685– 91. doi:10.1016/0014-2964(79)90142-7
- 40. Underwood JC, Melick RA, Loomes RS, Dangerfield VM, Crawford A, Coulton L, et al. Structural and functional correlations in parathyroid hormone responsive transplantable osteogenic sarcomas. *Eur J Cancer* (1979) 15(9):1151–8. doi:10.1016/0014-2964(79)90131-2
- 41. Bensted JP, Blackett NM, Lamerton LF. Studies on the development of radiationinduced bone tumours. *Acta Unio Int Contra Cancrum* (1959) 15:559–60.
- 42. Ek ET, Dass CR, Choong PF. Commonly used mouse models of osteosarcoma. *Crit Rev Oncol Hematol* (2006) 60(1):1–8. doi:10.1016/j.critrevonc.2006.03.006
- 43. Dass CR, Ek ET, Choong PF. Human xenograft osteosarcoma models with spontaneous metastasis in mice: clinical relevance and applicability for drug testing. *J Cancer Res Clin Oncol* (2007) 133(3):193–8. doi:10.1007/s00432-006-0157-x
- 44. Becher OJ, Holland EC. Genetically engineered models have advantages over xenografts for preclinical studies. *Cancer Res* (2006) 66(7):3355–8. doi:10.1158/0008-5472.CAN-05-3827 discussion 8-9,



- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation Studies
- Genetically
- Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References

- 45. Sharpless NE, Depinho RA. The mighty mouse: genetically engineered mouse models in cancer drug development. *Nat Rev Drug Discov* (2006) 5(9):741–54. doi:10.1038/nrd2110
- 46. Frese KK, Tuveson DA. Maximizing mouse cancer models. *Nat Rev Cancer* (2007) 7(9):645–58. doi:10.1038/nrc2192
- 47. Khanna C, Prehn J, Yeung C, Caylor J, Tsokos M, Helman L. An orthotopic model of murine osteosarcoma with clonally related variants differing in pulmonary metastatic potential. *Clin Exp Metastasis* (2000) 18(3):261–71. doi:10.1023/A:1006767007547
- Khanna C, Khan J, Nguyen P, Prehn J, Caylor J, Yeung C, et al. Metastasisassociated differences in gene expression in a murine model of osteosarcoma. *Cancer Res* (2001) 61(9):3750–9.
- 49. Sampson VB, Gorlick R, Kamara D, Anders Kolb E. A review of targeted therapies evaluated by the pediatric preclinical testing program for osteosarcoma. *Front Oncol* (2013) 3:132. doi:10.3389/ fonc.2013.00132
- 50. Kresse SH, Meza-Zepeda LA, Machado I, Llombart-Bosch A, Myklebost O. Preclinical xenograft models of human sarcoma show nonrandom loss of aberrations. *Cancer* (2012) 118(2):558–70. doi:10.1002/ cncr.26276
- 51. Berlin O, Samid D, Donthineni-Rao R, Akeson W, Amiel D, Woods VL Jr. Development of a novel spontaneous metastasis model of human osteosarcoma transplanted orthotopically into bone of athymic mice. *Cancer Res* (1993) 53(20):4890–5.

- 52. Crnalic S, Hakansson I, Boquist L, Lofvenberg R, Brostrom LA. A novel spontaneous metastasis model of human osteosarcoma developed using orthotopic transplantation of intact tumor tissue into tibia of nude mice. *Clin Exp Metastasis* (1997) 15(2):164–72. doi:10.1023/A:1018456911823
- 53. Yuan J, Ossendorf C, Szatkowski JP, Bronk JT, Maran A, Yaszemski M, et al. Osteoblastic and osteolytic human osteosarcomas can be studied with a new xenograft mouse model producing spontaneous metastases. *Cancer Invest* (2009) 27(4):435–42. doi:10.1080/07357900802491477
- 54. Janeway KA, Walkley CR. Modeling human osteosarcoma in the mouse: from bedside to bench. *Bone* (2010) 47(5):859–65. doi:10.1016/j.bone.2010.07.028
- 55. Ng AJ, Mutsaers AJ, Baker EK, Walkley CR. Genetically engineered mouse models and human osteosarcoma. *Clin Sarcoma Res* (2012) 2(1):19. doi:10.1186/2045-3329-2-19
- 56. Donehower LA, Harvey M, Slagle BL, McArthur MJ, Montgomery CA Jr, Butel JS, et al. Mice deficient for p53 are developmentally normal but susceptible to spontaneous tumours. *Nature* (1992) 356(6366):215–21. doi:10.1038/356215a0
- 57. Jacks T, Remington L, Williams BO, Schmitt EM, Halachmi S, Bronson RT, et al. Tumor spectrum analysis in p53-mutant mice. *Curr Biol* (1994) 4(1):1–7. doi:10.1016/ S0960-9822(00)00002-6
- 58. Lavigueur A, Maltby V, Mock D, Rossant J, Pawson T, Bernstein A. High incidence of lung, bone, and lymphoid tumors in transgenic mice overexpressing mutant alleles of the p53 oncogene. *Mol Cell Biol* (1989) 9(9):3982–91.

- 59. Lang GA, Iwakuma T, Suh YA, Liu G, Rao VA, Parant JM, et al. Gain of function of a p53 hot spot mutation in a mouse model of Li-Fraumeni syndrome. *Cell* (2004) 119(6):861–72. doi:10.1016/j. cell.2004.11.006
- 60. Olive KP, Tuveson DA, Ruhe ZC, Yin B, Willis NA, Bronson RT, et al. Mutant p53 gain of function in two mouse models of Li-Fraumeni syndrome. *Cell* (2004) 119(6):847–60. doi:10.1016/j. cell.2004.11.004
- 61. Williams BO, Remington L, Albert DM, Mukai S, Bronson RT, Jacks T. Cooperative tumorigenic effects of germline mutations in Rb and p53. *Nat Genet* (1994) 7(4):480–4. doi:10.1038/ng0894-480
- 62. Lee EY, Chang CY, Hu N, Wang YC, Lai CC, Herrup K, et al. Mice deficient for Rb are nonviable and show defects in neurogenesis and haematopoiesis. *Nature* (1992) 359(6393):288–94. doi:10.1038/359288a0
- 63. VanKoevering KK, Williams BO. Transgenic mouse strains for conditional gene deletion during skeletal development. *IBMS boneKEy* (2008) 5:151–70. doi:10.1138/20080312
- 64. Mohseny AB, Hogendoorn PC, Cleton-Jansen AM. Osteosarcoma models: from cell lines to zebrafish. *Sarcoma* (2012) 2012:417271. doi:10.1155/2012/417271
- 65. Lin PP, Pandey MK, Jin F, Raymond AK, Akiyama H, Lozano G. Targeted mutation of p53 and Rb in mesenchymal cells of the limb bud produces sarcomas in mice. *Carcinogenesis* (2009) 30(10):1789–95. doi:10.1093/carcin/bgp180



- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation Studies
- Genetically
- Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References

- Calo E, Quintero-Estades JA, Danielian PS, Nedelcu S, Berman SD, Lees JA. Rb regulates fate choice and lineage commitment in vivo. *Nature* (2010) 466(7310):1110–4. doi:10.1038/ nature09264
- Berman SD, Calo E, Landman AS, Danielian PS, Miller ES, West JC, et al. Metastatic osteosarcoma induced by inactivation of Rb and p53 in the osteoblast lineage. *Proc Natl Acad Sci U S A* (2008) 105(33):11851– 6. doi:10.1073/pnas.0805462105
- 68. Walkley CR, Qudsi R, Sankaran VG, Perry JA, Gostissa M, Roth SI, et al. Conditional mouse osteosarcoma, dependent on p53 loss and potentiated by loss of Rb, mimics the human disease. *Genes Dev* (2008) 22(12):1662–76. doi:10.1101/gad.1656808
- 69. Mutsaers AJ, Ng AJ, Baker EK, Russell MR, Chalk AM, Wall M, et al. Modeling distinct osteosarcoma subtypes in vivo using Cre:lox and lineage-restricted transgenic shRNA. *Bone* (2013) 55(1):166–78. doi:10.1016/j.bone.2013.02.016
- Lengner CJ, Steinman HA, Gagnon J, Smith TW, Henderson JE, Kream BE, et al. Osteoblast differentiation and skeletal development are regulated by Mdm2-p53 signaling. *J Cell Biol* (2006) 172(6):909–21. doi:10.1083/jcb.200508130
- 71. Molyneux SD, Di Grappa MA, Beristain AG, McKee TD, Wai DH, Paderova J, et al. Prkar1a is an osteosarcoma tumor suppressor that defines a molecular subclass in mice. J Clin Invest (2010) 120(9):3310–25. doi:10.1172/JCI42391
- 72. Logan M, Martin JF, Nagy A, Lobe C, Olson EN, Tabin CJ. Expression of Cre recombinase in the developing mouse limb bud driven by a Prxl enhancer. *Genesis* (2002) 33(2):77–80. doi:10.1002/ gene.10092

- 73. Rodda SJ, McMahon AP. Distinct roles for Hedgehog and canonical Wnt signaling in specification, differentiation and maintenance of osteoblast progenitors. *Development* (2006) 133(16):3231–44. doi:10.1242/dev.02480
- 74. Frendo JL, Xiao G, Fuchs S, Franceschi RT, Karsenty G, Ducy P. Functional hierarchy between two OSE2 elements in the control of osteocalcin gene expression in vivo. *J Biol Chem* (1998) 273(46):30509–16. doi:10.1074/jbc.273.46.30509
- 75. Thomas DM, Carty SA, Piscopo DM, Lee JS, Wang WF, Forrester WC, et al. The retinoblastoma protein acts as a transcriptional coactivator required for osteogenic differentiation. *Mol Cell* (2001) 8(2):303–16. doi:10.1016/S1097-2765(01)00327-6
- 76. Kuijjer ML, Namlos HM, Hauben EI, Machado I, Kresse SH, Serra M, et al. mRNA expression profiles of primary highgrade central osteosarcoma are preserved in cell lines and xenografts. *BMC Med Genomics* (2011) 4:66. doi:10.1186/1755-8794-4-66
- 77. Weisstein JS, Majeska RJ, Klein MJ, Einhorn TA. Detection of c-fos expression in benign and malignant musculoskeletal lesions. *J Orthop Res* (2001) 19(3):339–45. doi:10.1016/S0736-0266(00)90020-2
- Ruther U, Komitowski D, Schubert FR, Wagner EF. C-Fos expression induces bone tumors in transgenic mice. *Oncogene* (1989) 4(7):861–5.
- 79. Entz-Werle N, Choquet P, Neuville A, Kuchler-Bopp S, Clauss F, Danse JM, et al. Targeted apc;twist double-mutant mice: a new model of spontaneous osteosarcoma that mimics the human disease. *Transl Oncol* (2010) 3(6):344–53. doi:10.1593/ tlo.10169

- Krimpenfort P, Ijpenberg A, Song JY, van der Valk M, Nawijn M, Zevenhoven J, et al. p15Ink4b is a critical tumour suppressor in the absence of p16Ink4a. *Nature* (2007) 448(7156):943–6. doi:10.1038/ nature06084
- Sharpless NE, Bardeesy N, Lee KH, Carrasco D, Castrillon DH, Aguirre AJ, et al. Loss of p16Ink4a with retention of p19Arf predisposes mice to tumorigenesis. *Nature* (2001) 413(6851):86–91. doi:10.1038/35092592
- 82. Martin-Caballero J, Flores JM, Garcia-Palencia P, Serrano M. Tumor susceptibility of p21(Waf1/Cip1)-deficient mice. *Cancer Res* (2001) 61(16):6234–8.
- 83. Houghton PJ, Morton CL, Tucker C, Payne D, Favours E, Cole C, et al. The pediatric preclinical testing program: description of models and early testing results. *Pediatr Blood Cancer* (2007) 49(7):928–40. doi:10.1002/pbc.21078
- 84. Chawla SP, Staddon AP, Baker LH, Schuetze SM, Tolcher AW, D'Amato GZ, et al. Phase II study of the mammalian target of rapamycin inhibitor ridaforolimus in patients with advanced bone and soft tissue sarcomas. *J Clin Oncol* (2012) 30(1):78–84. doi:10.1200/ JC0.2011.35.6329
- 85. Grignani G, Palmerini E, Dileo P, Asaftei SD, D'Ambrosio L, Pignochino Y, et al. A phase II trial of sorafenib in relapsed and unresectable high-grade osteosarcoma after failure of standard multimodal therapy: an Italian Sarcoma Group study. *Ann Oncol* (2012) 23(2):508–16. doi:10.1093/annonc/mdr151



- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation Studies
- Genetically
- Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References

66 de 72

86. Malempati S, Weigel B, Ingle AM, Ahern CH, Carroll JM, Roberts CT, et al. Phase I/II trial and pharmacokinetic study of cixutumumab in pediatric patients with refractory solid tumors and Ewing sarcoma: a report from the Children's Oncology Group. J Clin Oncol (2012) 30(3):256–62. doi:10.1200/ JC0.2011.37.4355

Citation: Guijarro MV, Ghivizzani SC and Gibbs CP (2014) Animal models in osteosarcoma. *Front. Oncol.* 4:189. doi: 10.3389/fonc.2014.00189

Received: 31 May 2014; Accepted: 07 July 2014; Published online: 18 July 2014.

Edited by: Amancio Carnero, Instituto de Biomedicina de Sevilla, Spain



Reviewed by: Carmen Blanco Aparicio, Spanish National Cancer Research Centre, Spain Irene Ferrer, IBIS, Spain

Copyright: © 2014 Guijarro, Ghivizzani and Gibbs. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms. *Correspondence: Maria V. Guijarro, Department of Orthopaedics and Rehabilitation, University of Florida, 1600 Archer Road, MSB M2-212, Gainesville, FL 32610, USA e-mail: guijam@ufl.edu

This article is part of the Research Top-ic

Genetically modified mouse models of cancer



Tabla de contenido:

- Introduction
- Etiology of OS
- Animal Models in OS
- Canine Models
- Secondary OS after Radiation
- Xenotransplantation Studies
- Genetically
- Engineered Mouse Models
- Targeted Therapies in OS
- Conclusion
- Conflict of Interest
 Statement
- References



Comparte tu opinión con nosotros



https://axoncomunicacion.net/modelos-animales-enosteosarcoma/



Tres suplementos alimentarios que ayudan a mantener la función cardíaca

La adición de determinados suplementos aminoacídicos y componentes derivados de aminoácidos a la dieta de las mascotas puede ayudar al tratamiento de determinadas patologías. Además, la deficiencia de algunos de ellos puede ser la causa directa de enfermedades del corazón.



Taurina

La taurina es uno de los aminoácidos libres más abundantes. Se encuentra en elevadas concentraciones en los tejidos del músculo cardiaco, músculo esquelético, sistema nervioso central y plaquetas. Actúa en numerosos procesos metabólicos, ejerciendo diversas funciones:

- Antioxidación
- Actividad en las células fotorreceptoras de la retina
- Estabilización de las membranas neuronales
- Desarrollo del sistema nervioso
- Reducción de la agregación plaquetaria
- Reproducción

- Actividad miocárdica¹:
 - Modulación de las concentraciones de calcio en los tejidos y su disponibilidad.
 - Inactivación de los radicales libres y cambio de la osmolaridad celular.
 - Efectos en la osmorregulación del miocardio.
 - Otros mecanismos específicamente relacionados con la función miocárdica incluyen la N-metilación de los fosfolípidos de la membrana celular, efectos directos en las proteínas contráctiles e interacciones con el sistema renina-angiotensina-aldosterona.

¿Qué ocurre en casos de deficiencia de taurina?

En **gatos**, la taurina es un aminoácido esencial y su deficiencia puede causar miocardiopatía dilatada (MCD), degeneración de la retina y anomalías reproductivas. Existen evidencias de que la MCD causada por su deficiencia puede ser reversible con la suplementación de este aminoácido^{2,3}.

En **perros**, hasta hace unos años, la taurina no se consideraba un aminoácido esencial ni se conocía su papel en el desarrollo de la MCD⁴. Sin





embargo, diversos estudios han demostrado que sí lo es en perros alimentados con dietas restrictivas en proteína y que, al igual que los gatos, pueden desarrollar MCD secundaria a la deficiencia de taurina⁵.

L-carnitina

La L-carnitina es un derivado aminoacídico que se obtiene de la proteína de la dieta o por síntesis endógena en el hígado, siendo la lisina y la metionina los aminoácidos precursores. La síntesis requiere hierro, vitamina C y vitamina B6 como cofactores. El músculo esquelético y el cardiaco son los lugares donde se almacena hasta el 95-98% de la carnitina del cuerpo.

Entre las funciones de la carnitina, la más importante es la de cofactor de algunas enzimas necesarias para el transporte de ácidos grasos de cadena larga al interior de las mitocondrias, donde se oxidan para la generación de energía para el corazón, que obtiene de esta manera aproximadamente el 60% de su producción de energía total.

¿Qué es la miocardiopatía dilatada (MCD)?

Se trata de una enfermedad del corazón muy habitual, progresiva y, en gran medida, irreversible, que puede conducir a fallo cardiaco congestivo o muerte súbita. Es la segunda enfermedad cardiaca más habitual en perros, con una prevalencia superior al 50% en algunas razas10. La nutrición está actualmente aceptada como un importante adyuvante a la terapia médica en perros y gatos con MCD.

¿Qué ocurre en casos de deficiencia de L-carnitina?

La deficiencia de L-carnitina puede ser un trastorno primario o secundario.

- Las deficiencias primarias pueden aparecer por defectos genéticos en la síntesis, transporte, absorción o degradación. En personas se han asociado con cardiomiopatías.
- Las deficiencias secundarias son más comunes en pacientes que siguen dietas restrictivas

Se ha demostrado en perros que la deficiencia de L-carnitina puede favorecer el desarrollo de MCD en perros. Además, varios estudios^{6,7,8,9,10} han puesto de manifiesto que suplementar con carnitina mejora el tiempo de supervivencia de perros con MCD.

Hidrolizado de levadura de cerveza

La levadura de cerveza es un subproducto de la industria cervecera que puede ser un ingrediente beneficioso en la alimentación de las mascotas, ya que aporta el contenido nutricional que necesitan los perros y gatos¹¹. Esterilizada y sin poder leudante, es una levadura inactiva compuesta por el organismo unicelular Saccharomyces cerevisiae.

Su administración tiene efectos beneficiosos sobre la salud intestinal y la función inmune de los perros, estimulando las respuestas Th1 y, en consecuencia, la inflamación. Además, mejoran la palatabilidad de las dietas¹². Esto resulta especialmente útil para los perros con poco apetito a consecuencia de una enfermedad crónica.





Es una fuente proteínica rica en aminoácidos esenciales y vitaminas del grupo B:

- Los aminoácidos ayudan a la mascota a construir y mantener sus músculos, huesos, sangre, órganos, sistema inmunitario y pelaje y uñas. En particular, la arginina es un aminoácido esencial que reacciona con el oxígeno para producir óxido nítrico. El óxido nítrico relaja los músculos lisos de los vasos sanguíneos y reduce la presión arterial¹³. La hipertensión puede contribuir a las cardiopatías y a la insuficiencia cardíaca crónica, por lo que es conveniente controlar la tensión arterial de cualquier perro sospechoso de padecer una cardiopatía.
- Las vitaminas del grupo B contribuyen a la función cerebral, la fuerza muscular, la producción de glóbulos rojos y la digestión de los animales.

Por otro lado, se ha demostrado que estimula la producción de determinados marcadores de defensa antioxidantes, lo que ayuda a mejorar la salud cardiovascular de los animales¹⁴.

 A medida que progresa la insuficiencia cardiaca congestiva, aumenta el daño a las células cardiacas por la formación de radicales libres. Los estudios realizados en perros con insuficiencia cardíaca congestiva han demostrado que estos pacientes presentan un aumento de oxidantes reactivos y una disminución de antioxidantes a medida que progresa la enfermedad¹⁵.

En perros con fallo cardiaco, la oxigenación y el metabolismo celular no funcionan de forma apropiada, lo que conlleva la producción de elevadas cantidades de radicales libres. Los radicales libres son responsables de los principales daños celulares, lo que se denomina estrés oxidativo¹⁶.

Adicionalmente, la levadura de cerveza contiene sodio, calcio, magnesio y potasio. Muchos de los medicamentos utilizados para tratar las cardiopatías disminuyen los niveles sanguíneos de potasio y magnesio.

o Unos niveles inadecuados de potasio y magnesio pueden favorecer las arritmias cardiacas y debilitar las contracciones del músculo cardiaco¹⁷.

Bibliografía

- 1. Sanderson SL. Taurine and carnitine in canine cardiomyopathy. Vet Clin North Am Small Anim Pract. 2006 Nov;36(6):1325-43.
- 2. Pion,PD, Kittleson MD, Rogers QR & Morris JG. Myocardial failure in cats associated with low plasma taurine: A reversible cardiomyopathy. Science 237, 764–768 (1987).
- Pion PD, Kittleson MD, Thomas WP, Delellis LA & Rogers QR. Response of cats with dilated cardiomyopathy to taurine supplementation. J. Am. Vet. Med. Assoc. 201, 275–284 (1992).
- 4. Kramer GA, Kittleson MD, Fox PR et al. Plasma taurine concentration in normal dogs and dogs with heart disease. J Vet Intern Med 1995;9:253–8.
- Gray K. et al. The effect of 48-hour fasting on taurine status in healthy adult dogs.
 J. Anim. Physiol. Anim. Nutr. (Berl.) 100, 532–536 (2016). Sherry L. Sanderson, DVM, PhD, unpublished data, 1998).
- 6. Pion PD, Sanderson SL, Kittleson MD. The effectiveness of taurine and levocarnitine in dogs with heart disease. Vet Clin North Am Small Anim Pract 1998;28:1495–514.
- Sanderson S, Ogburn P, Osborne C. Heart disease management—Indications for nondrug therapies. Vet Forum 1996;13:36–43.





Doble protección para su corazón

El nuevo complemento alimentario en formato gel para perros y gatos que da soporte a BANACEP[®], nuestro fármaco indicado para el tratamiento de patologías cardíacas.



Compuesto por Benazepril









Compuesto por hidrolizado de levadura de cerveza, L-carnitina y Taurina

MOLÉCULA

ÚNICA









PALATABLE

FÁCIL ADMINISTRACIÓN









Cardiocep Gel

BANACEP[®] Vet 5 mg

BANACEP® Vet 20 mg

Hemeroteca

https://axoncomunicacion.net/ pequenos-animales-revolutionhemeroteca/







